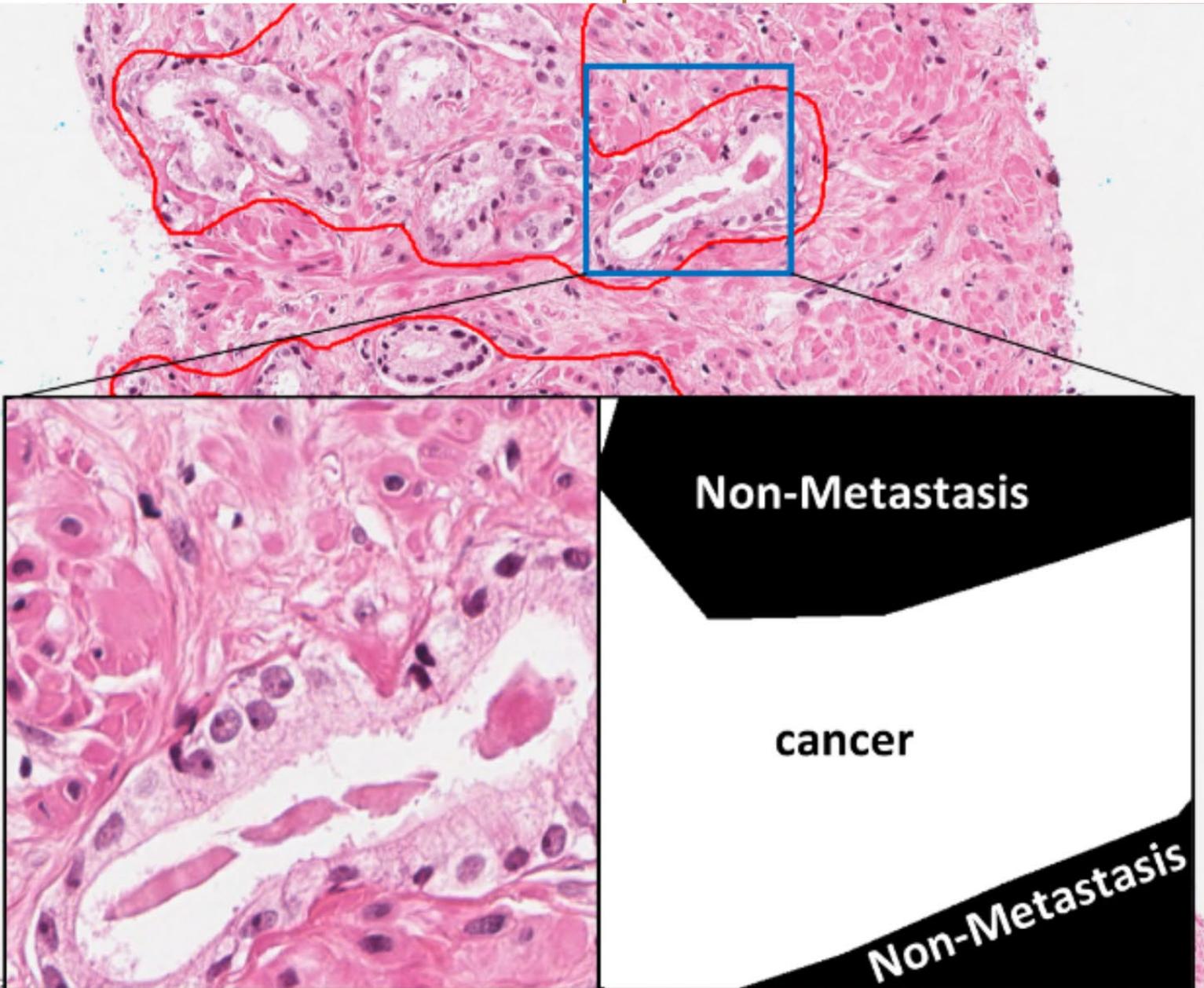


DEPARTMENT OF
PATHOLOGY
— & —
LABORATORY
MEDICINE

2022-2023
**Annual
Report**



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This publication has been produced by the Department of Pathology and Laboratory Medicine, a member of Alberta Precision Laboratories under Alberta Health Services and the Cumming School of Medicine, University of Calgary.

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Acknowledgments: Content Prepared by: Dr. Dylan Pillai, Dr. Tarek Bismar, and Ms. Katrina Epp.

Submissions from: Division/Section Heads Managers, Program Directors

We wish to thank all DPLM staff, administrators, and other team members whose efforts made this report possible.

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Territorial Land Acknowledgement

The University of Calgary, located in the heart of Southern Alberta, both acknowledges and pays tribute to the traditional territories of the peoples of Treaty 7, which include the Blackfoot Confederacy (comprised of the Siksika, the Piikani, and the Kainai First Nations), the Tsuut'ina First Nation, and the Stoney Nakoda (including Chiniki, Bearspaw, and Goodstoney First Nations).

The City of Calgary is also home to the Métis Nation of Alberta (Districts 5 and 6).

Department of Pathology and Laboratory Medicine Annual Report (2022-23)

Executive Summary



Photo credit: DPLM website

The Department of Pathology and Laboratory Medicine is now well past the 50-year mark and has seen many changes come and go in the model of clinical service delivery. Perhaps 2022-23 marked the most dramatic to and fro in service delivery seen in that 50 years. The two years saw the appearance of DynaLifeDx as the winning proponent of the request for proposals to provide laboratory services to the community sector in Calgary and indeed the rest of the province. The community versus acute care split was somewhat arbitrary and while it had performed well in the Edmonton zone, the onus was on the the private provider to demonstrate it could work provincially. In spite of a Herculean effort at a breakneck pace, DynaLifeDx struggled to meet the key performance indicators for laboratory service, with perhaps the most visible being the time to obtain an appointment at our patient service centres. As a result, the Department was informed in mid-2023 that DynaLifeDx would no longer retain the contract and all community services were to be re-integrated to Alberta Health Services (Alberta Precision Laboratories). On a more positive note, during these changes, a single laboratory information system was implemented for all of Alberta called “Connect Care”. This was a tremendous success and will hopefully enable big data science and clinical research at an unprecedented and integrated scale.

All this to say, there were consequences to the Department as the changes proved to be highly disruptive and drew resources and attention away from the core business of the Department: world-class clinical service, research and teaching. The transitions resulted in changes to research processes, teaching resources, and services rendered by pathologists, lab physicians and clinical scientists in a manner that challenged academic excellence. We have now emerged from this costly ‘flip-flop’ relatively unscathed with all residency programs intact and fellowship programs still functioning. Our academic staff continue to publish but at a lower rate in 2022-23 versus 2021 (perhaps as a consequence of the disruption) but still in leading international journals. We continue to garner extramural funding with considerable success from Tri-Council and other sources. Some of our leading lights have been recognized internationally for their efforts (eg. highly cited researchers by Clarivate) and contribute to guidelines that shape our future clinical practice. Discoveries continue to be translated into clinical practice. And our medical staff contribute to dozens of clinical trials by providing laboratory support. The future is challenged on the technical side with staff shortages and a pipeline that is struggling to replace that essential workforce, this being perhaps the biggest threat to our current service delivery model. Our equipment is aging, and

we are critically in a budget deficit that prevents us from ever-greening the equipment. In my estimation, a paradigm shift in service delivery is required through key innovations the department can help proffer. This will almost certainly include automation, digitization, and AI/machine learning. There is considerable work to do in this regard. We plan to recruit a GFT faculty member to lead this initiative. We also in collaboration with the Dept of Oncology and the Charbonneau Institute will launch the Translational Research Core (TRC) in the new Arthur Child Cancer Centre. The TRC will pioneer new diagnostic tests including digital imaging methods, molecular pathology, and mass spec techniques to support the Alberta ecosystem. The DPLM is sponsoring an AI initiative with digital images of prostate biopsies, biomarkers, and genomic data which holds promise to predict outcomes and impact patient management. Similar efforts are being pursued to create a translational core at the DSC in collaboration with the VPR office at U Calgary to support our clinical areas.

The Department leadership including the Deputy Head and Director of Education have continued to foster research and education excellence. The new triad model has proven effective in maintain DPLM core functions during all the changes in service delivery model. The annual research day was a smashing success with excellent abstracts presented followed by an awards ceremony. We also assembled at a GFT retreat where our academic faculty brainstormed on the future and a report will ensue in 2024 from this event. And finally, we were able to unwind at our Holiday Dinner in December 2022 and 2023 with a chance to reflect and perhaps give consideration to the more important things in life such as a wellness and our families.

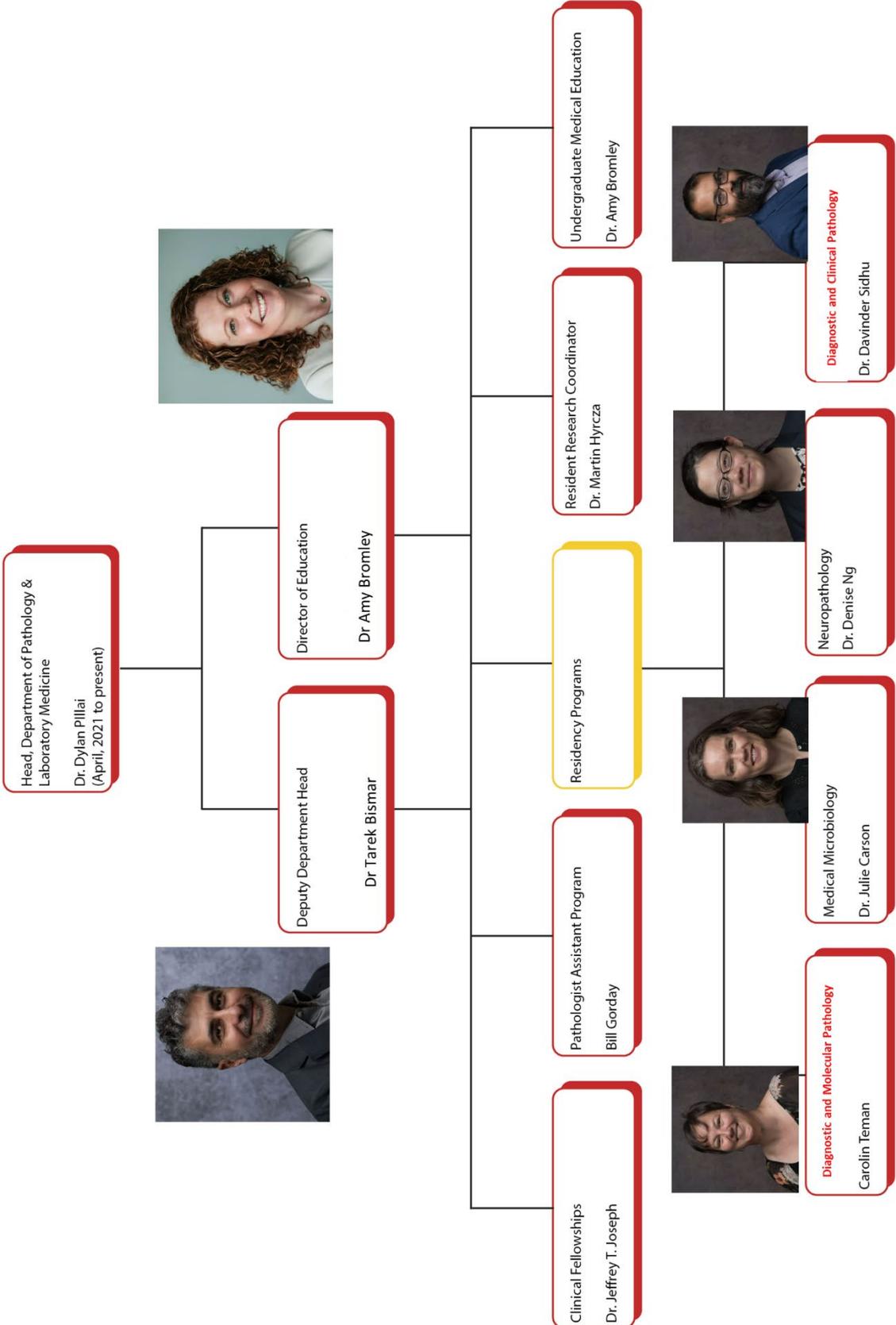
Lab will require a paradigm shift that requires new innovations in automation, digitization, and appropriateness using evidence all of which the DPLM is poised to provide as long as we protect the academic education and research activities. To not do so would be short-sighted and further undermine the laboratory mission.

Best wishes,

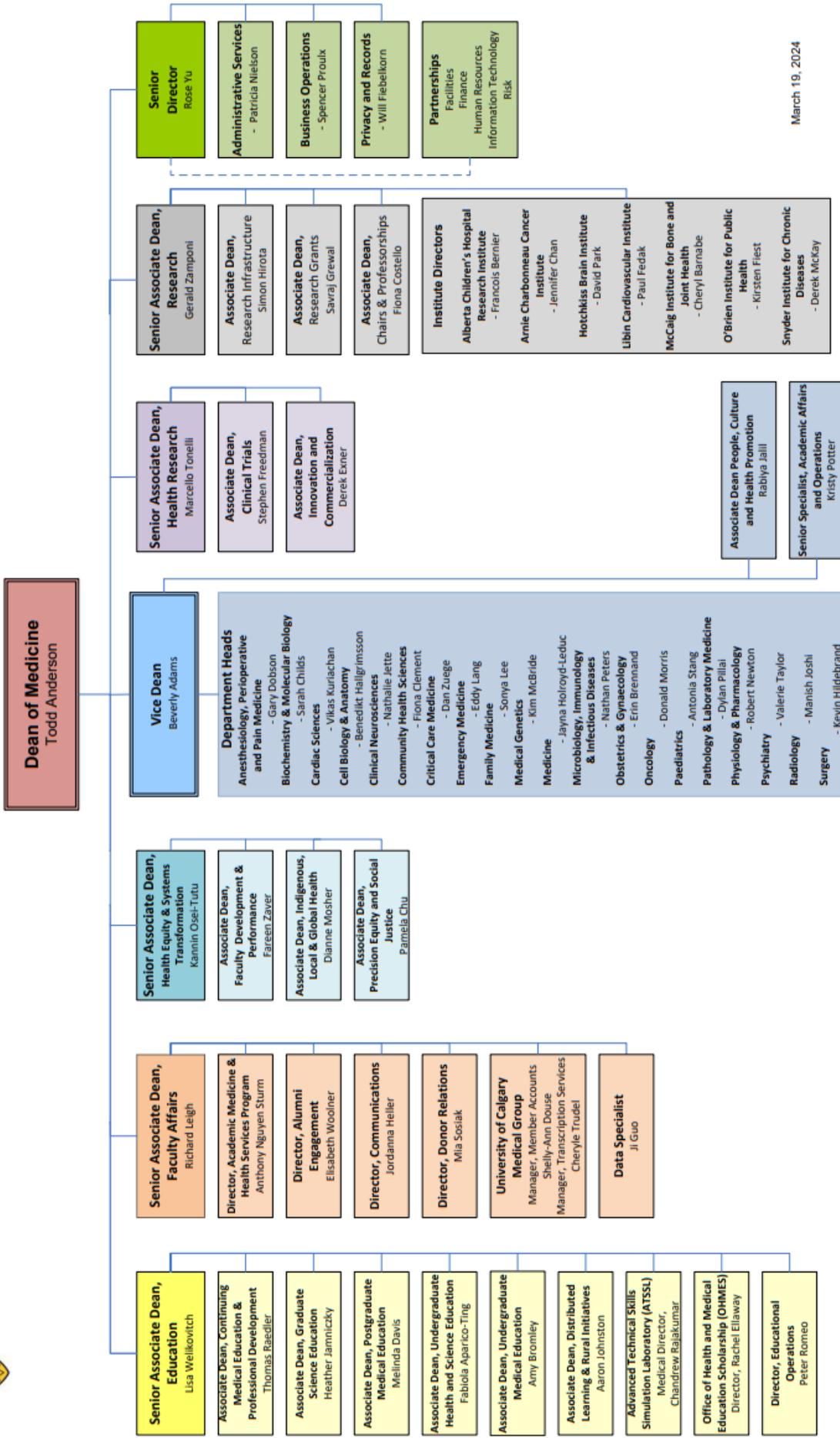
A handwritten signature in black ink, appearing to read 'Dylan Pillai', with a long horizontal flourish extending to the right.

Dylan Pillai MD, PhD. FRCP (C)
Professor & Head
Department of Pathology & Laboratory Medicine
University of Calgary

2024 DPLM Organizational Structure



2024 U of C Organizational Structure



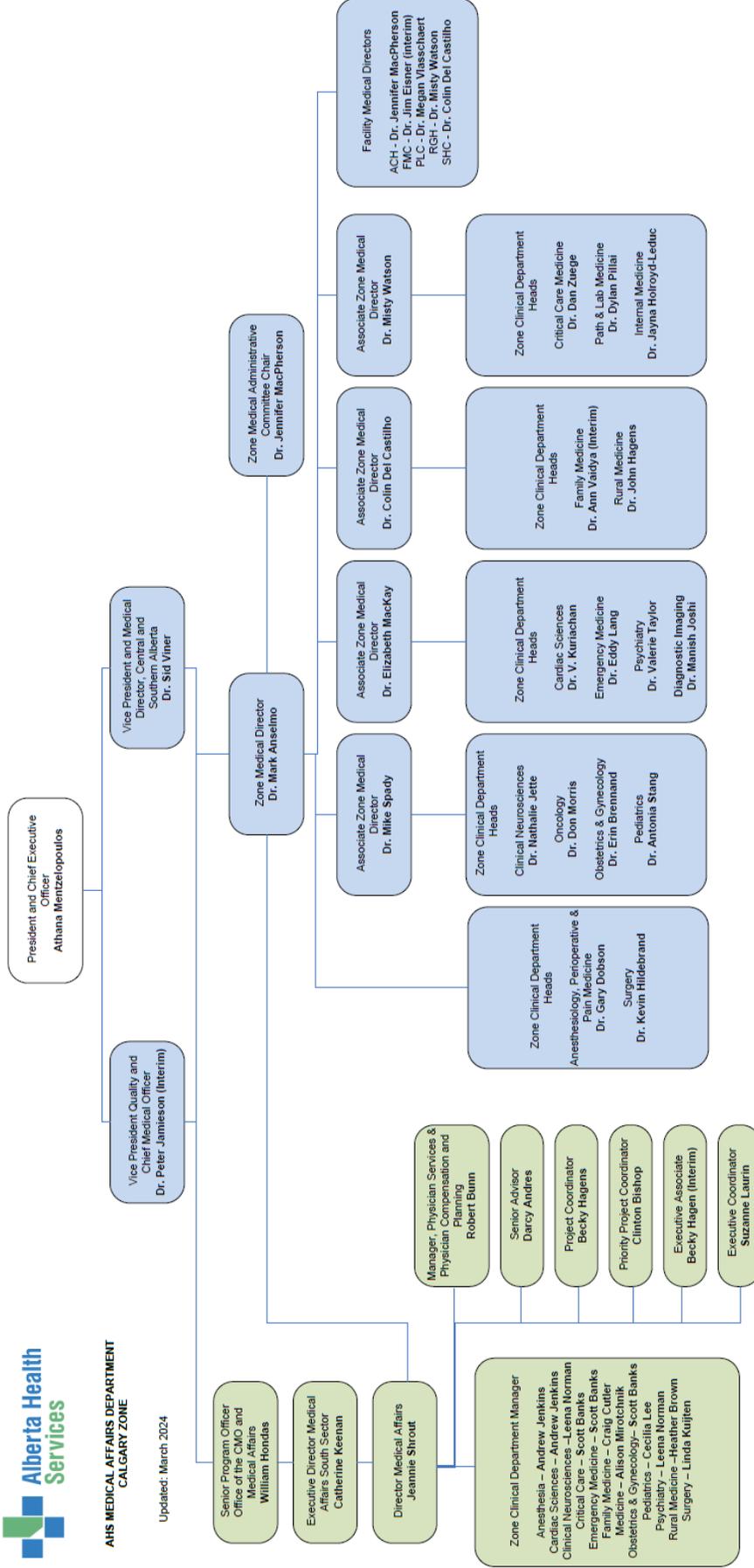
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2024 AHS Organizational Structure

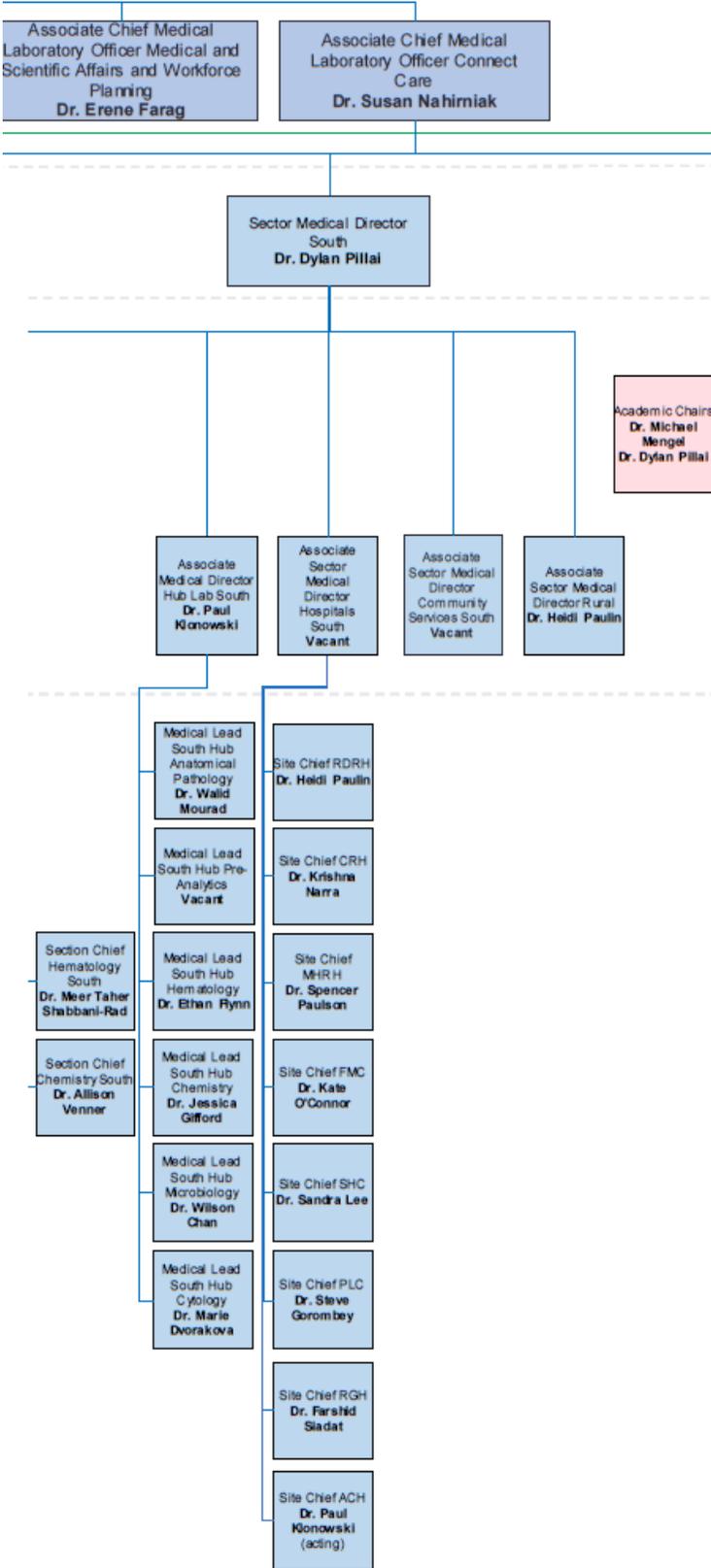


AHS MEDICAL AFFAIRS DEPARTMENT CALGARY ZONE

Updated: March 2024



2024 APL South Sector Organizational Structure



Clinical Section Updates



Photo credit: Totojang, Getty Images via Canva Pro

The Department of Pathology and Laboratory Medicine has stood out for its adaptability and commitment to excellence during the 2022-2023 period.

This portion of our annual report highlights our clinical advancements, how we've navigated through challenges, and the major changes that have shaped our department.

Clinical Section Updates

Diagnostic and Molecular Pathology (credit Drs Paul Klonowski/Ana Cota)

Major Changes

- Extensive planning and subsequent transition of staff, operations, equipment etc. to and then back again from the private provider (DynaLIFE).
- Extensive planning around and then execution of said plan to separate community specimens from acute care specimens, and to centralize some testing from regional sites to the DSC. General impact felt across most sites, most notably at DSC (community) and FMC (consolidation of STAT testing).
- Roll out of ConnectCare and Epic/Beaker, with the need to readjust some workflows, protocols, SOPs, etc. due to the slight differences in the nature of this LIS vs. the former. Extensive effort to reeducate staff, optimize workflow, and to find efficiencies, most of which is still ongoing.
- Major renovation and reconfiguration of space within DSC (performed by private provider), with the expansion of lab space in the histology/cytology areas, addition of a second gross room, second processor room, additional space for microtomy and embedding stations, new special stains area and file room/consult desk area, new shared accession space with ability to cross-train and pull resources to greatest workload demand in AP or Cytology. Entire cytology department moved into newly renovated space at the DSC.
- PLC lab moved to new space with addition of much needed lab and office space.

New Technology, Equipment, Innovations

- Connect Care implementation (May 2023 for most urban sites)
- Replacement of 2 Dako IHC stainers (1 “borrowed” from Lethbridge, 1 from Dako), plus the addition of 1 net new instrument (also Dako), to maintain the constantly increasing pressures on IHC lab.
- Addition (thanks to private provider) of 6 new tissue processors, 1 new embedding station, 4 new grossing stations, new IPC (cassette printer), and 5 new HE600 stainers.
- New Hologic Thin prep processors, imagers, and review scopes (cytology DSC)
- Additional grossing station(s) at PLC.
- Several new IHC antibodies approved and ready to validate or are making their way through the process.

Challenges

- Increased workload with inadequate staffing resources/workforce shortages exacerbated by transitions/integrations, both on the operational side as well as on the medical side, the latter resulting in excess overflow work. We have been unable to fill cytology technologist vacancies due to lack of qualified applicants. An aging work force with seemingly insufficient graduates/qualified candidates to fill looming retirements.
- Ongoing shortages of space within most lab operations, including office spaces for pathologists and PhDs. Some gains at the DSC (thanks to private provider) and PLC, but some areas, such as the DSC IHC lab, RGH, ACH, continue to be very cramped. FMC to see some space gains with transition of some staff to the new Cancer Centre (late 2024).
- Ongoing cuts/attrition and general lack of adequate staffing levels of clerical, admin, and other support staff resulting in daily challenges at every level of the organization, from

manning phones (such as at PLC and ACH with only part time staff available) to sorting slides, triaging consult/send out cases, to helping with minuting of meetings.

- Ongoing intermittent issues with staining quality, water quality, supply chain issues (delays in shipments of consumables and reagents, the changes in formulations of IHC aliquots, etc.)
- Ongoing issues with logistics, exacerbated by transition to private provider, with delays in delivery of slides to pathologists, some missed/delayed deliveries of specimens between sites.
- High volumes of sample deficiencies, leaking containers, missing clinical histories/demographics/etc. requiring mitigation and follow-up.
- Aging equipment, including microscopes, with insufficient replacement.
- Lack of dedicated equipment and space to process neuropath specimens, as well as IHC stain for prion protein, exacerbated by CJDSS severing its contract with Federal agencies, has made neuropathology/autopsy practice challenging, with extreme delays in processing of neuropath samples. A QAR resulting from contamination of 2 IHC processors as well as the other challenges forced us to explore other options – possibly solution/implementation still an ongoing effort.
- Increased testing volumes across the board, including additions of new IHC stains, with difficulty of keeping up with increased requests/workload due to staffing levels and delays in validating already approved stains. Ongoing difficulty in obtaining control tissues. Delays in implementation of on-slide controls due to lack of capital equipment funds (slide printer).

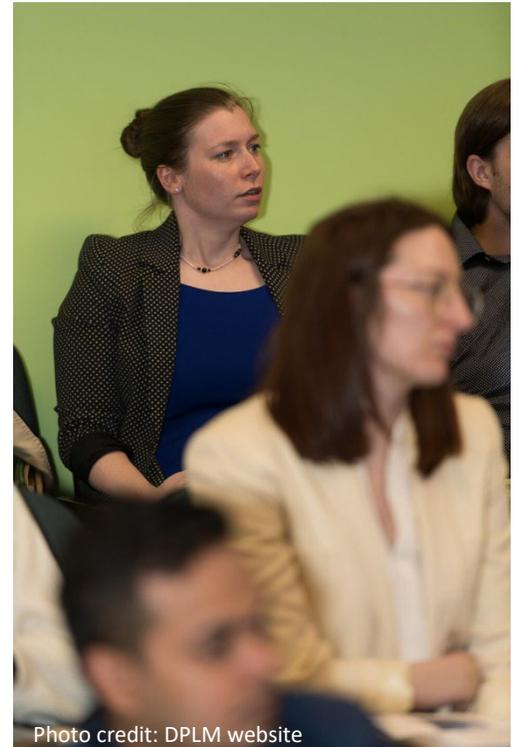


Photo credit: DPLM website

Diagnostic and Clinical Pathology (credit Dr Ethan Flynn)

Major Changes to Clinical Service & Testing

- Implementation of D-dimer testing for Outpatient collections in Calgary Zone on December 5, 2022 (repatriation from Calgary Zone RRL labs)
- Repatriation of consult to Pathologist cases for Routine CBC/diff performed at South Zone Rural test sites but warranting referral to Pathologist, on December 5, 2022 (formerly, smear to Pathologist consults on these South Zone Rural lab cases were performed by Medicine Hat and Lethbridge-based Pathologists)
- Repatriation of Hematology testing from South Zone testing sites to DSC Hub Lab on February 27, 2023 for South Zone outpatient collections through Patient Service Centres in Lethbridge, Brooks and Medicine Hat; as well as Routine lab tests collected at Rural Healthcare facilities in South Zone that could not be performed at that test site.
- PT/INR, PTT, D-dimer testing on South Zone collections - repatriated back to South Zone hospital lab sites on November 29, 2023.

New Technology, Equipment, Innovations

- DSC Hub Lab Hematology migration from Millennium LIS to Beaker LIS within Epic/Connect Care system on May 6, 2023
- DSC Hub Lab Hematology migration in Sysmex analytic middleware from WAM to Caresphere on May 6, 2023

Challenges

- DSC Hub Lab Hematology underwent significant challenges due to changes in administrative reporting structure, human resources and executive oversight during the 2022-2023 period:
 - Jan 1, 2022 – Dec 4, 2022: APL
 - Dec 5, 2022 – August 31, 2023: DynaLIFE
 - September 1, 2023 – Dec 31, 2023: APL
- Largest challenges occurred with assumption of testing of South Zone routine Hematology collections from South Zone in late 2022. DSC Hub Lab absorbed testing from Medicine Hat and Lethbridge in Fall 2022 (prior to Dec 5, 2022) due to staffing shortages at those lab sites.
 - Staffing shortages on technical/operational side
 - Difficulties and challenges encountered with South Zone Hematology labs operating in the Meditech LIS environment, while DSC Hub Lab Hematology lab operating in (1) Millennium LIS (Feb 27, 2023 until DSC Hub Lab Connect Care go-live on May 6, 2023) or in (2) Connect Care/Epic LIS (May 6, 2023 until South Zone Connect Care/Epic go-live on November 5, 2023).
 - Numerous specimen stability issues and resultant specimen cancellation impacts with specimens collected at South Zone sites but not arriving to DSC Hub Lab Hematology until many hours later (“timed-out”; multiple factors including Preanalytics and Logistics challenges on their end, however problem was worse before May 6, 2023 Connect Care go-live since South Zone PSCs were data entering into Meditech, and then DSC Hub Lab had to re-data enter into a different LIS, Millennium, upon arrival at DSC). Issues with missing samples and timed-out samples from South Zone, primarily with INR testing, has been alleviated since South Zone samples for coagulation testing were re-routed back to South Zone hospital testing labs on November 29, 2023.
- Challenge of smooth transition from Millennium to Epic/Connect Care LIS; compounding the challenge was the concurrent transition from WAM to Caresphere middleware within DSC Hub Lab Hematology: technical and medical training, validation, despite MLT staffing shortages.



Photo credit: Akram Huseyn, Unsplash

Medical Microbiology (credit Dr Wilson Chan/Byron Berenger)

Major Changes to Clinical Service & Testing

- Major changes in governance: transfer of department to DynaLIFE, then reintegration into Alberta Precision Laboratories, has led to major shifts not only of ownership, but in all the ancillary support structures (e.g., IT, HR, Ordering, Quality) and policies on which laboratory performance is based.
- Transfer of testing from rural/regional South Zone community sites has allowed us to consolidate testing, but also resulted in a ~20% increase in workload.
- Implementation of ConnectCare has changed the way the laboratory functions: many benefits, but also has exacted costs in efficiency and utilization.

New Technology, Equipment, Innovations

-
- Lab automation equipment (KiestrA) system has been partially acquired and implemented. Currently used for urine cultures, but with full deployment, application for more specimen types, and additional improvements slated for the future.
- Implementation of a LAMP assay for Group A Streptococcus detection in April 2022: improvements in performance characteristics and efficiency over previous method.

Challenges

- Workforce shortages, like everywhere else in health care, continue to be problematic, and have been exacerbated by the pace and magnitude of changes in the last two years.
- Gaining a measure of operational stability will allow for gains to be made in performance.
- Optimization of ConnectCare for the laboratory continues to be a work in progress.

Clinical Biochemistry (credit Dr Allison Venner/ Jessica Gifford)

Provincial

Major Changes to Clinical Service & Testing

- Insulin-Like Growth Factor-1 (IGF1) moved onto mass spectrometry for testing
- Hemoglobin A1c testing for Canadian Longitudinal Study on Aging (CLSA) testing was moved to Lethbridge, and included dried blood spot analysis validation
- DynaLIFE transition led to workload changes – some sites significant increase (e.g. FMC, SHC, DSC) and other sites significant decrease (e.g. regional site hospitals)
- New tests implemented at FMC: Cholesterol, Cholesterol (Fluid), Ferritin, Free Triiodothyronine (FT3), Free Thyroxine (FT4), HDL Cholesterol, LDL Cholesterol Calc/Non-HDL-C Calc, Thyroid Stimulating Hormone (TSH), Protein (Total Urine & Protein/Crea Ratio Urine), Total Prostate Specific Antigen, Carcinoembryonic antigen, Bile Acids, Iron, UIBC, TIBC/% sat calc, Chylomicron screen, Chylomicron screen (Fluid)
- Additional testing began at FMC for: APL Research, Community neonatal bilirubin, Community NT-proBNP testing, STAT testing from HCTL after hours
- PLC: Brand new lab space on 6th floor of the East tower June 2023 and successfully completed CPSA accreditation
- PLC: 2 Roche Cobas Pro analyzer lines, additional GEM 5000 blood gas analyzer, Osmopro and A2O osmometers

New Technology, Equipment, Innovations

- Implementation of Epic/Beaker (November 2022, May 2023, November 2023)
- ACH: 2x new Roche Cobas Pro c503 automated chemistry analyzers (includes updates to Methotrexate and Primidone assays), 2x new Medica Pro Type I water systems, Roche AST/ALT Gen 2 reagents were implemented
- ACH: Funding from the Alberta Children's Hospital Foundation for a Roche Cobas Pure e402 immunoassay analyzer to run high-sensitivity cardiac troponin T, Beta-hCG and NT-proBNP on-site. To be implemented in 2024
- FMC: 2 new Roche Cobas C8000 automated chemistry analyzer lines
- Analytical Tox: Working with KeyProteo, a company based in Seattle, assisted with validation of a novel newborn screening test
- Analytical Tox: Reviewing drugs of abuse testing panel
- MHRH: 2x new Roche Cobas Pro c503 automated chemistry analyzers
- CRH: 2x new Roche Cobas Pro c503 automated chemistry analyzers
- Rural sites in Central, Calgary and South: Implementation (or early of plans for) of new Vitros XT-3400s at several sites (continues into 2024)
- Development of a Provincial 2-hr Beckman Access High-sensitivity Troponin I algorithm
- Oilfields and Strathmore: New reagent fridges



Challenges

- Older equipment (e.g. water systems, mass spectrometry analyzers)
- Lack of viable backup analyzers (e.g. osmometers)
- Administrative support for effective RRL function
- Sites with increased workloads have been challenged to manage these increases alongside staffing challenges and new workflows
- Staffing challenges within MLA, MLT, operations and medical laboratory teams across sites/departments
- Staff change fatigue, burnout, and continuous request to do more with less resources
- APL/DynaLIFE transition and integration has had an enormous impact on test menu, routing and courier management/understanding, TATs, staffing levels
- Numerous changes with Connect Care changes required resources to manage, align, troubleshoot workflows and reporting
- Supply chain issues continue to be a problem that has resulted in additional calibrations and troubleshooting.

Calgary Community

Major Changes to Clinical Service & Testing

- In this period, we provided clinical service under two different lab entities:
 - APL: Jan 1-Dec 4, 2022
 - DynaLIFE: Dec 5, 2022-Aug 31, 2023
 - APL: Sept 1-Dec 31, 2023
- DynaLIFE transition on Dec 5, 2022 led to the go live of a number of new tests in DSC Chemistry and associated new instrumentation
 - Instrumentation to perform the Celiac Disease Screen testing
 - Instrumentation to measure serum free light chains
 - Instrumentation to perform hemoglobin electrophoresis and the sign out of results adding a new clinical service to the laboratory staff and medical/scientific staff working in DSC Chemistry: Sign out by Calgary team started Oct 1, 2023. This service was covered by Edmonton staff from Dec 5, 2022 to Oct 1, 2023.
 - Instrumentation to perform serum and urine osmolality testing
 - Mono testing which is a manual process
- DynaLIFE transition on Dec 5, 2022 resulted in DSC Chemistry performing testing for community STAT orders. Prior to this transition, this testing was performed by FMC and the RRLS.

New Technology, Equipment, Innovations

- January 2022 Roche Infinity middleware implementation
- January-October 2022 implementation of 4 new Roche Diagnostics cobas 8000 lines
- Dec 2022 implementation of the following instrumentation associated with the DynaLIFE transition
 - BioRad Bioplex (1)- Celiac testing
 - Binding Site Optilites (2)- serum free light chains
 - Sebia Capillary Flex II- transferred from Edmonton BaseLab when they received new instrumentation (2)- serum protein electrophoresis and hemoglobinopathy investigations
 - Advanced Instruments OsmoPro (1)- serum and urine osmolality testing
- EPIC ConnectCare LIS went live May 6, 2023
- Associated with the ConnectCare implementation, numerous provincial reference interval and reporting harmonization innovations went live

Challenges

- Transition of LIS from Millennium to Connect Care led to the loss of site-specific functionalities. Now, alternatives need to be explored or rebuilt into ConnectCare.
 - Eg. worklist formats, outstanding lists, expected lists
- Routing changes as well as logistic challenges affecting TAT
 - Delayed TATs resulting in the cancellation of time-sensitive analytes eg. potassium, glucose collected in South Zone
 - Delayed critical result reporting from specimen collected in South Zone due to transport time
- Aging equipment (pre-analytcs units, electrophoresis equipment, urinalysis equipment). A lot of downtimes either needing manual intervention or affecting TATs.
- Capacity issues

- 100 % on pre-analytics
- 97 % on chemistry modules on the Roche cobas lines
- Protein electrophoresis volume increases
- Lab staffing challenges including burn out. Not enough technologists to manage the increase in workload.

Neuropathology (credit Dr. Jeff Joseph)

Major Changes to Clinical Service & Testing

- Operationalize new molecular diagnostic platforms that relate to neoplastic neuropathology. These include how to incorporate new information into our synoptic reports and our grading of tumours. Major changes were made in Calgary in May-June 2023 that greatly impacted our diagnoses of neoplastic neuropathology.
- Use of antibodies for analysis of inflammatory muscle diseases. Antibodies are the cornerstone for the diagnosis of inflammatory muscle diseases and over these two years have standardized our use of several antibodies that allow us to make more refined and accurate diagnoses. These include: MxA, MCH-1, MCH-2, membrane attack complex, and p62. We have requested that these antibodies be available in Calgary and are working to either get these approved or validate them.
- Streamline CJD testing and handling of tissue. This has been a multi-year endeavour and is not yet complete.
- Approval of new antibodies for neoplastic testing (see below).
- Increasing complexity of autopsy neuropathology, including neurodegenerative and medical brain diseases. The 2022-2023 was the busiest autopsy neuropathology period in Calgary since 2007.



Photo credit: Drew Hays, Unsplash

New Technology, Equipment, Innovations

- Approval of several diagnostic antibodies (MxA, olig2, H3K27-Me3, BRAF-V600E, several for pediatric embryonal tumours)
- Working with the Molecular group, use of their new DNA Next-Generation sequencing and RNA translocation equipment. Quality control all cases that undergo molecular testing.
- In this timeframe, ConnectCare became active and required substantial reworking of how we complete cases and garner information.

Challenges

- Rapidly changing aspects of clinical work. Most notably, tumour molecular diagnostics but also the more detailed aspects of diagnosing muscle diseases, neurodegenerative diseases, and some changes in the diagnoses of epilepsy pathology. We have three staff members who focus on brain tumours from both diagnostic and research sides and update us on new diagnostic entities or grading. The information required in most of our reports is substantially greater than even a few years ago (most tumours, details of muscle diseases, details of neurodegenerative diseases). Ten - fifteen years ago, the majority of our muscle

biopsies were for various types of dystrophies and acquired myopathies; today, the majority of our biopsies are for inflammatory diseases and only a few are now to confirm a previous genetic test.

- Difficulty in having neuropathology staying "current" or providing the ever-changing standard of care on our cases.
- Resource difficulties with severely limited funding for new testing and minimal resources for innovation.
- Teaching demands in this era of changing expectations and increasing sub-specialization. This includes training of neuropathology residents, who now need to know a substantial amount of information that was not even imagined ten years ago or known five years ago.

Hematopathology (credit Dr. Meer-Taher Shabani-Rad)

Major Changes to Clinical Service & Testing

- Hempth group has contributed to the academic performance of the department by publishing multiple articles in peer-reviewed journals during 2023:
 - Published articles by hempth staff; First or last Author: 8 articles
 - Contribution to publications by another groups: 8 articles
 - Dr Adnan Mansoor has had another productive year with publication of 7 articles
 - The article published by Dr Afshin Shameli/Shabani-Rad was recognized as the most cited article of 2022 in the field of Flow cytometry
- Hematopathology staff have supported implementation and validation of Connect Care Launch 6, with physical presence in CZ hospitals since 4:30 AM and supervising and engagement in the validation process of CC for the transition of lab operation
- New platform for Hemoglobinopathy sign-out has been created and integrated to Connect Care
- Hempth group has continued to support APL's Molecular Pathology operation by significant engagement in providing clinical services in the field of molecular hematology
- Successfully secured two Clinical Fellowship positions for 2024 by significant support from DPLM



Photo credit: DPLM website

New Technology, Equipment, Innovations

- Planning for validation of Hempth Labs (flow cytometry and special Hematology labs) in Arthur Child New Cancer Center
- New State of art flow cytometry equipment has been delivered for Flow lab at ACCC

Cytopathology (credit Dr. Marie Dvorakova)

Major Changes to Clinical Service & Testing

- Provincial Pilot study for Self-sampling helping underserved/underscreened populations in AB. The Pilot study has just entered the laboratory validation phase and is expected to start

accepting patient samples likely in the Summer/Fall of 2024. Edmonton Prov Lab will be conducting the hrHPV testing for the Pilot.

- Preliminary discussions about primary HPV screening for cervical cancer are underway with Public Health/ACCSP. Expected phased implementation starting in ~3 years.

New Technology, Equipment, Innovations

- Calgary APL (DSC) was the Canadian testing site for the Hologic Genius Digital System using AI to recognize abnormal cells in cytologic preparations – the system for Gyne Cytology has now been granted Health Canada approval. The system allows for faster screening and slide organization/filing. Could allow sharing scanned slides between the Zones to better utilize Cytotechs for screening without the need to ship glass slides.

Challenges

- Staffing levels on the operational side, with expected retirements within the next 2-3 years. Cytotechs currently working OT to keep up with TAT/KPIs. Currently about 4 FTE short.



Education

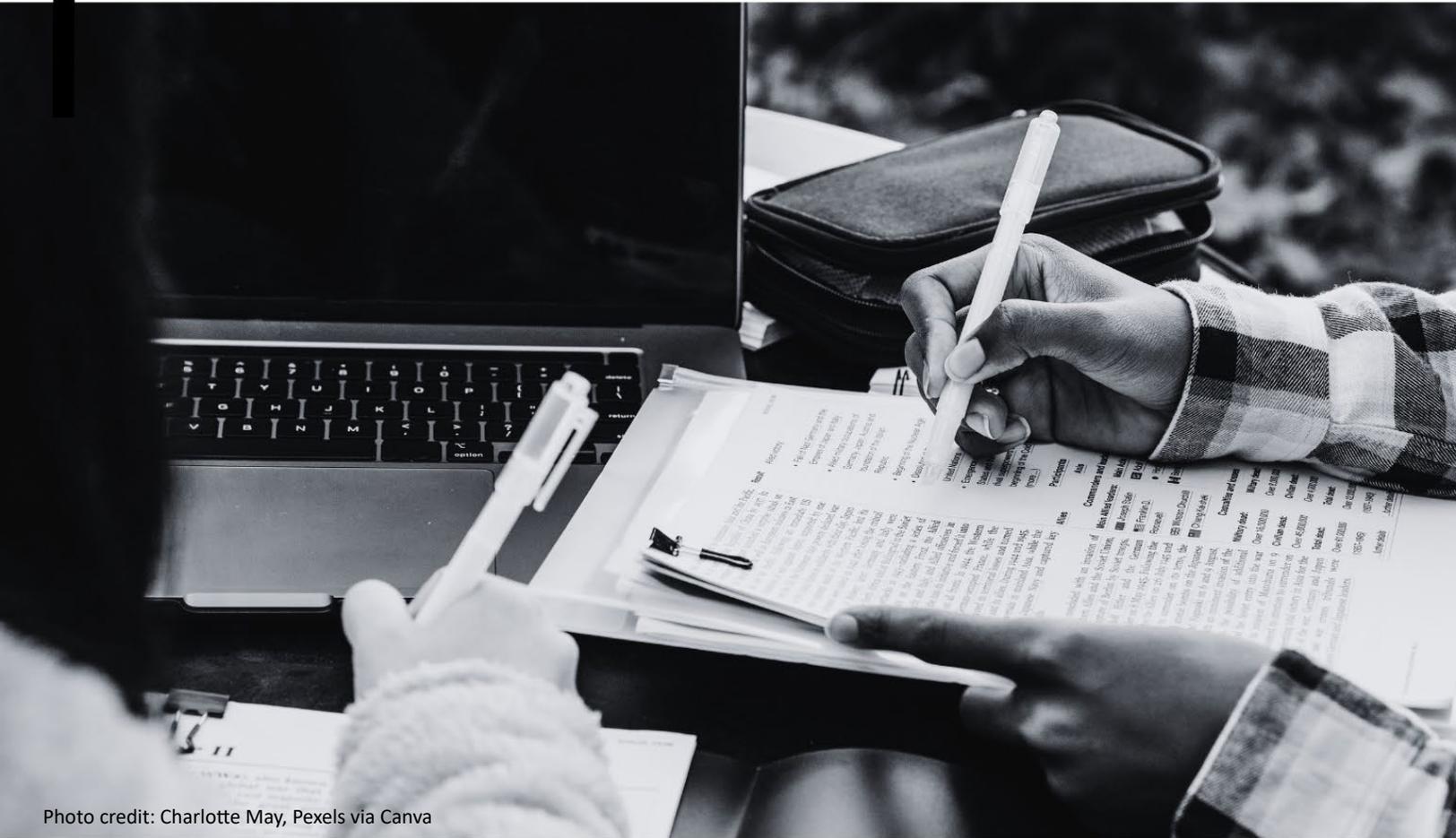


Photo credit: Charlotte May, Pexels via Canva

During the 2022-2023 period, the Department of Pathology and Laboratory Medicine focused on delivering robust educational programs and advancing research within the field. Our commitment to quality teaching and evaluation has been central to supporting the academic and professional development of our trainees.

Education

Diagnostic and Molecular Pathology Program (credit Dr. Carolin Teman/Konstantin Koro)

The Diagnostic and Molecular Pathology Program is a five-year program leading to certification in Diagnostic and Molecular Pathology by the Royal College of Physicians and Surgeons of Canada. In 2023, our specialty name was officially changed by the Royal College from Anatomical Pathology to Diagnostic and Molecular Pathology. Our program transitioned to Competency by Design (CBD) in July 2019, and is now almost exclusively comprised of residents in the CBD stream. The curriculum includes orientation, clinical rotations, core adult surgical and autopsy pathology rotations, subspecialty rotations, elective rotations, research, and 3-4 months of chief resident service. The curriculum is designed to provide graduated responsibility to residents. In the CBD stream, residents write the practical component of their Royal College Examination in the fourth year of training. In the final year, during Transition to Practice, residents are expected to perform at the level of a fellow or junior faculty member, recognizing that faculty-resident supervision is always occurring. In 2023, we welcomed three new residents through the CaRMS match. We currently have 18 residents in our program. Administrative support and training facilities are provided by the University of Calgary's Department of Pathology and Laboratory Medicine.

Teaching

A philosophy of supported self-directed learning underlies the program. Teaching takes place via a combination of dedicated educational events, group learning and one-on-one teaching. Structured educational events include departmental Continuing Medical Education rounds, resident-led Gross Pathology rounds, optional evening subspecialty slide sessions, and a dedicated weekly academic half day consisting of unknown slide rounds, autopsy rounds, and didactic teaching. Most of our educational activities are now offered virtually via Zoom, which has made it easier for trainees to attend even when they are rotating off-site. Some sessions are also now offered in hybrid format. Residents are also expected to read and study independently.



Evaluation

CBD residents are assessed based on their achievement of Entrustable Professional Activities (EPAs) as they progress through stages of training. Several rotations incorporate end-of-rotation slide exams or presentations into their assessments. PGY1-PGY4 residents are also assessed via biannual in-house examinations, including a full RCPSC-style examination each winter and spring, and the American Society of Clinical Pathology Resident In-Service Exam (RISE) each spring. The program director meets with each resident at least twice yearly to discuss the resident's academic progress, research projects, subspecialty interests, and fellowship/career plans.

Research

Involvement in research activities is an integral part of the program. Beginning in the PGY2 year, residents undertake one or more research projects with the advice and mentorship of the Resident Research Advisor. Research activities are generously supported through the DPLM's Trainee Led Project Support Initiative (TLPSI). Residents present their research findings at the annual departmental research day, as well as at national and international meetings.

Resident Progress & News:

Our program graduated two residents in 2023, who went on to complete subspecialty fellowships (Breast Pathology at University of Alberta and Forensic Pathology in Minneapolis). Our current senior residents will complete fellowships in GI Pathology (Memorial Sloan-Kettering Cancer Center), dermatopathology (University of Calgary), and Gynecologic Pathology (University of Calgary) next year. The University of Calgary's Diagnostic and Molecular Pathology residency program is well-regarded nationally and receives a large number of applicants for the annual CaRMS match. In 2023 and 2024 we filled all CaRMS positions with outstanding applicants.

Program Accreditation & Upcoming Changes:

The Diagnostic and Molecular Pathology residency program participated in the Royal College accreditation review in 2022 and received accreditation with follow-up by external review. Several areas for improvement were identified and are being actively addressed. Internal review by PGME is scheduled for summer 2024. Upcoming changes include the implementation of a redesigned longitudinal didactic curriculum that has undergone major organizational changes under the leadership of Dr. Ashely Flaman. The anticipated implementation date is mid-2024. Our program's greatest challenges include continued implementation and changes due to CBD, including faculty development with respect to utilization of EPAs.

Medical Microbiology Program (credit Dr. Julie Carson)

The Medical Microbiology residency training program at the University of Calgary is a full Royal College accredited five-year program that aims to train medical microbiologists to be competent and confident practitioners.

The program's training is focused at developing expertise and skills in the four major spheres of medical microbiology as outlined by the Royal College of Physicians and Surgeons of Canada: the medical, scientific, administrative direction and management of a clinical diagnostic laboratory; the provision of clinical consultation in infectious diseases; infection control and antimicrobial stewardship; and public health.



Resident Progress & News:

In 2022-2023 academic year, the Medical Microbiology program had three residents – PGY2, PGY3 and PGY5 level of training.

Program Accreditation & Upcoming Changes:

The Medical Microbiology residency program was fully accredited by the Royal College of Physicians and Surgeons of Canada in September 2022 with no major deficiencies.

The specialty of Medical Microbiology will implement Competency by Design in July 2024. Pre-launch activities and planning are underway.

Training Overview

The PGY-1 year provides an experience akin to the rotating internship, with rotations in a variety of related clinical disciplines to supplement the clinical knowledge and skillset of the trainee.

PGY years 2 through 5 involve a mix of rotations to meet the specialty training requirements. Diagnostic laboratory rotations (up to 22 four-week blocks) focus on bacteriology, virology, mycology, parasitology, and public health microbiology. Clinical infectious diseases rotations (13 four-week blocks), include both adult and pediatric, inpatient and outpatient services. The infection control and antimicrobial stewardship blocks (3 four-week blocks) are shared with the Infectious



Photo credit: DPLM website

Disease programs. There is a significant amount of elective time (up to 13 four-week blocks) included to allow trainees to further develop in subspecialties and research of their choosing.

The Medical Microbiology Program has three major laboratory training sites. The Diagnostic Scientific Center is the primary site for bacteriology, mycology, parasitology, and molecular training and where residents have the majority of their core laboratory training. The Public Health Lab – South (Calgary) location is the primary training site for virology, serology, molecular and public health lab microbiology. Our program

has an inter-university agreement with University of Alberta whereby our residents have one mandatory rotation at the Public Health Lab – North (Edmonton)/University of Alberta Hospital Laboratory to meet their mycobacteriology, containment level three and bacterial typing objectives.

Our residents also have opportunities in their Community Microbiology block to engage with our Microbiologists and technical teams (in person and remotely) in the regional centers microbiology laboratories – Red Deer Regional Hospital, Chinook Regional Hospital (Lethbridge) and Medicine Hat Regional Hospital.

Our clinical sites for training in Infectious Disease include all the major hospitals in Calgary: Foothills Medical Center, Peter Lougheed Center, Rockyview General Hospital, South Health Campus Hospital and the Alberta Children’s Hospital as well as the Sheldon M. Chumir Health Centre for several public health related outpatient clinics.

Our program also supports the training of Adult and Pediatric Infectious Disease Residents (12 weeks) and General Pathology Residents (24 weeks) and electives for these programs. We collaborate closely with our Infectious Disease programs with respect to shared curricula in Infection Control and Stewardship as well as our academic half day content.

Research

Medical Microbiology residents are expected to complete at least one research project during their residency. Our residents have been involved in several research projects, contributing to publications and presentations at local, national, and international meetings in the last year.

Medical Microbiology Residency Program Committee 2022-23

Name	Major Site Affiliation	Major Responsibility with the Program
Carson, Julie	DSC	Program Director, DSC Site Lead
McCartney, Nathan	n/a	Chief Resident
Gregson, Dan	DSC	Research/Academic Coordinator
Groeschel, Michael	DSC and Regional	Exam Chair
Jadavji, Taj	ACH	Pediatric Infectious Diseases representative
Kim, Joseph	RGH	Infection Control or Stewardship representative
Lisboa, Luiz	DSC and Regional	Competency by Design & Competency Committee Chair
Pitman, Shawna	DSC	Program Administrator
Vaughan, Stephen	SHC	Adult Infectious Diseases representative
Zhou, Hong Yuan	Public Health Lab	Public Health Lab Microbiology representative
Bromley, Amy	FMC	Academic Department Education Chair (ex-officio)
Lenz, Ryan	RGH	PGY1 RGH program director (corresponding member)
Pillai, Dylan	DSC	Academic Department Head (ex-officio)
Sidhu, Davinder	DSC/FMC	DGP program director (corresponding member)

The Neuropathology Program (credit Dr. Denise Ng)

One of four training programs in the country, our five-year program aims to train successful future neuropathologists while achieving qualifications for certification in Neuropathology by the Royal College of Physicians and Surgeons of Canada. The educational experiences are provided through a partnership between the University of Calgary and Alberta Precision Labs with added collaboration with the Ministry of Justice through the Office of the Chief Medical Examiner. The Neuropathology Residency Training Program in Calgary boasts extensive exposure to a high volume of neurosurgical and autopsy specimens from both adult and pediatric populations. Our training site is fortunate to have one of the most robust hospital autopsy services in the country and a strong collaboration with the Alberta Children's Hospital. Additionally, the site benefits from strong relationships built with other laboratory medicine colleagues, including but not exclusive to anatomical pathologists, molecular pathologists, forensic pathologists, medical microbiologists, and clinical biochemists. We have also fostered excellent relationships with clinicians, showcasing frequent collaboration as

demonstrated by frequent clinical discussions and multiple interdisciplinary rounds where pathology is presented and discussed.

Competency by Design:

Neuropathology as a specialty had officially transitioned in 2022 to the new Royal College mandated Competency By Design (CBD) curriculum structure. Although extensive planning has been underway in Calgary, initiatives have not been implemented as a new resident has not been matched since the transition.

In the transition, the program committee anticipates some changes to the structure of training. Formal phases of Transition to Discipline and Transition to Practice stages have been introduced. Although less formalized forms of these training phases been part of our traditional-stream program, these new educational stages have now been formalized with increased structure, clear objectives of training, and formal evaluation. Clinical rotations have been more stringently tailored to the neuropathology training needs, allowing for earlier focus on the Foundation and Core of training. The certification examinations have also been brought forward approximately six months to balance Foundation and Core training needs with exposure to Transition to Practice.



In future, the neuropathology program aims to include a multidisciplinary Competency Committee to review resident evaluation and feedback, as a subcommittee of the Neuropathology Residency Training Committee. We hope to also incorporate participation of neuropathologists from other sites, potentially expanding the expertise made available to our residents.

The program anticipates future modification to the curriculum will be a continuous improvement process, with feedback and support from our highly involved neuropathology staff and residency training leadership, including collaboration with medical leadership and other residency training programs in the Pathology and Laboratory Medicine department.

Current Program

Our program includes two very active traditional stream residents. In the last year, another successful resident completed training and was successful at the Neuropathology certification exams. In the last decade, every successfully graduated neuropathology resident that has applied to the Royal College Examination have successfully passed the Neuropathology certification exams by the Royal College of Physicians and Surgeons of Canada.

On-site Royal College accreditation survey/review of the University of Calgary's Neuropathology Program took place on September 20, 2022. The program was given accreditation with requirement for Action Plan Outcomes Report next year for areas of focused improvement. As part of a concerted effort for continuous improvement, the training program hopes to implement structural and process changes in the committee to address these previous challenges.

Members	Major Site Affiliation	Major Function with the Program
Dr. Denise Ng	Foothills Medical Centre	Program Director
Dr. Dylan Pillai	Diagnostic and Scientific Centre	Department Head
Dr. Amy Bromley	Diagnostic and Scientific Centre	General Pathology and Associate Dean, Undergraduate Medical Education
Dr. Jeffrey T. Joseph	Foothills Medical Centre	Neuropathology Division South Sector Lead, Committee member
Dr. Jennifer Chan	Arnie Charbonneau Cancer Institute	Institute Director of the Arnie Charbonneau Cancer Institute, Research advisor, Committee member
Dr. Kristopher Langdon	Foothills Medical Centre	Committee member
Dr. Ana Nikolic	Foothills Medical Centre	Research advisor, Committee member
Dr. Omid Rashidipour	Foothills Medical Centre	Committee member
Dr. Akmal Coetzee-Khan	Office of the Chief Medical Examiner (OCME)	Coordination of OCME site training
Dr. Marie-Anne Brundler	Alberta Children's Hospital (ACH)	Coordination of ACH site training
Chief Resident Chris Newell	Foothills Medical Centre	Resident Representative

Diagnostic and Clinical Pathology Program (credit Dr. Dave Sidhu)

Our program is a five-year program leading to certification in anatomic and clinical Pathology by the Royal College of Physicians and Surgeons of Canada. The University of Calgary through co-sponsorship with Alberta Precision Labs offer Diagnostic and Clinical Pathology Residency Training highlighting on laboratory management and pathology informatics. The Diagnostic and Clinical Pathology Residency Program includes approximately 4 plus years of laboratory medicine and multiple clinical training experiences. The clinical training is optimized to provide exposure to most of the medical and surgical services that rely heavily on the clinical and anatomical pathology laboratory. Upon successful completion of the education program, the residents will be competent to function as consultants in anatomical and clinical pathology and medical laboratory directors. In 2023 the residency program name was officially changed from General Pathology to Diagnostic and Clinical Pathology by the Royal College.

Competency by Design:

Since 2019 the Diagnostic and Clinical Pathology (DCP) program has continued to implement the new Royal College mandated Competency By Design (CBD) initiative for General Pathology evaluation and feedback. A Competency Committee, a subcommittee of the Residency Training Committee, oversees resident evaluation, promotion, and curriculum development within the CBD curriculum. A total of 9 residents are currently enrolled in the DCP Pathology CBD curriculum and have successfully completed multiple stages of training,



with all residents on track with expected progression through the curriculum. In addition to the CBD residents, we have a single legacy traditional stream resident continuing to progress through those training channels.

One important change within the CBD curriculum is that the Royal College certifying examination was moved from PGY5 to PGY4 to allow a period of transition to practice. This academic year saw our first cohort of DCP Pathology CBD residents participate in the Royal College certifying exam. Modification to the curriculum continues to be adjusted and improved with collaboration from residency training leadership in medical microbiology, hematopathology, anatomical pathology, and clinical chemistry to ensure core curriculum material is covered prior to the examination. The basic clinical rotations have been consolidated and modified to include disciplines more directly applicable to general pathology practice and will allow residents additional time within laboratory medicine training. We have also introduced new training opportunities including procedure simulation sessions with the Cumming School of Medicine Advanced Technical Skills Simulation Laboratory.

Current Program

Our program has successfully graduated every DCP pathology resident that has applied to the Royal College Examination, all of whom have successfully passed the General Pathology certification exams by the Royal College of Physicians and Surgeons of Canada.

Successful on-site Royal College accreditation survey/review of the University of Calgary’s General Pathology Residency Training Program took place on September 20, 2022, with no major deficiencies noted. The program was given accreditation with regular review in 8 years. We currently have 9 residents in the program and continue to deal with and overcome the current challenges of the ongoing laboratory services transition and postgraduate program funding as they relate to effective and safe resident education.

Members	Major Site Affiliation	Major Function with the program
Dr. Davinder Sidhu (Chair)	Foothills Medical Centre	Program Director
Dr. Dylan Pillai	Diagnostic and Scientific Centre	Department Head
Dr. Amy Bromley	Diagnostic and Scientific Centre	General Pathology and Associate Dean, Undergraduate Medical Education
TBA	Foothills Medical Centre	Director of Education.
Dr. Marie Dvorakova	Diagnostic and Scientific Centre	Coordination of DSC site training
Dr. Julie Carson	Diagnostic and Scientific Centre	Coordination of Medical Microbiology training
Dr. Tariq Roshan	Foothills Medical Centre	Coordination of Hematological Pathology training
Dr. Konstantin Koro Dr. Simon Walker	Foothills Medical Centre	Coordination of Anatomic Pathology training
Dr. Jessica Gifford	Diagnostic and Scientific Centre	Coordination of Medical Biochemistry training
Dr. Robby Wang	Red Deer Hospital	Coordination Red Deer Hospital training
Dr. Ryan Lenz Chief Resident Junior Resident	Rockyview General Hospital	Corresponding member PGY-1 site representative Rotating (5-6 months) Rotating (5-6 months)

Medical Scientific Staff Departures & Arrivals

Arrivals

Name	First	Effective Date	Site	Specialty
Vuong	Jennifer	10-Jan-22	FMC	Anatomic Pathology
Heaton	Sarah	07-Feb-22	DIASCI	Anatomic Pathology
Todorovic	Emilija	11-Apr-22	FMC	Anatomic Pathology
Al-Shamma	Zainab	01-Jul-22	FMC	Anatomic Pathology
Liao	Chiu-Hsiang	02-Aug-22	RGH	Anatomic Pathology
Wang	Linyuan	02-Aug-22	FMC	Anatomic Pathology
Walker	Simon	08-Aug-22	FMC	Anatomic Pathology
Flaman	Ashley	06-Sep-22	DIASCI	Anatomic Pathology
Tordon	Bryan	01-Mar-23	FMC	Transfusion Medicine
Seno	Rommel	01-Jul-23	FMC	Hematopathology
Cavazzi	Elena	14-Aug-23	FMC	Anatomic Pathology
Fung	Bonita	14-Aug-23	ACH	Pediatric Pathology
Tzafiriri	Ori	01-Sep-23	PLC	Anatomic Pathology
Rashidipour	Omid	01-Oct-23	FMC	Neuropathology/Molecular

Departures

Name	First	Effective Date	Site	Specialty
Thommasen	Amy	15-Apr-2022	DSC	General Pathology
Falck	Vince	30-Jun-2022	FMC	Anatomic Pathology
Paslawski	Doreen	30-Jun-2022	RGH	Anatomic Pathology
Wright	James (Jim)	30-Jun-2022	ACH	Pediatric Pathology
Mirza	Imran	30-Nov-2022	FMC	Molecular Pathology
Terzic	Tatjana	2-Dec-2022	DSC	Anatomic Pathology
Shameli	Afshin	31-Mar-2023	FMC	Hematopathology
Kurek	Kyle	30-Jun-2023	ACH	Pediatric Pathology
Liao	Chiu Hsiang (Connie)	31-Aug-2023	RGH	Anatomic Pathology
Brenn	Thomas	30-Nov-2023	DSC	Anatomic Pathology
Baskin	Leland	31-Dec-2023	DSC	General Pathology
Cota Schwarz	Ana	31-Mar-2024	FMC	Anatomic Pathology

Department of Pathology & Laboratory Medicine Chair's Awards

2022

Education Awards

Dr. Etienne Mahe

Dr. Julie Carson

Research Awards

Dr. Byron Berenger (Clinical Faculty)

Dr. Tarek Bismar (GFT Faculty)

Clinical Service Awards

Dr. Marie Dvorakova (Anatomic Pathology)

Dr. Angela Franko (Anatomic Pathology)

Dr. Michael Groeschel (Medical Microbiology)

Dr. Charlene Hunter (Anatomic Pathology)

Dr. Etienne Mahe (Hematopathology)

Dr. Shaun Medicott (Anatomic Pathology)

Dr. Denise Ng (Neuropathology)

Dr. Travis Ogilvie (Anatomic Pathology)

Dr. Heidi Paulin (General Pathology)

Dr. Hossein Sadrzadeh (Clinical Chemistry) [Special Long Service Award]

Dr. Anna Sienko (Anatomic Pathology)

Dr. Kiril Trpkov (Anatomic Pathology)

Dr. Ranjit Waghray (Anatomic Pathology) [Special Long Service Award]

Dr. Emma Whitcomb (Anatomic Pathology)



Photo credit: DPLM website

2023

Research

Dr. Margaret Kelly (GFT Faculty)

Dr. Ramin Zargham (Clinical Faculty)

Clinical Service Awards

Dr. Xiu Jian (Hematopathology)

Dr. Tom Griener (Medical Microbiology)

Dr. Amy Bromley (Anatomic Pathology)

Dr. Allison Venner (Clinical Biochemistry)

Dr. Leland Baskin (General Pathology)

Education Awards

Anna Thomas: Resident Appreciation Award

Dr. Mike Harvey: SS Collegiality Award

Dr. Kyo Farrington: Outstanding Teaching by a Resident

Drs. Zainab Al-Shamma & Amy Bromley Faculty Teacher of the Year



Photo credit: DPLM website



Research



Photo credit: Khunkorn, Canva

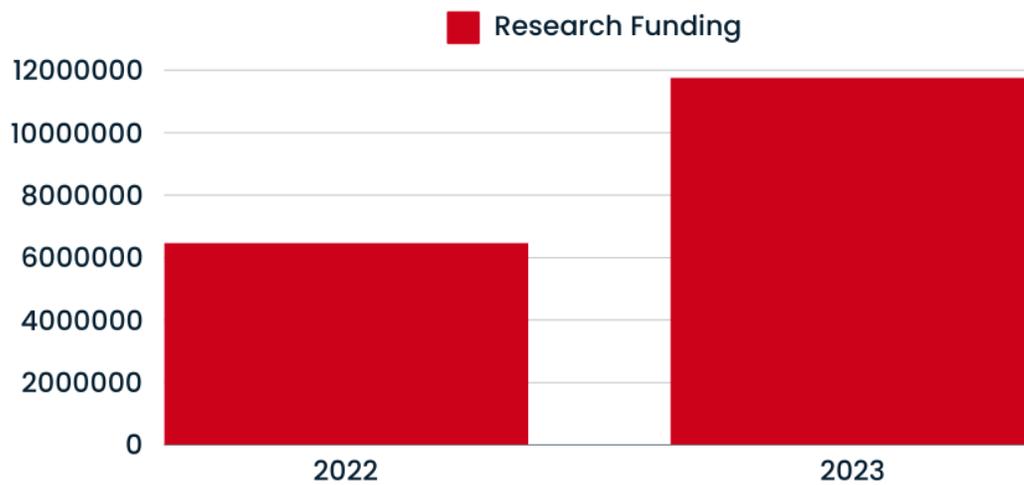
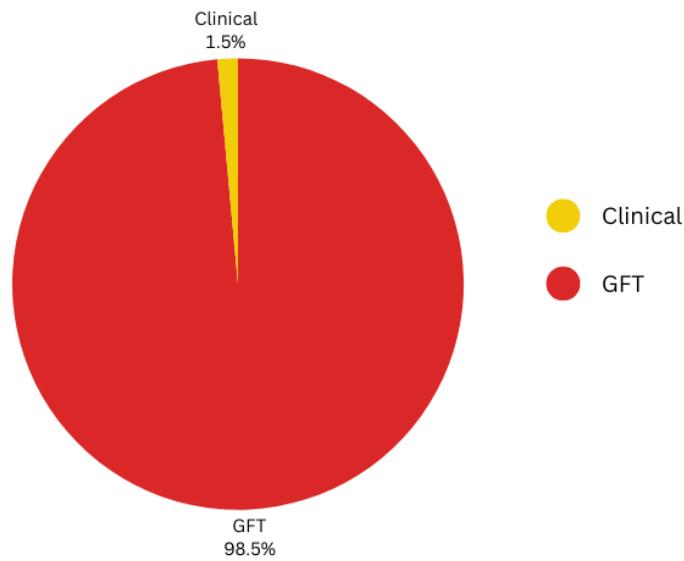
This section provides a comprehensive overview dedicated to the lifeblood of innovation and progress within the Department of Pathology and Laboratory Medicine. In 2022-2023, our department has been the fortunate recipient of numerous grants that have not only fueled our research endeavors but have also significantly enhanced our diagnostic and therapeutic capabilities. These funding sources, ranging from federal agencies to private foundations, underscore the confidence and belief in our mission to advance the field of pathology and laboratory medicine.

Research

2022-23 Funding by Appointment Type

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x x x x x
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x x x x x x
x x x x x x
x x x x x x



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RESEARCH FUNDING /YEAR

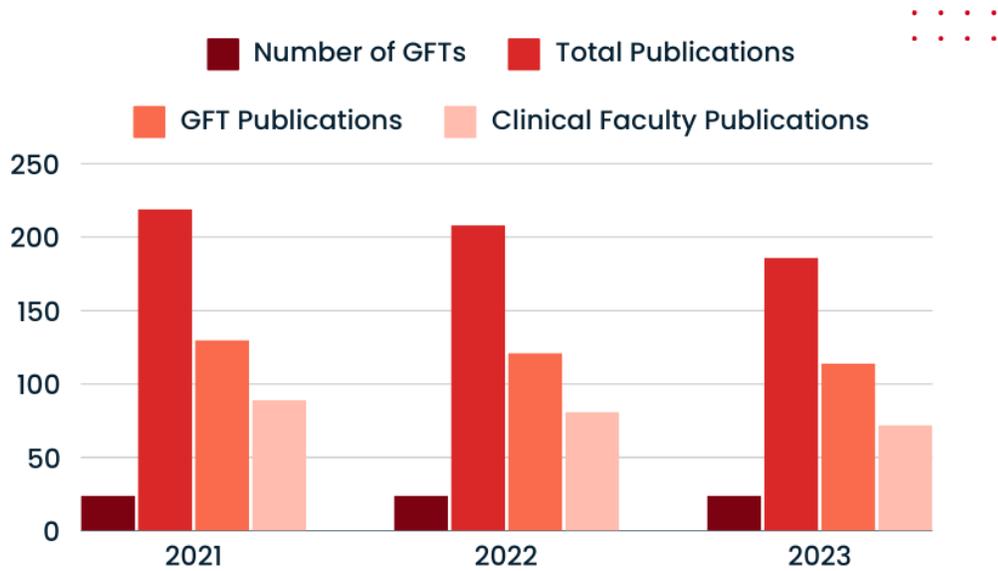
Principal Investigator/Co-Investigators	Year	Funding Source	Total Awarded
Berenger, Byron (Clinical)			
Rapid RNA sequencing of coronavirus for Public Health surveillance and transmission	2022	University of Alberta	\$19,000
Bismar, Tarek (GFT)			
Characterization of Novel Molecular Signature for Accurately Predicting Prostate Cancer Progression in Active Surveillance	2022-2023	Canadian Cancer Society	\$168,479
GU Oncology Fellowship	2023	Janssen Inc.	\$80,000
IOF for CFI # 14571	2023	Canada Foundation for Innovation	\$1,094
Mutational Landscape of Unclassified Renal Cell Carcinoma	2023	Alberta Health Services / Alberta Precision Laboratories	\$10,000
Box, Adrian (clinical)			
"Development of molecular testing for lung cancer biomarker (ROS) in Alberta Precision Laboratories"	2022	Calgary Health Foundation	\$12,000
Bromley, Amy (GFT)			
Internal Audit of the Hospital Autopsy Service/ Impact of the SARS-CoV-2 Pandemic on Mortality as Encountered in a Hospital Based Autopsy Service - Dr. Nicole House	2022	Alberta Health Services / Alberta Precision Laboratories	\$2,000
Chan, Jennifer/Charbonneau Institute (GFT)			
ACF Chair Funds for Pediatric Cancer	2022-2023	Alberta Cancer Foundation	\$100,000
ACTION - Calgary Foundation - Five Year Commitment.	2023	Calgary Foundation	\$150,000
ACTION Mechanisms to Medicines	2022-2023	Canadian Cancer Society	\$1,071,150
Alberta Cellular Therapy and Immune Oncology Initiative (ACTION)	2022	Multiple Sources	\$5,000
Arnie Charbonneau Cancer Institute Greatest Needs Fund	2022-2023	Multiple Sources	\$148,251
Brain Tumour Research within the Arnie Charbonneau Cancer Institute	2022-2023	Multiple Sources	\$30,925
Clark H Smith Integrative Brain Tumor Research Centre	2022-2023	Smith, Tony and Jane	\$30,648
Cancer Outcomes Research Program - Oncology Outcomes (O2) Program	2023	Multiple Sources	\$205,000
Cancer Prevention and Risk Reduction Research	2023	Multiple Sources	\$250,000

Cancer and Genetic Research Fund	2022	Multiple Sources	\$3,575
Charbonneau Biobank Donations	2023	Multiple Sources	\$16,225
Childhood Cancer Research Program - 1	2022	Alberta Children's Hospital Foundation	\$140,000
Donations to SACRI for supporting Cancer Research	2022	Multiple Sources	\$49,397
Drivers of Oligodendrocyte Precursor Cell Dysfunction in the Origin and Maintenance of Oligodendroglioma	2022	Canadian Institutes of Health Research	\$244,322
Emerging Cancer Scholars Program	2023	Multiple Sources	\$591,150
Impact in Glioblastoma (Brain Cancer) Research	2023	Multiple Sources	\$100
Kids Cancer Care Award in Childhood Cancer Research	2022	Kids Cancer Care Foundation of Alberta	\$100,000
Lisa Shea Trainee Travel Award	2023	Shea, Lisa	\$5,000
Lung Cancer Early Testing and Diagnostics	2023	Multiple Sources	\$200,000
Lung Cancer Research within the Arnie Charbonneau Cancer Institute	2022	Multiple Sources	\$3,525
MOHCCN Prairies Cancer Consortium (PR2C) - U of Calgary. MOH	2023	Terry Fox Research Institute	\$2,655,100
Marathon of Hope Cancer Centres Network (MOHCCN)	2023	Multiple Sources	\$247,922
Marathon of Hope Prairies Cancer Research Consortium (PR2C)	2022-2023	Terry Fox Research Institute	\$1,514,025
Mechanisms of epigenetic evolution in pediatric high-grade gliomas	2022-2023	Canadian Institutes of Health Research	\$258,060
Molecular Oncology and Cancer Immunotherapy Research Laboratories at the Calgary Cancer Centre	2023	Multiple Sources	\$1,035,238
New Recruitment in Translational Hematology	2023	Alberta Cancer Foundation	\$870,000
Oncology Outcomes O2 Infrastructure	2022-2023	Multiple Sources	\$57,702
PROFYLE Model Systems Activities in Calgary - Partnered with Terry Fox Research Institute (TFRI)	2022	Alberta Cancer Foundation	\$81,690
Precision Oncology for Young People: Biobanking and database development and coordination (PROFYLE)	2022	Terry Fox Research Institute	\$25,000
Recurrent 3D genome features in PFA ependymoma	2023	Canadian Institutes of Health Research	\$390,000
Rejeanne Taylor Research Prize in Precision Cancer Medicine	2022	Taylor, Robert W.	\$55,000
Richardson Oncology Surgery Fund	2022	Richardson, Dale	\$5,000

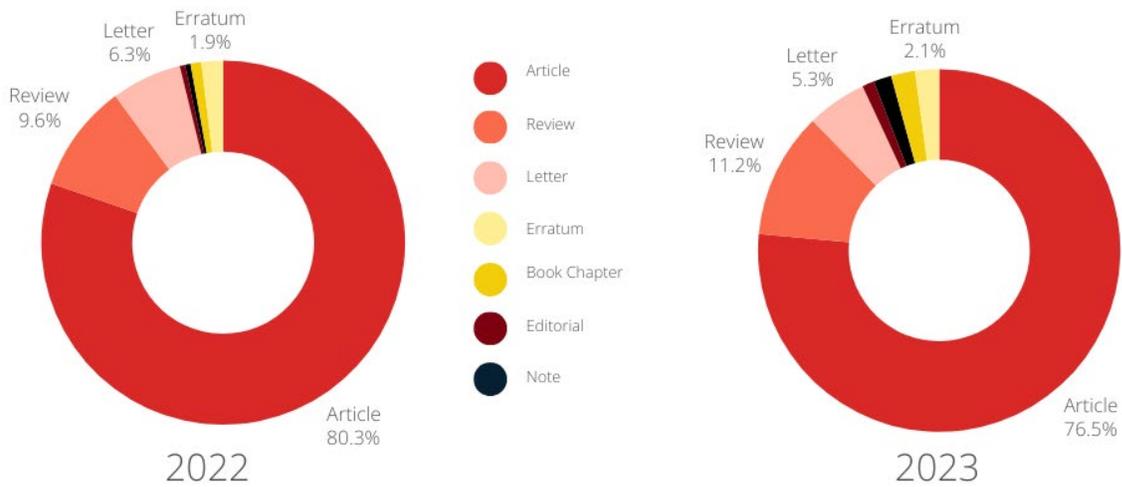
Robson DNA Science Centre	2022-2023	Calgary Foundation	\$1,423,148
Rod Chisholm Memorial Fund for Glioblastoma Research	2023	Multiple Sources	\$111,270
Southern Alberta Cancer Research Institute (SACRI) Cancer Genomics Program	2022-2023	Anonymous Donor	\$2,508,987
The Cancer Data Platform	2022	Antje Graupe Pryor Foundation	\$500,000
The Prairie Cancer Research Consortium (PCRC) - Pilot Project	2022-2023	Terry Fox Research Institute	\$309,375
The evolution of transcriptional cellular state dynamics in recurrent glioblastoma	2023	Terry Fox Research Institute	\$45,000
Training Tomorrow's Leaders in Hematology	2023	Alberta Cancer Foundation	\$100,000
Hycza, Martin (GFT)			
Head and Neck Tumour Banking Program	2022	Calgary Foundation	\$25,000
Trainee-Led Projects Support Initiative (TLPSI)	2023	Alberta Health Services / Alberta Precision Laboratories	\$145,000
Kelly, Margaret (GFT)			
Multiplexed Ion Beam Imaging (MIBI) Technology	2023	Alberta Children's Hospital Foundation	\$400,000
Khan, Faisal (GFT)			
5+14=0: A new maths based on KIR genes to reduce Graft versus host disease after allogeneic HCT	2022	Buckley Family Foundation	\$1,000
Early Detection of Acute Myeloid Leukemia Relapse after Hematopoietic Cell Transplantation	2022-2023	Alberta Cancer Foundation	\$371,500
Koebel, Martin (GFT)			
A Population Based Study of Ketorolac and Ovarian Cancer Survival	2023	University of Colorado Denver	\$14,038
A Population Based Study of Ketorolac and Ovarian Cancer Survival	2022	University of New Mexico	\$6,873
Mansoor, Adnan (GFT)			
Proteomic validation of Transducin beta-like 1 X-linked receptor 1 (TBL1XR1) gene expression in aggressive B-cell lymphomas arising at immune privileged sites	2023	Alberta Health Services / Alberta Precision Laboratories	\$4,946

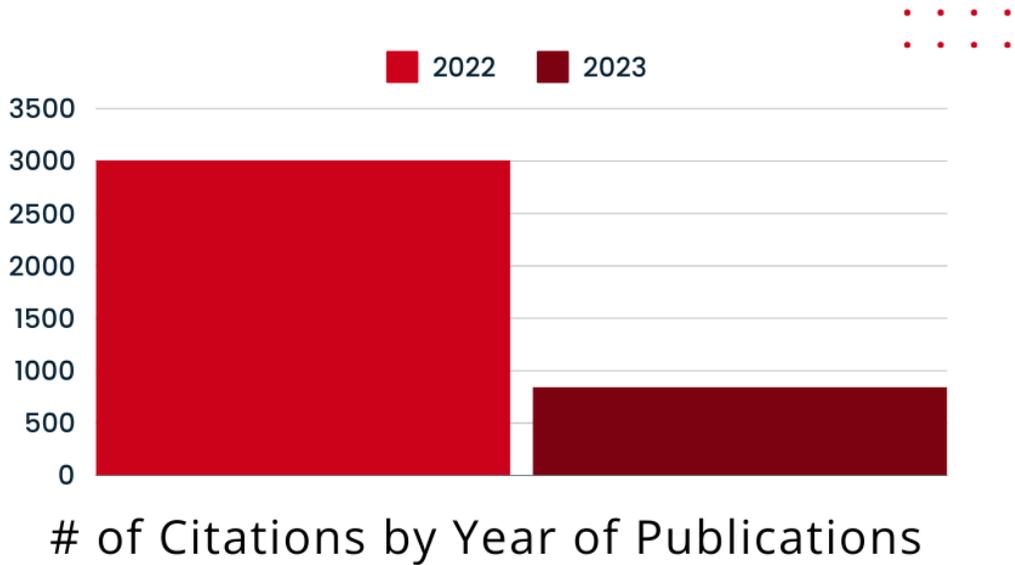
Minoo, Parham (Clinical)			
Appendiceal Mucocele: A preneoplastic lesion for mucinous neoplasm?	2023	Alberta Health Services / Alberta Precision Laboratories	\$7,452
Nohr, Erik (GFT)			
Advanced Stage Lung Adenocarcinoma: Assessing Liquid Cytology Molecular Biomarker Testing	2023	AstraZeneca Canada Inc.	\$49,969
Orton, Dennis (Clinical)			
Defining an optimal IGF-1 concentration in children undergoing in Treatment of Growth Hormone Deficiency	2022	Pfizer Canada	\$205,850
Pillai, Dylan (GFT)			
A multi-disciplinary approach for the development of a point of care solution for the detection of SARS-CoV-2	2022	Deep Biologics Inc.	\$48,924
Combating Antibiotic Resistance in Philippine Lakes: One-Health upstream interventions to reduce the burden	2022-2023	Canadian Institutes of Health Research	\$329,932
LAMP to detect malaria in pregnancy (LAMPREG)	2022-2023	Grand Challenges Canada	\$310,000
Quantification of SARS-CoV-2 viral load in clinical and environmental samples	2022	Canada Foundation for Innovation	\$24,452
The Alberta Diagnostics Ecosystem Platform for Translation (ADEPT)	2023	Alberta Innovates	\$235,000
Pitout, Johann (GFT)			
Genomic epidemiology of Escherichia coli ST167 with carbapenemases	2023	Alberta Health Services / Alberta Precision Laboratories	\$4,760
The Molecular Basis of the Carbapenem Resistance Epidemic	2022	Hackensack Meridian Health	\$68,322
Zargham, Ramin (Clinical)			
Characterization of alpha 8 integrin as a novel prognostic factor for invasive breast ductal carcinoma	2022	Alberta Health Services / Alberta Precision Laboratories	\$10,574
SWI/SNF defected Lung Adenocarcinoma	2022	Alberta Health Services / Alberta Precision Laboratories	\$11,774
Zhang, Kunyan (GFT)			
Center for Antimicrobial Resistance (CAR)	2022-2023	Alberta Health Services / Calgary Health Region	\$135,000

Publications (* indicates First or Last Author)

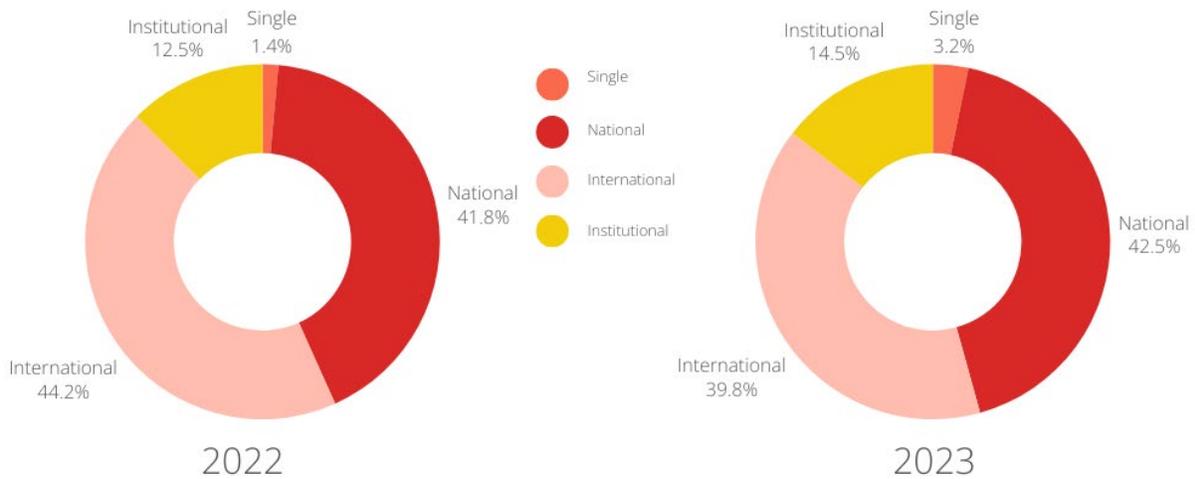


Publication Type by Year





Academia Collaboration by Year



Abdullah, Amid

- de Koning, L., Orton, D., Seiden Long, I., Boyd, J., Kellogg, M., **Abdullah, A.**, Naugler, C., Tsui, A., Strange, B., & Glaser, D. (2023). Distribution of videos demonstrating best practices in preventing hemolysis is associated with reduced hemolysis among nurse-collected specimens in hospitals. *Clinical Biochemistry*, 119, 110632. <https://doi.org/10.1016/j.clinbiochem.2023.110632>

2022

19.85

hindex

15

Pubs As First Author

55

of Pubs As Last Author

3

of Pubs As Single Author

1.55

Field Weighted Citation Impact

208

of Unique Publications

2365

Citations

2023

19.94

hindex

25

Pubs As First Author

62

of Pubs As Last Author

6

of Pubs As Single Author

1.41

Field Weighted Citation Impact

186

of Unique Publications

734

Citations

Collaboration Countries



Al-Shamma, Zainab

1. Alshamma, D. A., Zainab A. A., **Al-Shamma, Z.**, & Amer, A. (2022). Phytochemical Comparison Study for Evaluating the Hypolipidemic Effect Between Two Iraqi Pepper Spp in the Rats Model. *Biomedical and Pharmacology Journal*, 15(4), 2421–2435. <https://doi.org/10.13005/bpj/2580>

Bakker, Andrea

1. ***Bakker, A.**, Slack, J. C., Palanisamy, N., Carskadon, S., Ghosh, S., Khalifeh, I., & Bismar, T. A. (2022). Loss of *KLK4::KLK1* pseudogene expression by RNA chromogenic in-situ hybridization is associated with PTEN loss and increased risk of biochemical recurrence in a cohort of middle eastern men with prostate cancer. *Journal of Cancer Research and Clinical Oncology*, 149(7), 3721–3728. <https://doi.org/10.1007/s00432-022-04279-5>
2. Gamallat, Y., **Bakker, A.**, Khosh Kish, E., Choudhry, M., Walker, S., Aldakheel, S., Seyedi, S., Huang, K.-C., Ghosh, S., Gotto, G., & Bismar, T. A. (2022). The Association between Cyclin Dependent Kinase 2 Associated Protein 1 (CDK2AP1) and Molecular Subtypes of Lethal Prostate Cancer. *International Journal of Molecular Sciences*, 23(21), 13326. <https://doi.org/10.3390/ijms232113326>
3. Mah, J., Bakker, A., Tseng, C., Lafay-Cousin, L., Kuhn, S., Marie-Anne Brundler, & Lisboa, L. F. (2022). Isolated Pulmonary Emergomycosis in an Immunocompetent Patient in Alberta, Canada. *Open Forum Infectious Diseases*, 9(3). <https://doi.org/10.1093/ofid/ofac021>

Benediktsson, Hallgrimur

1. Bau, J. T., Churchill, L., Pandher, M., **Benediktsson, H.**, Tibbles, L. A., & Gill, S. (2022). Acute Kidney Allograft Rejection Following Coronavirus mRNA Vaccination: A Case Report. *Transplantation Direct*, 8(2), e1274. <https://doi.org/10.1097/txd.0000000000001274>
2. Brandt, P. J., Stepanchuk, A. A., Andonegui, G., **Hallgrimur Benediktsson**, Stys, P. K., & Muruve, D. A. (2022). Detection and Typing of Renal Amyloidosis by Fluorescence Spectroscopy Using the Environmentally Sensitive Fluorophore K114. *Molecular Imaging and Biology*, 25(1), 221–227. <https://doi.org/10.1007/s11307-022-01754-w>
3. Groves, R. A., Mapar, M., Raied Aburashed, Ponce, L. F., Bishop, S. L., Rydzak, T., Marija Drikic, Bihan, D. G., **Hallgrimur Benediktsson**, Clement, F., Gregson, D. B., & Lewis, I. A. (2022). Methods for Quantifying the Metabolic Boundary Fluxes of Cell Cultures in Large Cohorts by High-Resolution Hydrophilic Liquid Chromatography Mass Spectrometry. *Analytical Chemistry*, 94(25), 8874–8882. <https://doi.org/10.1021/acs.analchem.2c00078>
4. Lau, A., Rahn, J. J., Chappellaz, M., Chung, H., **Hallgrimur Benediktsson**, Bihan, D., Anne von Mässenhausen, Linkermann, A., Jenne, C. N., Robbins, S. M., Senger, D. L., Lewis, I. A., Chun, J., & Muruve, D. A. (2022). Dipeptidase-1 governs renal inflammation during ischemia reperfusion injury. *Science Advances*, 8(5). <https://doi.org/10.1126/sciadv.abm0142>

Berenger, Byron

1. ***Bibby, H. L.**, Lawrence de Koning, Isolde Seiden-Long, Zelyas, N., Church, D. L., & **Berenger, B. M.** (2022). A pragmatic randomized controlled trial of rapid on-site influenza and respiratory syncytial virus PCR testing in paediatric and adult populations. *BMC Infectious Diseases*, 22(1). <https://doi.org/10.1186/s12879-022-07796-3>
2. Flemons, K., Baylis, B., Khan, A. Z., Kirkpatrick, A. W., Whitehead, K., Moeini, S., Schreiber, A., Lapointe, S., Ashoori, S., Arif, M., **Berenger, B.**, Conly, J., & Hawkins, W. (2022). The use of drones for the delivery of diagnostic test kits and medical supplies to remote First Nations communities during Covid-19. *American Journal of Infection Control*, 50(8), 849–856. <https://doi.org/10.1016/j.ajic.2022.03.004>
3. Freedman, S. B., Oberding, L. K., Kim, K., Xie, J., **Berenger, B. M.**, Goulden, R., Weisbeck, S., & Pillai, D. R. (2022). SARS-CoV-2 Viral Load Quantification, Clinical Findings and Outcomes in Children Seeking Emergency Department Care: Prospective Cohort Study. *Pediatric Infectious Disease Journal*, 41(7), 566–569. <https://doi.org/10.1097/inf.0000000000003527>

4. Leal, J., O'Grady, H. M., Armstrong, L., Dixit, D., Khawaja, Z., Snedeker, K., Ellison, J., Erebor, J., Jamieson, P., Weiss, A., Salcedo, D., Roberts, K., Wiens, K., Croxen, M. A., **Berenger, B. M.**, Kanti Pabbaraju, Lin, Y.-C., Evans, D. H., & Conly, J. (2023). Patient and ward related risk factors in a multi-ward nosocomial outbreak of COVID-19: Outbreak investigation and matched case-control study. *Antimicrobial Resistance and Infection Control*, 12(1). <https://doi.org/10.1186/s13756-023-01215-1>
5. Lin, Y.-C., Malott, R. J., Ward, L., Kiplagat, L., Pabbaraju, K., Gill, K., **Berenger, B. M.**, Hu, J., Fonseca, K., Noyce, R. S., Louie, T., Evans, D. H., & Conly, J. M. (2022). Detection and quantification of infectious severe acute respiratory coronavirus-2 in diverse clinical and environmental samples. *Scientific Reports*, 12(1), 5418. <https://doi.org/10.1038/s41598-022-09218-5>
6. O'Grady, H. M., Harrison, R., Snedeker, K., Trufen, L., Yue, P., Ward, L., Fifen, A., Jamieson, P., Weiss, A., Coulthard, J., Lynch, T., Croxen, M. A., Li, V., Pabbaraju, K., Wong, A., Zhou, H. Y., Dingle, T. C., Hellmer, K., **Berenger, B. M.**, & Fonseca, K. (2023). A two-ward acute care hospital outbreak of SARS-CoV-2 delta variant including a point-source outbreak associated with the use of a mobile vital signs cart and sub-optimal doffing of personal protective equipment. *Journal of Hospital Infection*, 131, 1–11. <https://doi.org/10.1016/j.jhin.2022.09.019>
7. Razieh Salahandish, Jae Eun Hyun, Fatemeh Haghayegh, Hamed Osouli Tabrizi, Shirin Moossavi, Sultan Khetani, Giancarlo Ayala-Charca, **Berenger, B. M.**, Yan Dong Niu, Ebrahim Ghafar-Zadeh, & Amir Sanati Nezhad. (2023). CoVSense: Ultrasensitive Nucleocapsid Antigen Immunosensor for Rapid Clinical Detection of Wildtype and Variant SARS-CoV-2. *Advanced Science*, 10(15). <https://doi.org/10.1002/advs.202206615>
8. Razieh Salahandish, Jalali, P., Hamed Osouli Tabrizi, Jae Min Hyun, Fatemeh Haghayegh, Mahmood Khalghollah, Zare, A., **Berenger, B. M.**, Niu, Y., Ebrahim Ghafar-Zadeh, & Amir Sanati-Nezhad. (2022). A compact, low-cost, and binary sensing (BiSense) platform for noise-free and self-validated impedimetric detection of COVID-19 infected patients. *Biosensors and Bioelectronics*, 213(114459), 114459–114459. <https://doi.org/10.1016/j.bios.2022.114459>
9. Salahandish, R., Haghayegh, F., Ayala-Charca, G., Hyun, J. E., Khalghollah, M., Zare, A., Far, B., **Berenger, B. M.**, Niu, Y. D., Ghafar-Zadeh, E., & Sanati-Nezhad, A. (2022). Bi-ECDAQ: An electrochemical dual-immuno-biosensor accompanied by a customized bi-potentiostat for clinical detection of SARS-CoV-2 Nucleocapsid proteins. *Biosensors and Bioelectronics*, 203, 114018. <https://doi.org/10.1016/j.bios.2022.114018>
10. Stokes, W., **Berenger, B. M.**, Scott, B., Szelewicki, J., Singh, T., Portnoy, D., Turnbull, L., Pabbaraju, K., Shokoples, S., Wong, A. A., Gill, K., Hu, J., & Tipples, G. (2022). One Swab Fits All: Performance of a Rapid, Antigen-Based SARS-CoV-2 Test Using a Nasal Swab, Nasopharyngeal Swab for Nasal Collection, and RT-PCR Confirmation from Residual Extraction Buffer. *The Journal of Applied Laboratory Medicine*. <https://doi.org/10.1093/jalm/jfac004>
11. Stokes, W., **Berenger, B. M.**, Venner, A. A., Deslandes, V., & Julie. (2022). Point of care molecular and antigen detection tests for COVID-19: current status and future prospects. *Expert Review of Molecular Diagnostics*, 22(8), 797–809. <https://doi.org/10.1080/14737159.2022.2122712>
12. *Stokes, W., Venner, A. A., Buss, E., Tipples, G., & **Berenger, B. M.** (2023). Evaluation of the ID NOW among symptomatic individuals during the Omicron wave. *Journal of Medical Microbiology*, 72(2). <https://doi.org/10.1099/jmm.0.001669>
13. *Stokes, W., Venner, A. A., Buss, E., Tipples, G., & **Berenger, B. M.** (2023a). Prospective population-level validation of the Abbott ID NOW severe acute respiratory syndrome coronavirus 2 device implemented in multiple settings for testing asymptomatic and symptomatic individuals. *Clinical Microbiology and Infection*, 29(2), 247–252. <https://doi.org/10.1016/j.cmi.2022.08.025>
14. Thornton, C. S., Huntley, K., **Berenger, B. M.**, Bristow, M., Evans, D. H., Fonseca, K., Franko, A., Gillrie, M. R., Lin, Y.-C., Povitz, M., Shafey, M., Conly, J. M., & Tremblay, A. (2022). Prolonged

SARS-CoV-2 infection following rituximab treatment: clinical course and response to therapeutic interventions correlated with quantitative viral cultures and cycle threshold values. *Antimicrobial Resistance & Infection Control*, 11(1). <https://doi.org/10.1186/s13756-022-01067-1>

15. Vayalunkal, J. V., Soraisham, A. S., Ayman Abou Mehrem, Ghosh, A., Jessica, Fonseca, K., Zhou, H., **Berenger, B. M.**, Chan, E. S., Marie-Anne Brundler, Lin, Y.-C., Evans, D. H., Rousso, S., Kuret, V., & Conly, J. M. (2023). Congenital SARS-CoV-2 Infection in Two Neonates with Confirmation by Viral Culture of the Placenta in One Case. *Viruses*, 15(6), 1310–1310. <https://doi.org/10.3390/v15061310>
16. Xie, J., Kim, K., **Berenger, B. M.**, Chui, L., Vanderkooi, O. G., Silviu Grisaru, & Freedman, S. B. (2023). Comparison of a Rapid Multiplex Gastrointestinal Panel with Standard Laboratory Testing in the Management of Children with Hematochezia in a Pediatric Emergency Department: Randomized Controlled Trial. *Microbiology Spectrum*, 11(3). <https://doi.org/10.1128/spectrum.00268-23>

Berka, Nouredine

1. *Abu-Khader, A., Gareau, A. J., Burns, C., Khan, F., & **Berka, N.** (2022). Next-generation sequencing identifies two novel HLA class II alleles, HLA-DRB1*01:115 and HLA-DRB1*14:224. *HLA*, 100(2), 176–177. <https://doi.org/10.1111/tan.14623>
2. *Abu-Khader, A., Hu, Q., Kamar, F., Iwona Galaszkiwicz, Wang, W., Khan, F., & **Berka, N.** (2023). Low incidence of de novo HLA antibodies after COVID-19 vaccination: A cohort study of patients awaiting kidney transplantation. *Transplant Infectious Disease*, 25(6). <https://doi.org/10.1111/tid.14105>
3. Yang, G., Abu-Khader, A., Burns, C., Galaszkiwicz, I., & **Berka, N.** (2022). The novel HLA class II allele, DPB1*1284:01, identified using next-generation sequencing. *HLA*. <https://doi.org/10.1111/tan.14587>
4. *Hu, Q., Burns, C., Christian, D., Yang, G., & **Berka, N.** (2023). Discovery of three novel HLA-DPA1 alleles, HLA-DPA1*01:147N, 01:03:47, and 02:106 using next-generation sequencing. *HLA: Immune Response Genetics*, 102(3), 385–387. <https://doi.org/10.1111/tan.15125>
5. *Hu, Q., Burns, C., McKinley, C., Christian, D., & **Berka, N.** (2022). Identification of a novel allele with a frameshift mutation, HLA-DRB4*01:165N, using next-generation sequencing. *HLA: Immune Response Genetics*, 101(6), 699–700. <https://doi.org/10.1111/tan.14954>
6. *Hu, Q., Christian, D., Yang, G., Burns, C., & **Berka, N.** (2023b). Identification of four novel HLA-DQ alleles, HLA-DQA1*01:106, -DQA1*01:107, -DQA1*05:74 and -DQB1*05:01:48. *HLA: Immune Response Genetics*, 102(2), 254–256. <https://doi.org/10.1111/tan.15076>
7. *Hu, Q., Christian, D., Yang, G., McKinley, C., & **Berka, N.** (2023). Two novel HLA class I alleles, HLA-C*04:493 and -A*26:01:78, identified using next-generation sequencing. *HLA: Immune Response Genetics*, 101(6), 686–687. <https://doi.org/10.1111/tan.14985>
8. Tripathi, G., Khanolkar, R. A., Faridi, R. M., Kalra, A., Poonam Dharmani-Khan, Meer-Taher Shabani-Rad, **Berka, N.**, Daly, A., Storek, J., & Khan, F. M. (2022). Donor Genetic Predisposition to High Interleukin-10 Production Appears Protective against Acute Graft-Versus-Host Disease. *International Journal of Molecular Sciences*, 23(24), 15888–15888. <https://doi.org/10.3390/ijms232415888>
9. *Yang, G., Abu-Khader, A., Burns, C., Galaszkiwicz, I., & **Berka, N.** (2022). The novel HLA class II allele,

Bismar, Tarek

1. Kiciak, A., Clark, W., Uhlich, M., Letendre, A., Littlechild, R., Lightning, P., Vásquez, C., Singh, R., Broomfield, S., Anaïs Medina Martín, Huang, G., Fairey, A., Kolinsky, M., Christopher J.D. Wallis,

- Fung, C., Hyndman, E., Yip, S., **Bismar, T. A.**, Lewis, J. D., & Ghosh, S. (2023). Disparities in prostate cancer screening, diagnoses, management, and outcomes between Indigenous and non-Indigenous men in a universal health care system. *Cancer*, 129(18), 2864–2870. <https://doi.org/10.1002/cncr.34812>
2. *Ahmed Al Marouf, Alhajj, R., Rokne, J. G., Ghose, S., & **Bismar, T. A.** (2023). Exploring Gene Expression and Clinical Data for Identifying Prostate Cancer Severity Levels using Machine Learning Methods. *Canadian Conference on Electrical and Computer Engineering*. <https://doi.org/10.1109/ccece58730.2023.10288946>
 3. *Bakker, A., Slack, J. C., Nalla Palanisamy, Carskadon, S., Ghosh, S., Khalifeh, I., & **Bismar, T. A.** (2022b). Loss of KLK4::KLKP1 pseudogene expression by RNA chromogenic in-situ hybridization is associated with PTEN loss and increased risk of biochemical recurrence in a cohort of middle eastern men with prostate cancer. *Journal of Cancer Research and Clinical Oncology*, 149(7), 3721–3728. <https://doi.org/10.1007/s00432-022-04279-5>
 4. *Choudhry, M., Gamallat, Y., Khosh Kish, E., Seyedi, S., Gotto, G., Ghosh, S., & **Bismar, T. A.** (2023). Downregulation of BUD31 Promotes Prostate Cancer Cell Proliferation and Migration via Activation of p-AKT and Vimentin In Vitro. *International Journal of Molecular Sciences*, 24(7), 6055. <https://doi.org/10.3390/ijms24076055>
 5. *Choudhry, M., Yaser Gamallat, Ghosh, S., & **Bismar, T. A.** (2023). Cleavage and Polyadenylation-Specific Factor 4 (CPSF4) Expression Is Associated with Enhanced Prostate Cancer Cell Migration and Cell Cycle Dysregulation, In Vitro. *International Journal of Molecular Sciences*, 24(16), 12961–12961. <https://doi.org/10.3390/ijms241612961>
 6. *Gamallat, Y., Bakker, A., Khosh Kish, E., Choudhry, M., Walker, S., Aldakheel, S., Seyedi, S., Huang, K.-C., Ghosh, S., Gotto, G., & **Bismar, T. A.** (2022b). The Association between Cyclin Dependent Kinase 2 Associated Protein 1 (CDK2AP1) and Molecular Subtypes of Lethal Prostate Cancer. *International Journal of Molecular Sciences*, 23(21), 13326. <https://doi.org/10.3390/ijms232113326>
 7. *Gamallat, Y., Zaaluk, H., Kish, E. K., Abdelsalam, R., Liosis, K., Ghosh, S., & **Bismar, T. A.** (2022). ARPC1B Is Associated with Lethal Prostate Cancer and Its Inhibition Decreases Cell Invasion and Migration In Vitro. *International Journal of Molecular Sciences*, 23(3), 1476. <https://doi.org/10.3390/ijms23031476>
 8. *Khosh Kish, E., Choudhry, M., Gamallat, Y., Buharideen, S. M., D, D., & **Bismar, T. A.** (2022). The Expression of Proto-Oncogene ETS-Related Gene (ERG) Plays a Central Role in the Oncogenic Mechanism Involved in the Development and Progression of Prostate Cancer. *International Journal of Molecular Sciences*, 23(9), 4772. <https://doi.org/10.3390/ijms23094772>
 9. *Khosh Kish, E., Gamallat, Y., Choudhry, M., Ghosh, S., Seyedi, S., & **Bismar, T. A.** (2023). Glycyl-tRNA Synthetase (GARS) Expression Is Associated with Prostate Cancer Progression and Its Inhibition Decreases Migration, and Invasion In Vitro. *International Journal of Molecular Sciences*, 24(5), 4260. <https://doi.org/10.3390/ijms24054260>
 10. Konstantinos Christos Liosis, Ahmed Al Marouf, Rokne, J. G., Ghosh, S., **Bismar, T. A.**, & Alhajj, R. (2023). Genomic Biomarker Discovery in Disease Progression and Therapy Response in Bladder Cancer Utilizing Machine Learning. *Cancers*, 15(19), 4801–4801. <https://doi.org/10.3390/cancers15194801>
 11. Seltzer, S., Giannopoulos, P. N., **Bismar, T. A.**, Trifiro, M., & Miltiadis Paliouras. (2022). Investigation of androgen receptor-dependent alternative splicing has identified a unique subtype of lethal prostate cancer. *Asian Journal of Andrology*, 25(3), 296–308. <https://doi.org/10.4103/aja202263>
 12. *Yaser Gamallat, & **Bismar, T. A.** (2023). Editorial: The Application of Proteogenomics to Urine Analysis for the Identification of Novel Biomarkers of Prostate Cancer: An Exploratory Study. *Cancers*, 15(16), 4143–4143. <https://doi.org/10.3390/cancers15164143>

13. *Yaser Gamallat, Choudhry, M., Li, Q., Jon George Rokne, Alhadj, R., Abdelsalam, R., Ghosh, S., Jaron Arbet, Boutros, P. C., & **Bismar, T. A.** (2023). Serrate RNA Effector Molecule (SRRT) Is Associated with Prostate Cancer Progression and Is a Predictor of Poor Prognosis in Lethal Prostate Cancer. *Cancers*, 15(10), 2867–2867. <https://doi.org/10.3390/cancers15102867>
14. *Yaser Gamallat, Mitra Afsharpad, Soufiane El Hallani, Maher, C. A., Nimira Alimohamed, Hyndman, E., & **Bismar, T. A.** (2023). Large, Nested Variant of Urothelial Carcinoma Is Enriched with Activating Mutations in Fibroblast Growth Factor Receptor-3 among Other Targetable Mutations. *Cancers*, 15(12), 3167–3167. <https://doi.org/10.3390/cancers15123167>

Box, Adrian

1. Brenner, D. R., O’Sullivan, D. E., Tamer Jarada, Yusuf, A., Boyne, D. J., Mather, C., **Box, A.**, Morris, D., Cheung, W. Y., & Mirza, I. (2023). The impact of population-based EGFR testing in non-squamous metastatic non-small cell lung cancer in Alberta, Canada. *Lung Cancer*, 175, 60–67. <https://doi.org/10.1016/j.lungcan.2022.11.017>
2. Eszlinger, M., Stewardson, P., McIntyre, J. B., **Box, A.**, Khalil, M., Hycza, M., Konstantin Koro, Ruether, D., Wu, J., & Paschke, R. (2022). Systematic population-based identification of NTRK and RET fusion-positive thyroid cancers. *European Thyroid Journal*, 11(1). <https://doi.org/10.1530/etj-21-0061>
3. Gibson, A. J. W., **Box, A.**, Cheung, W. Y., Dean, M. L., Elegbede, A. A., Hao, D., Pabani, A., Sangha, R., & Bebb, D. G. (2022). Real-World Management and Outcomes of Crizotinib-Treated ROS1-Rearranged NSCLC: A Retrospective Canadian Cohort. *Current Oncology*, 29(3), 1967–1982. <https://doi.org/10.3390/curroncol29030160>
4. Slack, J. C., Marie-Anne Bründler, **Box, A.**, & Konstantin Koro. (2022). A Subset of Pancreatoblastomas May Arise From an Adenomatous Precursor: An Ampullary Pancreatoblastoma and Adjacent Adenoma With a Shared Molecular Phenotype in an Adult Patient. *Pancreas*, 51(10), 1455–1460. <https://doi.org/10.1097/mpa.0000000000002189>
5. Stewardson, P., Eszlinger, M., Wu, J., Khalil, M., **Box, A.**, Perizzolo, M., Zoya Punjwani, Ziehr, B., Sanyal, R., Demetrick, D. J., & Paschke, R. (2023). Prospective Validation of ThyroSPEC Molecular Testing of Indeterminate Thyroid Nodule Cytology Following Diagnostic Pathway Optimization. *Thyroid*, 33(12), 1423–1433. <https://doi.org/10.1089/thy.2023.0255>
6. Stockley, T. L., Lo, B., **Box, A.**, Andrea Gomez Corredor, DeCoteau, J., Desmeules, P., Feilotter, H., Grafodatskaya, D., Hawkins, C., Weei Yuarn Huang, Iyare Izevbaye, Lepine, G., Papadakis, A. I., Park, P. C., Sheffield, B. S., Danh Tran-Thanh, Yip, S., & Ming Sound Tsao. (2023). Consensus Recommendations to Optimize the Detection and Reporting of NTRK Gene Fusions by RNA-Based Next-Generation Sequencing. *Current Oncology*, 30(4), 3989–3997. <https://doi.org/10.3390/curroncol30040302>
7. Stockley, T., Lo, B., **Box, A.**, Corredor, A., DeCoteau, J. F., Desmeules, P., Feilotter, H., Grafodatskaya, D., Greer, W. L., Hawkins, C., Weei Yuarn Huang, Iyare Izevbaye, Guylaine Lépine, Sebastiao, Papadakis, A. I., Park, P. C., Jean-Baptiste Rivière, Sheffield, B. S., Spatz, A., & Spriggs, E. (2023). CANTRK: A Canadian Ring Study to Optimize Detection of NTRK Gene Fusions by Next-Generation RNA Sequencing. *The Journal of Molecular Diagnostics*, 25(3), 168–174. <https://doi.org/10.1016/j.jmoldx.2022.12.004>

Boyd, Jessica

1. de Koning, L., Orton, D., Seiden Long, I., **Boyd, J.**, Kellogg, M., Abdullah, A., Naugler, C., Tsui, A., Strange, B., & Glaser, D. (2023b). Distribution of videos demonstrating best practices in preventing hemolysis is associated with reduced hemolysis among nurse-collected specimens

in hospitals. *Clinical Biochemistry*, 119, 110632.

<https://doi.org/10.1016/j.clinbiochem.2023.110632>

2. Wickramarachchi, P., **Boyd, J. M.**, & Orton, D. J. (2022). Biological variation in clozapine and metabolite reporting during therapeutic drug monitoring. *Clinica Chimica Acta*, 531, 183–187. <https://doi.org/10.1016/j.cca.2022.04.005>

Brenn, Thomas

1. *Caldeira, C., Deolino João Camilo-Júnior, Luiz, A., Paulo, P., Roberto, A., & **Brenn, T.** (2022). Value of Immunohistochemistry to Differentiate Digital Papillary Adenocarcinoma From Acral Hidradenoma With Papillary Structures. *The American Journal of Dermatopathology*, 44(11), 843–845. <https://doi.org/10.1097/dad.0000000000002291>
2. *Hornick, J. L., Plaza, J. A., Mentzel, T., Gru, A. A., & **Brenn, T.** (2023). PRAME Expression Is a Useful Tool in the Diagnosis of Primary and Metastatic Dedifferentiated and Undifferentiated Melanoma. *The American Journal of Surgical Pathology*, 47(12), 1390–1397. <https://doi.org/10.1097/pas.0000000000002125>
3. Kinaschuk, K., Cheng, T., **Brenn, T.**, J. Gregory McKinnon, & Temple-Oberle, C. (2023). Not Waiting to Progress; How the COVID-19 Pandemic Nudged Neoadjuvant Therapy for Stage III Locally Advanced Melanoma Patients. *Current Oncology*, 30(5), 4402–4411. <https://doi.org/10.3390/curroncol30050335>
4. *Lach, K., Gharpuray-Pandit, D., Franko, A., & **Brenn, T.** (2023). Distant metastasis form morphologically low-grade spiradenocarcinoma: a report of two cases. *Journal of Cutaneous Pathology*. <https://doi.org/10.1111/cup.14388>
5. *Mehta, A., Davey, J., Deepa Gharpuray-Pandit, Plaza, J. A., Billings, S. D., Balogh, P., Nairi Tchraikian, Sharmila Selvan, Wiedemeyer, K., Hornick, J. L., & **Brenn, T.** (2022). Cutaneous Myoepithelial Neoplasms on Acral Sites Show Distinctive and Reproducible Histopathologic and Immunohistochemical Features. *The American Journal of Surgical Pathology*, 46(9), 1241–1249. <https://doi.org/10.1097/pas.0000000000001896>
6. *Mitteldorf, C., & **Brenn, T.** (2022). Ist die Nomenklatur atypisches Fibroxanthom und pleomorphes dermales Sarkom noch zeitgemäß? *Journal Der Deutschen Dermatologischen Gesellschaft*, 20(12), 1563–1564. https://doi.org/10.1111/ddg.14972_g
7. Plaza, J. A., **Brenn, T.**, Gru, A. A., Matoso, A., Sheldon, J., & Sangueza, M. (2022). Poorly differentiated cutaneous apocrine carcinomas: histopathological clues and immunohistochemical analysis for the diagnosis of this unusual neoplasm. *Histopathology*, 82(2), 276–284. <https://doi.org/10.1111/his.14809>
8. *Plaza, J. A., Wakely, P., Roman, J., Gru, A. A., J. Martin Sangueza, Davey, J., & **Brenn, T.** (2023). Low-grade Hidradenocarcinomas: A Clinicopathologic Study of an Unusual Carcinoma That Can Mimic its Benign Counterpart. *The American Journal of Surgical Pathology*, 47(8), 907–914. <https://doi.org/10.1097/pas.0000000000002065>
9. Pourmaleki, M., Jones, C. J., Ariyan, C. E., Zeng, Z., Mono Pirun, Navarrete, D. A., Li, Y., Zhang, M., Subhiksha Nandakumar, Campos, C., Nadeem, S., Klimstra, D. S., Temple-Oberle, C. F., **Brenn, T.**, Lipson, E. J., Schenk, K. M., Stein, J. E., Taube, J. M., White, M. G., & Traweek, R. (2022). Tumor MHC Class I Expression Associates with Intralesional IL2 Response in Melanoma. *Cancer Immunology Research*, 10(3), 303–313. <https://doi.org/10.1158/2326-6066.cir-21-1083>
10. Vanderbilt, C., **Brenn, T.**, Moy, A. P., Harloe, G., Ariyan, C., Athanasian, E., & Busam, K. J. (2022). Association of HPV42 with digital papillary adenocarcinoma and the use of in situ hybridization for its distinction from acral hidradenoma and diagnosis at non-acral sites. *Modern Pathology*, 35(10), 1405–1410. <https://doi.org/10.1038/s41379-022-01094-8>

11. *Wiedemeyer, K., & **Brenn, T.** (2023). Dedifferentiated and undifferentiated melanomas: a practical approach to a challenging diagnosis. *Human Pathology*, 140, 22–31. <https://doi.org/10.1016/j.humpath.2023.03.014>
12. Wiedemeyer, K., **Brenn, T.**, & Naert, K. (2022). Necrotizing Infundibular Crystalline Folliculitis—A Case Report of a Rare Entity and Review of the Literature. *The American Journal of Dermatopathology*, 44(10), 774–777. <https://doi.org/10.1097/dad.0000000000002239>

Bromley, Amy

1. Sarden, N., Sinha, S., Potts, K. G., Pernet, E., Hiroki, C. H., Hassanabad, M. F., Nguyen, A. P., Lou, Y., Farias, R., Winston, B. W., **Bromley, A.**, Snarr, B. D., Zucoloto, A. Z., Andonegui, G., Muruve, D. A., McDonald, B., Sheppard, D. C., Mahoney, D. J., Divangahi, M., & Rosin, N. (2022b). A B1a–natural IgG–neutrophil axis is impaired in viral- and steroid-associated aspergillosis. *Science Translational Medicine*, 14(674). <https://doi.org/10.1126/scitranslmed.abq6682>

Brundler, Marie-Ann

1. Espinoza, **M. L.**, **Brundler**, M.-A., Hasan, S. U., Mohammad, K., Momin, S., Al Shaikh, B., & Yusuf, K. (2022). Placental pathology as a marker of brain injury in infants with hypoxic ischemic encephalopathy. *Early Human Development*, 174, 105683. <https://doi.org/10.1016/j.earlhumdev.2022.105683>
2. Ginter, D., Ramien, M., **Marie-Anne Brundler**, Swaney, L. C., Paivi Miettunen, & Luca, N. (2022). A rare case of suspected lupus erythematosus panniculitis as the presenting skin feature of juvenile dermatomyositis: A case report. *SAGE Open Medical Case Reports*, 10, 2050313X2210863–2050313X2210863. <https://doi.org/10.1177/2050313x221086317>
3. Mah, J., Bakker, A., Tseng, C., Lafay-Cousin, L., Kuhn, S., **Marie-Anne Brundler**, & Lisboa, L. F. (2022). Isolated Pulmonary Emergomycosis in an Immunocompetent Patient in Alberta, Canada. *Open Forum Infectious Diseases*, 9(3). <https://doi.org/10.1093/ofid/ofac021>
4. Slack, J. C., **Marie-Anne Brundler**, Box, A., & Konstantin Koro. (2022). A Subset of Pancreatoblastomas May Arise From an Adenomatous Precursor: An Ampullary Pancreatoblastoma and Adjacent Adenoma With a Shared Molecular Phenotype in an Adult Patient. *Pancreas*, 51(10), 1455–1460. <https://doi.org/10.1097/mpa.0000000000002189>
5. Vayalumkal, J. V., Soraisham, A. S., Ayman Abou Mehrem, Ghosh, A., Jessica, Fonseca, K., Zhou, H., Berenger, B. M., Chan, E. S., **Marie-Anne Brundler**, Lin, Y.-C., Evans, D. H., Rousso, S., Kuret, V., & Conly, J. M. (2023b). Congenital SARS-CoV-2 Infection in Two Neonates with Confirmation by Viral Culture of the Placenta in One Case. *Viruses*, 15(6), 1310–1310. <https://doi.org/10.3390/v15061310>
6. Wright, J. R., Chan, S., Morgen, E. K., Maung, R. T. A., **Marie-Anne Brundler**, Dina El Demellawy, Fraser, R. B., Kurek, K. C., Magee, F., Nizalik, E., Oligny, L. L., Somers, G. R., Stefanovici, C., & Terry, J. (2022). Workload Measurement in Subspecialty Placental Pathology in Canada. *Pediatric and Developmental Pathology*, 25(6), 604–610. <https://doi.org/10.1177/10935266221118150>

Caragea, Mara

1. Choi, K.-Y., Mack, L., **Caragea, M.**, Monument, M., Puloski, S., & Bouchard-Fortier, A. (2022). MDM2 analysis in the management of benign lipomas versus atypical lipomatous tumors/well-differentiated liposarcomas: A useful prognostication tool? *The American Journal of Surgery*, 224(2), 747–750. <https://doi.org/10.1016/j.amjsurg.2022.03.052>

Carson, Julie

1. Bourassa-Blanchette, S., Biesheuvel, M. M., Lam, J. C., Kipp, A., Church, D., **Carson, J.**, Dalton, B., Parkins, M. D., Barkema, H. W., & Gregson, D. B. (2023). Incidence, susceptibility and outcomes of candidemia in adults living in Calgary, Alberta, Canada (2010–2018). *BMC Infectious Diseases*, 23(1). <https://doi.org/10.1186/s12879-023-08050-0>
2. Chow, B. L., Groeschel, M., **Carson, J.**, Griener, T. P., & Church, D. L. (2022). Development and evaluation of a novel fast broad-range PCR and sequencing assay (FBR-PCR/S) using dual priming oligonucleotides targeting the ITS/LSU gene regions for rapid diagnosis of invasive fungal diseases: multi-year experience in a large Canadian healthcare zone. *BMC Infectious Diseases*, 22(1). <https://doi.org/10.1186/s12879-022-07356-9>

Chan, Elaine

1. ***Chan E. S.**, Suchet I, Yu W, Somerset D, Soliman N, Kuret V, Chadha R. (2023) Absence of Ductus Venosus: A Comparison of 2 Distinctive Fetal Autopsy Cases and Embryologic Perspectives. *Pediatric and Developmental Pathology*. doi: 10.1177/10935266231211760.
2. ***Chan, E. S.**, & Stefanovici, C. (2022). Obstructed Hemivagina and Ipsilateral Renal Anomaly (OHVIRA) – A Fetal Autopsy Case. *Journal of Pediatric and Adolescent Gynecology*, 35(5), 593–596. <https://doi.org/10.1016/j.jpbg.2022.02.003>
3. ***Chan, E. S.**, Chadha, R., & de Koning, L. (2023). Maternal Hemoglobin A1c in the Third-Trimester May Underestimate Maternal Hyperglycemia and Its Impact on Offspring in Perinatal Demise Associated With Gestational Diabetes Mellitus: An Autopsy Case Series. *Pediatric and Developmental Pathology*, 26(5), 472–481. <https://doi.org/10.1177/10935266231194697>
4. ***Chan, E. S.**, de Koning, L., Yu, W., & Chadha, R. (2023). C4d Staining Is Present in Normal Placentas From Pregnancies Prior to Pregnancy Loss Associated With Chronic Histiocytic Intervillositis and Is Reduced by Immunomodulatory Therapy in Subsequent Pregnancies. *Pediatric and Developmental Pathology*, 26(4), 374–387. <https://doi.org/10.1177/10935266231176682>
5. ***Chan, E. S.**, Haider, S., Subramanian, S., Yu, W., Nohr, E. W., & de Koning, L. (2022). Congenital cytomegalovirus infection is associated with congenital rickets: a retrospective autopsy cohort study. *Archives of Disease in Childhood: Fetal & Neonatal*, 108(4), 435–436. <https://doi.org/10.1136/archdischild-2022-324760>
6. ***Chan, E. S.**, Mehrem, A. A., de Koning, L., Stritzke, A., & Zhou, H. Y. (2023). Extrahepatic biliary atresia in a premature neonate with congenital cytomegalovirus infection. *Pathology*, 55(4), 573–576. <https://doi.org/10.1016/j.pathol.2022.10.013>
7. ***de Koning, L.**, Crawford, S., Nohr, E., Chadha, R., Horn, C., Wright, J. R., & **Chan, E. S.** (2022). Recurrence risk of villitis of unknown etiology: Analysis of a large retrospective cohort study, systematic review and meta-analysis. *Placenta*, 120, 32–39. <https://doi.org/10.1016/j.placenta.2022.02.006>
8. ***Horn, C.**, Engel, N., Drouin, D., Haley, J., Holder, C., Hung, L., Royall, L., McInnis, P., de Koning, L., & **Chan, E. S.** (2023). Time Required for Gross Examination of Routine Second and Third Trimester Singleton Placentas by Pathologists' Assistants. *Pediatric and Developmental Pathology*, 26(5), 466–471. <https://doi.org/10.1177/10935266231196015>
9. Sarnat, H. B., **Chan, E. S.**, Ng, D., & Yu, W. (2023). Maturation of metastases in peripheral neuroblastic tumors (neuroblastoma) of children. *Journal of Neuropathology and Experimental Neurology*, 82(10), 853–864. <https://doi.org/10.1093/jnen/nlad068>
10. Vayalunkal, J. V., Soraisham, A. S., Mehrem, A. A., Ghosh, A., Jessica, Fonseca, K., Zhou, H., Berenger, B. M., **Chan, E. S.**, Brundler, M.-A., Lin, Y.-C., Evans, D. H., Rousso, S., Kuret, V., & Conly, J. M. (2023). Congenital SARS-CoV-2 Infection in Two Neonates with Confirmation by Viral

Culture of the Placenta in One Case. *Viruses*, 15(6), 1310–1310.
<https://doi.org/10.3390/v15061310>

Chan, Jennifer

1. Berger, N. D., Brownlee, P. M., Chen, M. J., Morrison, H., Osz, K., Ploquin, N. P., **Chan, J. A.**, & Goodarzi, A. A. (2022). High replication stress and limited Rad51-mediated DNA repair capacity, but not oxidative stress, underlie oligodendrocyte precursor cell radiosensitivity. *NAR Cancer*, 4(2). <https://doi.org/10.1093/narcan/zcac012>
2. ***Chan, J. A.**, & Vercauteren, S. M. (2022). Processing and Cryopreservation of Blood, Cancer Tissues, and Cancer Cells for Viable Biobanking. *Methods in Molecular Biology*, 45–58. https://doi.org/10.1007/978-1-0716-2376-3_5
3. Hendrikse, L. D., Haldipur, P., Saulnier, O., Millman, J., Sjoboen, A. H., Erickson, A. W., Ong, W., Gordon, V., Coudière-Morrison, L., Mercier, A. L., Shokouhian, M., Suárez, R. A., Ly, M., Borlase, S., Scott, D. S., Vladiu, M. C., Farooq, H., Sirbu, O., Nakashima, T., & Nambu, S. (2022). Author Correction: Failure of human rhombic lip differentiation underlies medulloblastoma formation. *Nature*, 612(7940), E12–E12. <https://doi.org/10.1038/s41586-022-05578-0>
4. Hendrikse, L. D., Haldipur, P., Saulnier, O., Millman, J., Sjoboen, A. H., Erickson, A. W., Ong, W., Gordon, V., Coudière-Morrison, L., Mercier, A. L., Shokouhian, M., Suárez, R. A., Ly, M., Borlase, S., Scott, D. S., Vladiu, M. C., Farooq, H., Sirbu, O., Nakashima, T., & Nambu, S. (2022a). Failure of human rhombic lip differentiation underlies medulloblastoma formation. *Nature*, 1–9. <https://doi.org/10.1038/s41586-022-05215-w>
5. Karimi, E., Yu, M. W., Maritan, S. M., Perus, L. J. M., Rezanejad, M., Sorin, M., Dankner, M., Fallah, P., Doré, S., Zuo, D., Fiset, B., Kloosterman, D. J., Ramsay, L., Wei, Y., Lam, S., Alsajjan, R., Watson, I. R., Roldan Urgoiti, G., Park, M., & Brandsma, D. (2023). Single-cell spatial immune landscapes of primary and metastatic brain tumours. *Nature*, 614(7948), 555–563. <https://doi.org/10.1038/s41586-022-05680-3>
6. LeBlanc, V. G., Trinh, D. L., Aslanpour, S., Hughes, M., Livingstone, D., Jin, D., Ahn, B. Y., Blough, M. D., Cairncross, J. G., **Chan, J. A.**, Kelly, J. J. P., & Marra, M. A. (2022). Single-cell landscapes of primary glioblastomas and matched explants and cell lines show variable retention of inter- and intratumor heterogeneity. *Cancer Cell*, 40(4), 379–392.e9. <https://doi.org/10.1016/j.ccell.2022.02.016>
7. Nikolic, A., Maule, F., Boby, A., Ellestad, K., So Ya Paik, Marhon, S. A., Mehdipour, P., Lun, X., Chen, H.-M., Mallard, C., Hay, A., Johnston, M. J., Gafuik, C., Zemp, F. J., Shen, Y., Ninkovic, N., Osz, K., Labit, E., Berger, N. D., & Brownsey, D. K. (2023). macroH2A2 antagonizes epigenetic programs of stemness in glioblastoma. *Nature Communications*, 14(1). <https://doi.org/10.1038/s41467-023-38919-2>
8. Takemon, Y., LeBlanc, V. G., Song, J., Chan, S., Lee, S. D., Trinh, D. L., Ahmad, S., Brothers, W. R., Corbett, R., Gagliardi, A., Moradian, A., Cairncross, J. G., Yip, S., Aparicio, S., **Chan, J. A.**, Hughes, C. S., Morin, G. B., Gorski, S. M., Chittaranjan, S., & Marra, M. A. (2023). Multi-Omic Analysis of CIC's Functional Networks Reveals Novel Interaction Partners and a Potential Role in Mitotic Fidelity. *Cancers*, 15(10), 2805–2805. <https://doi.org/10.3390/cancers15102805>
9. Tatari, N., Khan, S., Livingstone, J., Zhai, K., Mckenna, D., Ignatchenko, V., Chokshi, C., Gwynne, W. D., Singh, M., Revill, S., Mikolajewicz, N., Zhu, C., **Chan, J.**, Hawkins, C., Lu, J.-Q., Provias, J. P., Ask, K., Morrissy, S., Brown, S., & Weiss, T. (2022). The proteomic landscape of glioblastoma recurrence reveals novel and targetable immunoregulatory drivers. *Acta Neuropathologica*, 144(6), 1127–1142. <https://doi.org/10.1007/s00401-022-02506-4>
10. Zagozewski, J., Borlase, S., Guppy, B. J., Coudière-Morrison, L., Shahriary, G. M., Gordon, V., Liang, L., Cheng, S., Porter, C. J., Kelley, R., Hawkins, C., **Chan, J. A.**, Liang, Y., Gong, J., Nör, C., Saulnier, O., Wechsler-Reya, R. J., Ramaswamy, V., & Werbowetski-Ogilvie, T. E. (2022). Combined MEK

and JAK/STAT3 pathway inhibition effectively decreases SHH medulloblastoma tumor progression. *Communications Biology*, 5(1). <https://doi.org/10.1038/s42003-022-03654-9>

Chan, Wilson

1. Lam, J. C., **Chan, W. W.**, & Walsh, J. F. (2022). Disseminated nocardiosis in an immunocompetent host with occupational exposure. *IDCases*, 30, e01620. <https://doi.org/10.1016/j.idcr.2022.e01620>

Church, Deirdre

1. *Leber, A. L., Burnham, C.-A. D., & Gregson, D., **Church, D.** (2022). "Aerobic bacteriology" *Clinical microbiology procedures handbook*. Wiley ; ASM Press.
2. *Leber, A. L., Burnham, C.-A. D., & Gregson, D., **Church, D.** (2022). "Epidemiologic and infection control microbiology" *Clinical microbiology procedures handbook*. Wiley ; ASM Press.
3. Bibby, H. L., de Koning, L., Seiden-Long, I., Zelyas, N., **Church, D. L.**, & Berenger, B. M. (2022). A pragmatic randomized controlled trial of rapid on-site influenza and respiratory syncytial virus PCR testing in paediatric and adult populations. *BMC Infectious Diseases*, 22(1). <https://doi.org/10.1186/s12879-022-07796-3>
4. Bourassa-Blanchette, S., Biesheuvel, M. M., Lam, J. C., Kipp, A., **Church, D.**, Carson, J., Dalton, B., Parkins, M. D., Barkema, H. W., & Gregson, D. B. (2023). Incidence, susceptibility and outcomes of candidemia in adults living in Calgary, Alberta, Canada (2010–2018). *BMC Infectious Diseases*, 23(1). <https://doi.org/10.1186/s12879-023-08050-0>
5. *Chow, B. L., Groeschel, M., Carson, J., Griener, T. P., & **Church, D. L.** (2022). Development and evaluation of a novel fast broad-range PCR and sequencing assay (FBR-PCR/S) using dual priming oligonucleotides targeting the ITS/LSU gene regions for rapid diagnosis of invasive fungal diseases: multi-year experience in a large Canadian healthcare zone. *BMC Infectious Diseases*, 22(1). <https://doi.org/10.1186/s12879-022-07356-9>
6. ***Church, D. L.**, Griener, T., & Gregson, D. (2022). Multi-year comparison of VITEK® MS and 16S rRNA gene sequencing performance for the identification of rarely encountered anaerobes causing invasive human infections in a large Canadian region: can our laboratory abandon 16S rRNA gene sequencing? *Anaerobe*, 102640. <https://doi.org/10.1016/j.anaerobe.2022.102640>
7. ***Church, D. L.**, & Naugler, C. (2022). Using a systematic approach to strategic innovation in laboratory medicine to bring about change. *Critical Reviews in Clinical Laboratory Sciences*, 59(3), 178–202. <https://doi.org/10.1080/10408363.2021.1997899>
8. *Gillrie, M., Chow, B., Griener, T., Johnson, A., & **Church, D.** (2023). Hepatosplenic mucormycosis due to *Rhizomucor pusillus* identified by panfungal PCR/sequencing of ribosomal ITS2 and LSU regions in a patient with acute myelogenous leukemia: A case report. *JAMMI*, 8(1), 105–110. <https://doi.org/10.3138/jammi-2022-0020>
9. Izydorczyk, C., Waddell, B. J., Thornton, C. S., Conly, J. M., Rabin, H. R., Somayaji, R., Surette, M. G., **Church, D. L.**, & Parkins, M. D. (2023). *Stenotrophomonas maltophilia* natural history and evolution in the airways of adults with cystic fibrosis. *Frontiers in Microbiology*, 14. <https://doi.org/10.3389/fmicb.2023.1205389>
10. Izydorczyk, C., Waddell, B. J., Weyant, R. B., Surette, M. G., Somayaji, R., Rabin, H. R., Conly, J. M., **Church, D. L.**, & Parkins, M. D. (2022). The natural history and genetic diversity of *Haemophilus influenzae* infecting the airways of adults with cystic fibrosis. *Scientific Reports*, 12(1). <https://doi.org/10.1038/s41598-022-19240-2>
11. *Lang, R., Gill, M. J., Viczko, J., Naugler, C., & **Church, D.** (2022). Risk Factors and Outcomes of Bloodstream Infections Among People With Human Immunodeficiency Virus: A Longitudinal Cohort Study From 2000 to 2017. *PubMed*, 9(8), ofac318–ofac318. <https://doi.org/10.1093/ofid/ofac318>

12. *Lieu, A., Mah, J., Peirano, G., Somayaji, R., & **Church, D. L.** (2022). Microbiological Characterization of *Actinotignum schaalii* Strains Causing Invasive Infections during a Multiyear Period in a Large Canadian Health Care Region. *Microbiology Spectrum*, 10(6). <https://doi.org/10.1128/spectrum.03442-22>
13. *Mah, J., Lee, A., Scott, J. N., & **Church, D.** (2022). Pneumococcal otogenic meningitis complicated by pneumocephalus, seizures, right-sided hemiplegia and cortical venous thrombosis. *IDCases*, 29, e01601. <https://doi.org/10.1016/j.idcr.2022.e01601>
14. *Mah, J., Lieu, A., Somayaji, R., & **Church, D.** (2022). Characterizing *Actinotignum schaalii* infections in a large Canadian healthcare region. *Future Microbiology*, 17(17), 1353–1362. <https://doi.org/10.2217/fmb-2022-0049>
15. Zaidi, S.-Z., Zaheer, R., Barbieri, R., Cook, S. R., Hannon, S. J., Booker, C. W., **Church, D.**, Van Domselaar, G., Zovoilis, A., & McAllister, T. A. (2022). Genomic Characterization of *Enterococcus hirae* From Beef Cattle Feedlots and Associated Environmental Continuum. *Frontiers in Microbiology*, 13. <https://doi.org/10.3389/fmicb.2022.859990>

de Koning, Lawrence

1. Bibby, H. L., **de Koning, L.**, Seiden-Long, I., Zelyas, N., Church, D. L., & Berenger, B. M. (2022). A pragmatic randomized controlled trial of rapid on-site influenza and respiratory syncytial virus PCR testing in paediatric and adult populations. *BMC Infectious Diseases*, 22(1). <https://doi.org/10.1186/s12879-022-07796-3>
2. *Butorin, Y., Seiden-Long, I., & **de Koning, L.** (2024). A verification and implementation plan for 50 blood gas analyzers across a large geographic area. *Clinical Biochemistry*, 123, 110702–110702. <https://doi.org/10.1016/j.clinbiochem.2023.110702>
3. Chan, E. S., **de Koning, L.**, Yu, W., & Chadha, R. (2023). C4d Staining Is Present in Normal Placentas From Pregnancies Prior to Pregnancy Loss Associated With Chronic Histiocytic Intervillositis and Is Reduced by Immunomodulatory Therapy in Subsequent Pregnancies. *Pediatric and Developmental Pathology*, 26(4), 374–387. <https://doi.org/10.1177/10935266231176682>
4. *Chan, E. S., Haider, S., Subramanian, S., Yu, W., Nohr, E. W., & **de Koning, L.** (2022). Congenital cytomegalovirus infection is associated with congenital rickets: a retrospective autopsy cohort study. *Archives of Disease in Childhood: Fetal & Neonatal*, 108(4), 435–436. <https://doi.org/10.1136/archdischild-2022-324760>
5. Chan, E. S., Mehrem, A. A., **de Koning, L.**, Stritzke, A., & Zhou, H. Y. (2023). Extrahepatic biliary atresia in a premature neonate with congenital cytomegalovirus infection. *Pathology*, 55(4), 573–576. <https://doi.org/10.1016/j.pathol.2022.10.013>
6. ***de Koning, L.**, Crawford, S., Nohr, E., Chadha, R., Horn, C., Wright, J. R., & Chan, E. S. (2022). Recurrence risk of villitis of unknown etiology: Analysis of a large retrospective cohort study, systematic review and meta-analysis. *Placenta*, 120, 32–39. <https://doi.org/10.1016/j.placenta.2022.02.006>
7. ***de Koning, L.**, Orton, D., Seiden Long, I., Boyd, J., Kellogg, M., Abdullah, A., Naugler, C., Tsui, A., Strange, B., & Glaser, D. (2023). Distribution of videos demonstrating best practices in preventing hemolysis is associated with reduced hemolysis among nurse-collected specimens in hospitals. *Clinical Biochemistry*, 119, 110632. <https://doi.org/10.1016/j.clinbiochem.2023.110632>
8. Horn, C., Engel, N., Drouin, D., Haley, J., Holder, C., Hung, L., Royall, L., McInnis, P., **de Koning, L.**, & Chan, E. S. (2023). Time Required for Gross Examination of Routine Second and Third Trimester Singleton Placentas by Pathologists' Assistants. *Pediatric and Developmental Pathology*, 26(5), 466–471. <https://doi.org/10.1177/10935266231196015>

9. Wayne Lewis, C., Butorin, Y., **de Koning, L.**, Paul, H. A., Gifford, J. L., Venner, A. A., & Seiden-Long, I. (2023). Low partial pressure of oxygen causes significant and unrecognized under-recovery of glucose on blood gas analyzers. *Clinical Biochemistry*, 111, 60–65. <https://doi.org/10.1016/j.clinbiochem.2022.10.008>
10. *Wright, J. R., & **de Koning, L.** (2023). Eosinophilic/T-cell Chorionic Vasculitis: Its Incidence Is Increasing but It Does Not Appear to Recur in Subsequent Pregnancies. *Pediatric and Developmental Pathology*, 26(3), 281–286. <https://doi.org/10.1177/10935266231156332>
11. *Wright, J. R., Yu, W., & **de Koning, L.** (2022). Author(s)' Reply - Vitelline Vessel Remnant – Derived Funisitis: Semantics, Severity, and Significance. *Pediatric and Developmental Pathology*, 25(5), 572–573. <https://doi.org/10.1177/10935266221113450>
12. *Yang, J., Dong, Y., Naugler, C. T., & **de Koning, L.** (2022). Serum 25-hydroxyvitamin D and risk of cancer in a large community population under investigation for cardiovascular disease: a retrospective cohort study. *BMJ Open*, 12(1), e056543–e056543. <https://doi.org/10.1136/bmjopen-2021-056543>

Dharmani-Khan, Poonam

1. Khanolkar, R. A., Tripathi, G., **Dharmani-Khan, P.**, Dabas, R., Kinzel, M., Kalra, A., Puckrin, R., Jimenez-Zepeda, V., Jamani, K., Duggan, P. R., Chaudhry, A., Bryant, A., Stewart, D. A., Khan, F. M., & Storek, J. (2022). Incomplete chimerism following myeloablative and anti-thymocyte globulin-conditioned hematopoietic cell transplantation is a risk factor for relapse and chronic graft-versus-host disease. *Cytotherapy*, 24(12), 1225–1231. <https://doi.org/10.1016/j.jcyt.2022.07.013>
2. Kinzel, M., Kalra, A., Tripathi, G., **Dharmani-Khan, P.**, Khan, F., & Storek, J. (2022). Lack of both donor and recipient anti-EBV T cells in EBV seronegative recipients of grafts from seropositive donors. *Bone Marrow Transplantation*. <https://doi.org/10.1038/s41409-022-01884-3>
3. Shameli, A., **Dharmani-Khan, P.**, Auer, I., & Shabani-Rad, M.-T. (2023). Deep immunophenotypic analysis of the bone marrow progenitor cells in myelodysplastic syndromes. *Leukemia Research*, 134, 107401–107401. <https://doi.org/10.1016/j.leukres.2023.107401>
4. Tripathi, G., Khanolkar, R. A., Faridi, R. M., Kalra, A., **Dharmani-Khan, P.**, Shabani-Rad, M.-T., Berka, N., Daly, A., Storek, J., & Khan, F. M. (2022). Donor Genetic Predisposition to High Interleukin-10 Production Appears Protective against Acute Graft-Versus-Host Disease. *International Journal of Molecular Sciences*, 23(24), 15888–15888. <https://doi.org/10.3390/ijms232415888>

Dingle, Tanis

1. Chiang, D., **Dingle, T. C.**, Belga, S., Kabbani, D., Bhanji, R. A., Walter, J., Abraldes, J. G., & Cervera, C. (2022). Association between gut colonization of vancomycin-resistant enterococci and liver transplant outcomes. *Transplant Infectious Disease*, 24(3). <https://doi.org/10.1111/tid.13821>
2. de Hoog, S., Walsh, T. J., Ahmed, S. A., Alastruey-Izquierdo, A., Alexander, B. D., Arendrup, M. C., Babady, E., Bai, F.-Y., Balada-Llasat, J.-M., Borman, A., Chowdhary, A., Clark, A., Colgrove, R. C., Cornely, O. A., **Dingle, T. C.**, Dufresne, P. J., Fuller, J., Gangneux, J.-P., Gibas, C., & Glasgow, H. (2023). A conceptual framework for nomenclatural stability and validity of medically important fungi: a proposed global consensus guideline for fungal name changes supported by ABP, ASM, CLSI, ECMM, ESCMID-EFISG, EUCAST-AFST, FDLC, IDSA, ISHAM, MMSA, and MSGERC. *Journal of Clinical Microbiology*, 61(11). <https://doi.org/10.1128/jcm.00873-23>
3. ***Dingle, T. C.**, & Dufresne, P. J. (2022). Discontinuation of DNA Probes for Identification of Dimorphic Fungi Growing in Culture: What's a Lab To Do? *Clinical Microbiology Newsletter*, 44(21), 191–197. <https://doi.org/10.1016/j.clinmicnews.2022.11.001>

4. ***Dingle, T. C.**, Gamage, D., Gomez-Villegas, S., Hanson, B. M., Reyes, J., Abbott, A., Burnham, C.-A. D., Dien Bard, J., Fritz, S., Miller, W. R., Westblade, L. F., Zimmer, B., Arias, C. A., & Butler-Wu, S. (2022). Prevalence and Characterization of the Cefazolin Inoculum Effect in North American Methicillin-Susceptible *Staphylococcus aureus* Isolates. *Journal of Clinical Microbiology*, *60*(7). <https://doi.org/10.1128/jcm.02495-21>
5. ***Dingle, T. C.**, Jansen, B., Walker, C., Sam, M., Verity, B., Purdy, D., Paul, P., & Schwartz, I. S. (2022). Implantation subcutaneous phaeohyphomycosis caused by *Rhytidhysterion rufulum*: A case report. *Medical Mycology Case Reports*, *36*, 16–18. <https://doi.org/10.1016/j.mmcr.2022.03.002>
6. ***Dingle, T. C.**, & Pitout, J. (2022). The Ins and Outs of Susceptibility Testing for New β -Lactam/ β -Lactamase Inhibitor Combinations for Gram-Negative Organisms. *Journal of Clinical Microbiology*, *60*(7). <https://doi.org/10.1128/jcm.00807-21>
7. Humphries, R. M., Ambler, J., Mitchell, S. L., Castanheira, M., **Dingle, T.**, Hindler, J. A., Koeth, L., & Sei, K. (2023). Erratum for Humphries et al., “CLSI Methods Development and Standardization Working Group Best Practices for Evaluation of Antimicrobial Susceptibility Tests.” *Journal of Clinical Microbiology*, *61*(10). <https://doi.org/10.1128/jcm.00739-23>
8. Kariyawasam, R. M., **Dingle, T. C.**, Kula, B. E., Vandermeer, B., Sligl, W. I., & Schwartz, I. S. (2022). Defining COVID-19-associated pulmonary aspergillosis: systematic review and meta-analysis. *Clinical Microbiology and Infection*. <https://doi.org/10.1016/j.cmi.2022.01.027>
9. Kariyawasam, R. M., Julien, D. A., Jelinski, D. C., Larose, S. L., Rennert-May, E., Conly, J. M., **Dingle, T. C.**, Chen, J. Z., Tyrrell, G. J., Ronksley, P. E., & Barkema, H. W. (2022). Antimicrobial resistance (AMR) in COVID-19 patients: a systematic review and meta-analysis (November 2019–June 2021). *Antimicrobial Resistance & Infection Control*, *11*(1). <https://doi.org/10.1186/s13756-022-01085-z>
10. *Ma, A., Ferrato, C., Martin, I., Smyczek, P., Gratrix, J., & **Dingle, T. C.** (2023). Use of genome sequencing to resolve differences in gradient diffusion and agar dilution antimicrobial susceptibility testing performance of *Neisseria gonorrhoeae* isolates in Alberta, Canada. *Journal of Clinical Microbiology*, *61*(11). <https://doi.org/10.1128/jcm.00606-23>
11. *Mataseje, L. F., Pitout, J., Croxen, M., Mulvey, M. R., & **Dingle, T. C.** (2023). Three separate acquisitions of blaNDM-1 in three different bacterial species from a single patient. *European Journal of Clinical Microbiology & Infectious Diseases*, *42*(10), 1275–1280. <https://doi.org/10.1007/s10096-023-04651-4>
12. O’Grady, H. M., Harrison, R., Snedeker, K., Trufen, L., Yue, P., Ward, L., Fifen, A., Jamieson, P., Weiss, A., Coulthard, J., Lynch, T., Croxen, M. A., Li, V., Pabbaraju, K., Wong, A., Zhou, H. Y., **Dingle, T. C.**, Hellmer, K., Berenger, B. M., & Fonseca, K. (2023). A two-ward acute care hospital outbreak of SARS-CoV-2 delta variant including a point-source outbreak associated with the use of a mobile vital signs cart and sub-optimal doffing of personal protective equipment. *Journal of Hospital Infection*, *131*, 1–11. <https://doi.org/10.1016/j.jhin.2022.09.019>
13. Sligl, W. I., Chen, J. Z., Wang, X., Boehm, C., Fong, K., Crick, K., Garrido Clua, M., Codan, C., **Dingle, T. C.**, Gregson, D., Prosser, C., Sadrzadeh, H., Yan, C., Chen, G., Tse-Chang, A., Garros, D., Doig, C. J., Zygun, D., Opgenorth, D., & Conly, J. M. (2023). Antimicrobial stewardship, procalcitonin testing, and rapid blood-culture identification to optimize sepsis care in critically ill adult patients: A quality improvement initiative. *Antimicrobial Stewardship & Healthcare Epidemiology*, *3*(1). <https://doi.org/10.1017/ash.2023.183>
14. Zhang, K., Potter, R. F., Marino, J., Muenks, C. E., Lammers, M. G., Dien Bard, J., **Dingle, T. C.**, Humphries, R. M., Westblade, L. F., Burnham, C. D., & Dantas, G. (2023). Comparative genomics reveals the correlations of stress response genes and bacteriophages in developing antibiotic resistance of *Staphylococcus saprophyticus*. *MSystems*. <https://doi.org/10.1128/msystems.00697-23>

Duggan, Maire

1. Abubakar, M., Klein, A., Fan, S., Lawrence, S., Mutreja, K., Henry, J. E., Pfeiffer, R. M., **Duggan, M. A.**, & Gierach, G. L. (2023). Host, reproductive, and lifestyle factors in relation to quantitative histologic metrics of the normal breast. *Breast Cancer Research*, 25(1). <https://doi.org/10.1186/s13058-023-01692-7>
2. Ahearn, T. U., Choudhury, P. P., Derkach, A., Wiafe-Addai, B., Awuah, B., Yarney, J., Edusei, L., Titiloye, N., Adjei, E., Vanderpuye, V., Aitpillah, F., Dedey, F., Oppong, J., Osei-Bonsu, E. B., **Duggan, M. A.**, Brinton, L. A., Allen, J., Luccarini, C., Baynes, C., & Carvalho, S. (2022). Breast Cancer Risk in Women from Ghana Carrying Rare Germline Pathogenic Mutations. *Cancer Epidemiology, Biomarkers & Prevention*, 31(8), 1593–1601. <https://doi.org/10.1158/1055-9965.epi-21-1397>
3. Felix, A. S., Sinnott, J. A., Cohn, D. E., **Duggan, M. A.**, Havrilesky, L. J., Olawaiye, A. B., Mariani, A., Rodriguez, M., Brett, M. A., Dinoi, G., Meade, C. E., Hall, B., Goldfeld, E., Elishaev, E., Sherman, M. E., & Suarez, A. A. (2023). Intraluminal tumor cells and prognostic accuracy of endometrial cancer stage criteria: A multi-institution study. *Gynecologic Oncology*, 178, 130–137. <https://doi.org/10.1016/j.ygyno.2023.10.004>
4. Park, K. J., Selinger, C. I., Alvarado-Cabrero, I., **Duggan, M. A.**, Kiyokawa, T., Mills, A. M., Ordi, J., Otis, C. N., Plante, M., Stolnicu, S., Talia, K. L., Wiredu, E. K., Lax, S. F., & McCluggage, W. G. (2022). Dataset for the Reporting of Carcinoma of the Cervix: Recommendations From the International Collaboration on Cancer Reporting (ICCR). *International Journal of Gynecological Pathology*, 41, S64. <https://doi.org/10.1097/PGP.0000000000000909>

Franko, Angela

1. Cohen, R. A., Rose, C. S., Go, L. H. T., Zell-Baran, L. M., Almberg, K. S., Sarver, E. A., Lowers, H. A., Iwaniuk, C., Clingerman, S. M., Richardson, D. L., Abraham, J. L., Cool, C. D., **Franko, A. D.**, Hubbs, A. F., Murray, J., Orandle, M. S., Sanyal, S., Vorajee, N. I., Petsonk, E. L., & Zulfikar, R. (2022). Pathology and Mineralogy Demonstrate Respirable Crystalline Silica is a Major Cause of Severe Pneumoconiosis in US Coal Miners. *Annals of the American Thoracic Society*. <https://doi.org/10.1513/annalsats.202109-1064oc>
2. Go, L. H. T., Rose, C. S., Zell-Baran, L. M., Almberg, K. S., Iwaniuk, C., Clingerman, S., Richardson, D. L., Abraham, J. L., Cool, C. D., **Franko, A. D.**, Green, F. H. Y., Hubbs, A. F., Murray, J., Orandle, M. S., Sanyal, S., Vorajee, N. I., Sarver, E. A., Petsonk, E. L., & Cohen, R. A. (2023). Historical shift in pathological type of progressive massive fibrosis among coal miners in the USA. *Occupational and Environmental Medicine*, 80(8), 425–430. <https://doi.org/10.1136/oemed-2022-108643>
3. Lach, K., Gharpuray-Pandit, D., **Franko, A.**, & Brenn, T. (2023). Distant metastasis form morphologically low-grade spiradenocarcinoma: a report of two cases. *Journal of Cutaneous Pathology*. <https://doi.org/10.1111/cup.14388>
4. Thornton, C. S., Huntley, K., Berenger, B. M., Bristow, M., Evans, D. H., Fonseca, K., **Franko, A.**, Gillrie, M. R., Lin, Y.-C., Povitz, M., Shafey, M., Conly, J. M., & Tremblay, A. (2022). Prolonged SARS-CoV-2 infection following rituximab treatment: clinical course and response to therapeutic interventions correlated with quantitative viral cultures and cycle threshold values. *Antimicrobial Resistance & Infection Control*, 11(1). <https://doi.org/10.1186/s13756-022-01067-1>
5. Zell-Baran, L. M., Go, L. H. T., Sarver, E., Almberg, K. S., Iwaniuk, C., Green, F. H. Y., Abraham, J. L., Cool, C., **Franko, A.**, Hubbs, A. F., Murray, J., Orandle, M. S., Sanyal, S., Vorajee, N., Cohen, R. A., & Rose, C. S. (2022). Mining Tenure and Job Duties Differ Among Contemporary and Historic Underground Coal Miners With Progressive Massive Fibrosis. *Journal of Occupational & Environmental Medicine*, 65(4), 315–320. <https://doi.org/10.1097/jom.0000000000002746>

Gifford, Jessica

1. ***Gifford, J. L.**, & Seiden-Long, I. (2022). Filling in the GAPS: validation of anion gap (AGAP) measurement uncertainty estimates for use in clinical decision making. *Clinical Chemistry and Laboratory Medicine*, 60(6), 851–858. <https://doi.org/10.1515/cclm-2021-1279>
2. Higgins, V., Garcia, L., **Gifford, J. L.**, Volodko, N., Beriault, D. R., Parker, M. L., Estey, M. P., Proctor, D. T., & Ismail, O. Z. (2023). Validating the NIH LDL-C equation for provincial implementation in Alberta. *Clinical Biochemistry*, 121-122, 110678–110678. <https://doi.org/10.1016/j.clinbiochem.2023.110678>
3. Paul, H. A., Chi, Q., **Gifford, J. L.**, & Seiden-Long, I. (2022). Not T too! False elevations in high-sensitivity cardiac troponin T (hs-TnT) following specimen transport. *Clinical Biochemistry*. <https://doi.org/10.1016/j.clinbiochem.2022.08.008>
4. Wayne Lewis, C., Butorin, Y., de Koning, L., Paul, H. A., **Gifford, J. L.**, Venner, A. A., & Seiden-Long, I. (2023). Low partial pressure of oxygen causes significant and unrecognized under-recovery of glucose on blood gas analyzers. *Clinical Biochemistry*, 111, 60–65. <https://doi.org/10.1016/j.clinbiochem.2022.10.008>

Gregson, Daniel

1. *Leber, A. L., Burnham, C.-A. D., & **Gregson, D.** (2022). “Epidemiologic and infection control microbiology” Clinical microbiology procedures handbook. Wiley ; ASM Press.
2. *Bourassa-Blanchette, S., Biesheuvel, M. M., Lam, J. C., Kipp, A., Church, D., Carson, J., Dalton, B., Parkins, M. D., Barkema, H. W., & **Gregson, D. B.** (2023). Incidence, susceptibility and outcomes of candidemia in adults living in Calgary, Alberta, Canada (2010–2018). *BMC Infectious Diseases*, 23(1). <https://doi.org/10.1186/s12879-023-08050-0>
3. *Church, D. L., Griener, T., & **Gregson, D.** (2022). Multi-year comparison of VITEK® MS and 16S rRNA gene sequencing performance for the identification of rarely encountered anaerobes causing invasive human infections in a large Canadian region: can our laboratory abandon 16S rRNA gene sequencing? *Anaerobe*, 102640. <https://doi.org/10.1016/j.anaerobe.2022.102640>
4. Groves, R. A., Chan, C. C. Y., Wildman, S. D., **Gregson, D. B.**, Rydzak, T., & Lewis, I. A. (2022). Rapid LC–MS assay for targeted metabolite quantification by serial injection into isocratic gradients. *Analytical and Bioanalytical Chemistry*, 415(2), 269–276. <https://doi.org/10.1007/s00216-022-04384-x>
5. Groves, R. A., Mapar, M., Aburashed, R., Ponce, L. F., Bishop, S. L., Rydzak, T., Drikkic, M., Bihan, D. G., Benediktsson, H., Clement, F., **Gregson, D. B.**, & Lewis, I. A. (2022). Methods for Quantifying the Metabolic Boundary Fluxes of Cell Cultures in Large Cohorts by High-Resolution Hydrophilic Liquid Chromatography Mass Spectrometry. *Analytical Chemistry*, 94(25), 8874–8882. <https://doi.org/10.1021/acs.analchem.2c00078>
6. Henderson, A., Cheng, M. P., Chew, K. L., Coombs, G. W., Davis, J. S., Grant, J. M., **Gregson, D.**, Giulieri, S. G., Howden, B. P., Lee, T. C., Nguyen, V., Mora, J. M., Morpeth, S. C., Robinson, J. O., Tong, S. Y. C., Van Hal, S. J., & Microbiology Working Group of the Staphylococcus aureus Network Adaptive Platform (SNAP) Trial Group. (2023). A multi-site, international laboratory study to assess the performance of penicillin susceptibility testing of Staphylococcus aureus. *The Journal of Antimicrobial Chemotherapy*, 78(6), 1499–1504. <https://doi.org/10.1093/jac/dkad116>
7. Kanji, J. N., Isaac, A., **Gregson, D.**, Mierzejewski, M., Shpeley, D., Tomlin, P., Groeschel, M., Lindsay, L. R., Lachance, L., & Kowalewska-Grochowska, K. (2022). Epidemiology of ticks submitted from human hosts in Alberta, Canada (2000-2019). *Emerging Microbes & Infections*, 11(1), 284–292. <https://doi.org/10.1080/22221751.2022.2027217>
8. Mossman, A. K., Svishchuk, J., Waddell, B., Izydorczyk, C., Buckley, P. T., Hilliard, J. J., Al-Ghalith, G. A., Zheng, L., Lynch, A. S., Mody, C. H., Lisboa, L. F., **Gregson, D. B.**, & Parkins, M. D. (2022).

- Staphylococcus aureus* in Non-Cystic Fibrosis Bronchiectasis: Prevalence and Genomic Basis of High Inoculum β -Lactam Resistance. *Annals of the American Thoracic Society*, 19(8), 1285–1293. <https://doi.org/10.1513/annalsats.202108-965oc>
9. Sligl, W. I., Chen, J. Z., Wang, X., Boehm, C., Fong, K., Crick, K., Garrido Clua, M., Codan, C., Dingle, T. C., **Gregson, D.**, Prosser, C., Sadrzadeh, H., Yan, C., Chen, G., Tse-Chang, A., Garros, D., Doig, C. J., Zygun, D., Opgenorth, D., & Conly, J. M. (2023). Antimicrobial stewardship, procalcitonin testing, and rapid blood-culture identification to optimize sepsis care in critically ill adult patients: A quality improvement initiative. *Antimicrobial Stewardship & Healthcare Epidemiology*, 3(1). <https://doi.org/10.1017/ash.2023.183>
 10. Sohani, Z. N., Butler-Laporte, G., Aw, A., Belga, S., Benedetti, A., Carignan, A., Cheng, M. P., Coburn, B., Costiniuk, C. T., Ezer, N., **Gregson, D.**, Johnson, A., Khwaja, K., Lawandi, A., Leung, V., Lothar, S., MacFadden, D., McGuinty, M., Parkes, L., & Qureshi, S. (2022). Low-dose trimethoprim-sulfamethoxazole for the treatment of *Pneumocystis jirovecii* pneumonia (LOW-TMP): protocol for a phase III randomised, placebo-controlled, dose-comparison trial. *BMJ Open*, 12(7), e053039. <https://doi.org/10.1136/bmjopen-2021-053039>
 11. Svishchuk, J., Ebbert, K., Waddell, B., Izydorczyk, C., Acosta, N., Somayaji, R., Rabin, H. R., Bjornson, C. L., Lisboa, L., **Gregson, D. B.**, Conly, J. M., Surette, M. G., & Parkins, M. D. (2023). Epidemiology and impact of methicillin-sensitive *Staphylococcus aureus* with β -lactam antibiotic inoculum effects in adults with cystic fibrosis. *Antimicrobial Agents and Chemotherapy*, 67(12). <https://doi.org/10.1128/aac.00136-23>

Griener, Thomas

1. Chow, B. L., Groeschel, M., Carson, J., **Griener, T. P.**, & Church, D. L. (2022). Development and evaluation of a novel fast broad-range PCR and sequencing assay (FBR-PCR/S) using dual priming oligonucleotides targeting the ITS/LSU gene regions for rapid diagnosis of invasive fungal diseases: multi-year experience in a large Canadian healthcare zone. *BMC Infectious Diseases*, 22(1). <https://doi.org/10.1186/s12879-022-07356-9>
2. Church, D. L., **Griener, T.**, & Gregson, D. (2022). Multi-year comparison of VITEK® MS and 16S rRNA gene sequencing performance for the identification of rarely encountered anaerobes causing invasive human infections in a large Canadian region: can our laboratory abandon 16S rRNA gene sequencing? *Anaerobe*, 102640. <https://doi.org/10.1016/j.anaerobe.2022.102640>
3. Gillrie, M., Chow, B., **Griener, T.**, Johnson, A., & Church, D. (2023). Hepatosplenic mucormycosis due to *Rhizomucor pusillus* identified by panfungal PCR/sequencing of ribosomal ITS2 and LSU regions in a patient with acute myelogenous leukemia: A case report. *JAMMI*, 8(1), 105–110. <https://doi.org/10.3138/jammi-2022-0020>

Groeschel, Michael

1. Chow, B. L., **Groeschel, M.**, Carson, J., Griener, T. P., & Church, D. L. (2022). Development and evaluation of a novel fast broad-range PCR and sequencing assay (FBR-PCR/S) using dual priming oligonucleotides targeting the ITS/LSU gene regions for rapid diagnosis of invasive fungal diseases: multi-year experience in a large Canadian healthcare zone. *BMC Infectious Diseases*, 22(1). <https://doi.org/10.1186/s12879-022-07356-9>
2. Kanji, J. N., Isaac, A., Gregson, D., Mierzejewski, M., Shpeley, D., Tomlin, P., **Groeschel, M.**, Lindsay, L. R., Lachance, L., & Kowalewska-Grochowska, K. (2022). Epidemiology of ticks submitted from human hosts in Alberta, Canada (2000-2019). *Emerging Microbes & Infections*, 11(1), 284–292. <https://doi.org/10.1080/22221751.2022.2027217>

Hunter, Charlene

1. Green, R., **Hunter, C.**, & Chia, J. C. (2022). A case of inflammatory linear verrucous epidermal nevus successfully treated with Brodalumab. *Pediatric Dermatology*. <https://doi.org/10.1111/pde.15058>

Hycza, Martin

2. Arora, R., Cao, C., Kumar, M., Sinha, S., Chanda, A., McNeil, R., Samuel, D., Arora, R. K., Matthew, T. W., Chandarana, S., Hart, R. D., Dort, J. C., Biernaskie, J., Neri, P., **Hycza, M.**, & Bose, P. (2022). Spatial transcriptomics reveals distinct and conserved tumor core and edge architectures that predict survival and targeted therapy response. *BioRxiv (Cold Spring Harbor Laboratory)*. <https://doi.org/10.1101/2022.09.04.505581>
3. Baněčková, M., Thompson, L. D. R., **Hycza, M. D.**, Vaněček, T., Agaimy, A., Laco, J., Simpson, R. H. W., Di Palma, S., Stevens, T. M., Brcic, L., Etebarian, A., Dimnik, K., Majewska, H., Stárek, I., O'Regan, E., Salviato, T., Helliwell, T., Horáková, M., Biernat, W., & Onyuma, T. (2023). Salivary Gland Secretory Carcinoma. *The American Journal of Surgical Pathology*, 47(6), 661–677. <https://doi.org/10.1097/pas.0000000000002043>
4. Bradová, M., Lester, **Hycza, M.**, Vaněček, T., Grossman, P., Michal, M., Hájková, V., Taheri, T., Rupp, N., Suster, D., Lakhani, S., Nikolov, D. H., Žalud, R., Skálová, A., Michal, M., & Agaimy, A. (2023). Branchioma: immunohistochemical and molecular genetic study of 23 cases highlighting frequent loss of retinoblastoma 1 immunoexpression. *Virchows Archiv*. <https://doi.org/10.1007/s00428-023-03697-1>
5. Ernst, M. S., Lysack, J. T., **Hycza, M. D.**, Chandarana, S. P., & Hao, D. (2022). TRK Inhibition with Entrectinib in Metastatic Salivary Secretory Carcinoma (SC): A Case Report. *Current Oncology*, 29(6), 3933–3939. <https://doi.org/10.3390/curroncol29060314>
6. Eszlinger, M., Stewardson, P., McIntyre, J. B., Box, A., Khalil, M., **Hycza, M.**, Koro, K., Ruether, D., Wu, J., & Paschke, R. (2022). Systematic population-based identification of NTRK and RET fusion-positive thyroid cancers. *European Thyroid Journal*, 11(1). <https://doi.org/10.1530/etj-21-0061>
7. Grace, S., **Hycza, M. D.**, & Monzon, J. G. (2023). Treatment of Unresectable Cutaneous Squamous Cell Carcinoma with Cemiplimab in a Patient on Dialysis. *Case Reports in Oncology*, 143–148. <https://doi.org/10.1159/000528414>
8. ***Hycza, M. D.**, Lindenmuth, T. R., & Auerbach, A. (2023). Top Ten Lymphoproliferative Lesions Not to Miss When Evaluating Oral Ulcer Biopsies. *Head and Neck Pathology*, 17(1), 99–118. <https://doi.org/10.1007/s12105-023-01532-2>
9. ***Hycza, M. D.**, Martins-Filho, S. N., Spatz, A., Wang, H.-J., Purgina, B. M., Desmeules, P., Park, P. C., Bigras, G., Jung, S., Cutz, J.-C., Xu, Z., Berman, D. M., Sheffield, B. S., Cheung, C. C., Leduc, C., Hwang, D. M., Ionescu, D., Klonowski, P., Chevarie-Davis, M., & Chami, R. (2024). Canadian Multicentric Pan-TRK (CANTRK) Immunohistochemistry Harmonization Study. *Modern Pathology*, 37(1), 100384–100384. <https://doi.org/10.1016/j.modpat.2023.100384>
10. Koonar, E., Ramazani, F., **Hycza, M.**, & Chau, J. (2023). Temporal bone manifestation of primary extranodal Rosai–Dorfman disease: a case report. *Journal of Medical Case Reports*, 17(1). <https://doi.org/10.1186/s13256-023-03790-8>
11. Michal, M., Skálová, A., **Hycza, M.**, Laco, J., Vaněček, T., Rupp, N. J., Michal, M., Michalová, K., Agaimy, A., & Bradová, M. (2023). Nasal and sinonasal tumors formed by atypical adenomatous lesions arising in respiratory epithelial adenomatoid hamartoma/seromucinous hamartoma. *Virchows Archiv*. <https://doi.org/10.1007/s00428-023-03719-y>
12. Mudhar, H. S., Krishna, Y., Cross, S., Auw-Haedrich, C., Barnhill, R., Cherepanoff, S., Eagle, R., Farmer, J., Folberg, R., Grossniklaus, H., Herwig-Carl, M. C., **Hycza, M.**, Lassalle, S., Loeffler, K. U., Moulin, A., Milman, T., Verdijk, R. M., Heegaard, S., & Coupland, S. E. (2024). A Multicenter

Study Validates the WHO 2022 Classification for Conjunctival Melanocytic Intraepithelial Lesions With Clinical and Prognostic Relevance. *Laboratory Investigation*, 104(1), 100281–100281. <https://doi.org/10.1016/j.labinv.2023.100281>

13. *Skalova, A., & **Hycza, M. D.** (2023). Proceedings of the North American Society of Head and Neck Pathology Companion Meeting, New Orleans, LA, March 12, 2023: Classification of Salivary Gland Tumors: Remaining Controversial Issues? *Head and Neck Pathology*, 17(2), 285–291. <https://doi.org/10.1007/s12105-023-01541-1>
14. Skálová, A., **Hycza, M. D.**, & Leivo, I. (2022). Update from the 5th Edition of the World Health Organization Classification of Head and Neck Tumors: Salivary Glands. *Head and Neck Pathology*, 16(1), 40–53. <https://doi.org/10.1007/s12105-022-01420-1>
15. Thierauf, J. C., Farahani, A. A., Indave, B. I., Bard, A. Z., White, V. A., Smith, C. R., Marble, H., **Hycza, M. D.**, John, Bishop, J., Shi, Q., Ely, K., Agaimy, A., Martinez-Lage, M., Nose, V., Rivera, M., Nardi, V., Dias-Santagata, D., Garg, S., & Sadow, P. (2022). Diagnostic Value of MAML2 Rearrangements in Mucoepidermoid Carcinoma. *International Journal of Molecular Sciences*, 23(8), 4322–4322. <https://doi.org/10.3390/ijms23084322>
16. Weis, E., Surgeoner, B., Salopek, T. G., Cheng, T., **Hycza, M.**, Kostaras, X., Larocque, M., McKinnon, G., McWhae, J., Menon, G., Monzon, J., Murtha, A. D., Walker, J., & Temple-Oberle, C. (2023). Management of Uveal Melanoma: Updated Cancer Care Alberta Clinical Practice Guideline. *Current Oncology*, 31(1), 24–41. <https://doi.org/10.3390/curroncol31010002>
17. White, V. A., **Hycza, M. D.**, Lennerz, J. K., Thierauf, J., Lokuhetty, D., Cree, I. A., & Indave, B. I. (2022). Mucoepidermoid carcinoma (MEC) and adenosquamous carcinoma (ASC), the same or different entities? *Modern Pathology*, 35(10), 1484–1493. <https://doi.org/10.1038/s41379-022-01100-z>
18. Wu, J., Hu, X. Y., Ghaznavi, S., Kinnear, S., Symonds, C. J., Grundy, P., Parkins, V. M., Sharma, P., Lamb, D., Khalil, M., **Hycza, M.**, Chandarana, S. P., Pasieka, J. L., Harvey, A., Warshawski, J., Hart, R., Deutschman, M., Randall, D. R., & Paschke, R. (2022). The Prospective Implementation of the 2015 ATA Guidelines and Modified ATA Recurrence Risk Stratification System for Treatment of Differentiated Thyroid Cancer in a Canadian Tertiary Care Referral Setting. *Thyroid*, 32(12), 1509–1518. <https://doi.org/10.1089/thy.2022.0055>

Joseph, Jeffrey

1. Ambeskovic, M., Hopkins, G., Hoover, T., **Joseph, J. T.**, Montina, T., & Metz, S. (2023). Metabolomic Signatures of Alzheimer's Disease Indicate Brain Region-Specific Neurodegenerative Progression. *International Journal of Molecular Sciences*, 24(19), 14769–14769. <https://doi.org/10.3390/ijms241914769>
2. *Newell, C., Kline, G. A., & **Joseph, J. T.** (2022). The Curious Case of Hypopituitarism. *Journal of Neuropathology & Experimental Neurology*. <https://doi.org/10.1093/jnen/nlac046>
3. Perera, T., Khayambashi, S., Jewett, G., Hahn, C., McCulloch, S., **Joseph, J. T.**, & Chhibber, S. (2023). Plasmapheresis for Treatment of Light Chain Amyloidosis Related Myopathy. *Canadian Journal of Neurological Sciences*, 1–3. <https://doi.org/10.1017/cjn.2023.38>
4. Roczkowsky, A., Limonta, D., Fernandes, J. P., Branton, W. G., Clarke, M., Hlavay, B., Noyce, R. S., **Joseph, J. T.**, Ogando, N. S., Das, S. K., Elaihs, M., Arbour, N., Evans, D. H., Langdon, K., Hobman, T. C., & Power, C. (2023). COVID-19 Induces Neuroinflammation and Suppresses Peroxisomes in the Brain. *Annals of Neurology*, 94(3), 531–546. <https://doi.org/10.1002/ana.26679>
5. Stepanchuk, A. A., Morgan, M. L., **Joseph, J. T.**, & Stys, P. K. (2022). Dual-probe fluorescence spectroscopy for sensitive quantitation of Alzheimer's amyloid pathology. *Acta Neuropathologica Communications*, 10(1). <https://doi.org/10.1186/s40478-022-01456-y>

Kelly, Margaret

1. Fortin, M., Liberman, M., Delage, A., Dion, G., Martel, S., Rolland, F., Soumagne, T., Trahan, S., Assayag, D., Albert, E., **Kelly, M. M.**, Johannson, K. A., Guenther, Z., Leduc, C., Manganas, H., Prenovault, J., & Provencher, S. (2023). Transbronchial Lung Cryobiopsy and Surgical Lung Biopsy: A Prospective Multi-Centre Agreement Clinical Trial (CAN-ICE). *American Journal of Respiratory and Critical Care Medicine*, 207(12), 1612–1619. <https://doi.org/10.1164/rccm.202209-1812oc>
2. Vargas, F., Nguyen, R., Willson, M., Davoli-Ferreira, M., David, B. A., **Kelly, M. M.**, Lee, W.-Y., Kratofil, R. M., Zhang, W. X., Bui-Marinos, M. P., Corcoran, J. A., & Kubes, P. (2023). Intravital imaging of three different microvascular beds in SARS-CoV-2-infected mice. *Blood Advances*, 7(15), 4170–4181. <https://doi.org/10.1182/bloodadvances.2022009430>

Khalil, Moosa

1. Craig, S., Stretch, C., Farshidfar, F., Sheka, D., Alabi, N., Siddiqui, A., Kopciuk, K., Park, Y. J., **Khalil, M.**, Khan, F., Harvey, A., & Bathe, O. F. (2023). A clinically useful and biologically informative genomic classifier for papillary thyroid cancer. *Frontiers in Endocrinology*, 14. <https://doi.org/10.3389/fendo.2023.1220617>
2. Eszlinger, M., Stephenson, A., Mirhadi, S., Patyra, K., Moran, M. F., **Khalil, M.**, Kero, J., & Paschke, R. (2023). Activation of mitogen-activated protein kinase signaling and development of papillary thyroid carcinoma in thyroid-stimulating hormone receptor D633H knockin mice. *European Thyroid Journal*, 12(6). <https://doi.org/10.1530/etj-23-0049>
3. Eszlinger, M., Stewardson, P., McIntyre, J. B., Box, A., **Khalil, M.**, Hyrcza, M., Koro, K., Ruether, D., Wu, J., & Paschke, R. (2022). Systematic population-based identification of NTRK and RET fusion-positive thyroid cancers. *European Thyroid Journal*, 11(1). <https://doi.org/10.1530/etj-21-0061>
4. Ghaznavi, S. A., Clayton, H., Eszlinger, M., **Khalil, M.**, Symonds, C. J., & Paschke, R. (2022). Accuracy of Thyroid Fine-Needle Aspiration Cytology: A Cyto-Histologic Correlation Study in an Integrated Canadian Health Care Region with Centralized Pathology Service. *Acta Cytologica*, 66(3), 171–178. <https://doi.org/10.1159/000521562>
5. Stewardson, P., Eszlinger, M., Wu, J., **Khalil, M.**, Box, A., Perizzolo, M., Punjwani, Z., Ziehr, B., Sanyal, R., Demetrick, D. J., & Paschke, R. (2023). Prospective Validation of ThyroSPEC Molecular Testing of Indeterminate Thyroid Nodule Cytology Following Diagnostic Pathway Optimization. *Thyroid*, 33(12), 1423–1433. <https://doi.org/10.1089/thy.2023.0255>
6. Wu, J., Hu, X. Y., Ghaznavi, S., Kinnear, S., Symonds, C. J., Grundy, P., Parkins, V. M., Sharma, P., Lamb, D., **Khalil, M.**, Hyrcza, M., Chandarana, S. P., Pasiaka, J. L., Harvey, A., Warshawski, J., Hart, R., Deutschman, M., Randall, D. R., & Paschke, R. (2022). The Prospective Implementation of the 2015 ATA Guidelines and Modified ATA Recurrence Risk Stratification System for Treatment of Differentiated Thyroid Cancer in a Canadian Tertiary Care Referral Setting. *Thyroid*, 32(12), 1509–1518. <https://doi.org/10.1089/thy.2022.0055>
7. *Zhao, L., Cheng, T., & **Khalil, M.** (2023). Upper gastrointestinal bleed as the initial presentation of seminoma involving the duodenum in an elderly patient with 9 year follow up: A case report and literature review. *Diagnostic Cytopathology*. <https://doi.org/10.1002/dc.25119>

Khan, Faisal

1. Abu-Khader, A., Hu, Q., Kamar, F., Galaszkiwicz, I., Wang, W., **Khan, F.**, & Berka, N. (2023). Low incidence of de novo HLA antibodies after COVID-19 vaccination: A cohort study of patients awaiting kidney transplantation. *Transplant Infectious Disease*, 25(6). <https://doi.org/10.1111/tid.14105>

2. Abu-Khader, A., Gareau, A. J., Burns, C., **Khan, F.**, & Berka, N. (2022). Next-generation sequencing identifies two novel HLA class II alleles, *HLA-DRB1*01:115* and *HLA-DRB1*14:224*. *HLA*, *100*(2), 176–177. <https://doi.org/10.1111/tan.14623>
3. Craig, S., Stretch, C., Farshidfar, F., Sheka, D., Alabi, N., Siddiqui, A., Kopciuk, K., Park, Y. J., Khalil, M., **Khan, F.**, Harvey, A., & Bathe, O. F. (2023). A clinically useful and biologically informative genomic classifier for papillary thyroid cancer. *Frontiers in Endocrinology*, *14*. <https://doi.org/10.3389/fendo.2023.1220617>
4. Khanolkar, R. A., Tripathi, G., Dharmani-Khan, P., Dabas, R., Kinzel, M., Kalra, A., Puckrin, R., Jimenez-Zepeda, V., Jamani, K., Duggan, P. R., Chaudhry, A., Bryant, A., Stewart, D. A., **Khan, F. M.**, & Storek, J. (2022). Incomplete chimerism following myeloablative and anti-thymocyte globulin-conditioned hematopoietic cell transplantation is a risk factor for relapse and chronic graft-versus-host disease. *Cytotherapy*, *24*(12), 1225–1231. <https://doi.org/10.1016/j.jcyt.2022.07.013>
5. Kinzel, M., Kalra, A., Khanolkar, R. A., Williamson, T. S., Li, N., **Khan, F.**, Puckrin, R., Duggan, P. R., Shafey, M., & Storek, J. (2023). Rituximab Toxicity after Preemptive or Therapeutic Administration for Post-Transplant Lymphoproliferative Disorder. *Transplantation and Cellular Therapy*, *29*(1), 43.e1–43.e8. <https://doi.org/10.1016/j.jtct.2022.10.013>
6. Kinzel, M., Kalra, A., Tripathi, G., Dharmani-Khan, P., **Khan, F.**, & Storek, J. (2022). Lack of both donor and recipient anti-EBV T cells in EBV seronegative recipients of grafts from seropositive donors. *Bone Marrow Transplantation*. <https://doi.org/10.1038/s41409-022-01884-3>
7. Lewis, J., Greenway, S. C., **Khan, F.**, Singh, G., Bhatia, M., & Guilcher, G. M. T. (2022). Assessment of donor cell engraftment after hematopoietic stem cell transplantation for sickle cell disease: A review of current and future methods. *American Journal of Hematology*, *97*(10), 1359–1371. <https://doi.org/10.1002/ajh.26599>
8. Tripathi, G., Khanolkar, R. A., Faridi, R. M., Kalra, A., Dharmani-Khan, P., Shabani-Rad, M.-T., Berka, N., Daly, A., Storek, J., & **Khan, F. M.** (2022). Donor Genetic Predisposition to High Interleukin-10 Production Appears Protective against Acute Graft-Versus-Host Disease. *International Journal of Molecular Sciences*, *23*(24), 15888–15888. <https://doi.org/10.3390/ijms232415888>

Klonowski, Paul

1. Hyrcza, M. D., Martins-Filho, S. N., Spatz, A., Wang, H.-J., Purgina, B. M., Desmeules, P., Park, P. C., Bigras, G., Jung, S., Cutz, J.-C., Xu, Z., Berman, D. M., Sheffield, B. S., Cheung, C. C., Leduc, C., Hwang, D. M., Ionescu, D., **Klonowski, P.**, Chevarie-Davis, M., & Chami, R. (2024). Canadian Multicentric Pan-TRK (CANTRK) Immunohistochemistry Harmonization Study. *Modern Pathology*, *37*(1), 100384–100384. <https://doi.org/10.1016/j.modpat.2023.100384>

Koebel, Martin

1. Bolton, K. L., Chen, D., Corona, R. I., Fu, Z., Murali, R., Bel, M. K., Tazi, Y., Cunningham, J. M., Chan, I. C. C., Wiley, B. J., Moukarzel, L. A., Winham, S. J., Armasu, S. M., Lester, J., Elishaev, E., Laslavic, A., Kennedy, C. J., Piskorz, A., Sekowska, M., & Brand, A. H. (2022). Molecular Subclasses of Clear Cell Ovarian Carcinoma and Their Impact on Disease Behavior and Outcomes. *Clinical Cancer Research*, *28*(22), 4947–4956. <https://doi.org/10.1158/1078-0432.ccr-21-3817>
2. Cheasley, D., Fernandez, M. L., **Köbel, M.**, Kim, H., Dawson, A., Hoenisch, J., Bittner, M., Chiu, D. S., Talhouk, A., Gilks, C. B., Jayawardana, M. W., Pishas, K. I., Mes-Masson, A.-M., Provencher, D., Nigam, A., Hacker, N. F., Gorringer, K. L., Campbell, I. G., & Carey, M. S. (2022). Molecular characterization of low-grade serous ovarian carcinoma identifies genomic aberrations

- according to hormone receptor expression. *Npj Precision Oncology*, 6(1).
<https://doi.org/10.1038/s41698-022-00288-2>
3. *Eun Young Kang, Terzic, T., Ghatage, P., Woo, L., Gorringer, K. L., Karnezis, A. N., Lee, C. H., & **Köbel, M.** (2023). TP53 as a Diagnostic Aid in the Distinction of Ovarian Mucinous Borderline Tumors From Mucinous Carcinoma. *International Journal of Gynecological Pathology*.
<https://doi.org/10.1097/pgp.0000000000000967>
 4. Farahani, H., Boschman, J., Farnell, D., Darbandsari, A., Zhang, A., Ahmadvand, P., Jones, S. J. M., Huntsman, D., **Köbel, M.**, Gilks, C. B., Singh, N., & Bashashati, A. (2022). Deep learning-based histotype diagnosis of ovarian carcinoma whole-slide pathology images. *Modern Pathology*, 35(12), 1983–1990. <https://doi.org/10.1038/s41379-022-01146-z>
 5. Gilks, C. B., Selinger, C., Davidson, B., **Köbel, M.**, Ledermann, J. A., Lim, D., Malpica, A., Mikami, Y., Singh, N., Srinivasan, R., Vang, R., Lax, S., & McCluggage, W. G. (2022). Data Set for the Reporting of Ovarian, Fallopian Tube and Primary Peritoneal Carcinoma: Recommendations From the International Collaboration on Cancer Reporting (ICCR). *International Journal of Gynecological Pathology*, 41(Supplement 1), S119–S142. <https://doi.org/10.1097/pgp.0000000000000908>
 6. Hamilton, S. N., Tinker, A. V., Kwon, J., Lim, P., Kong, I., Sihra, S., **Koebel, M.**, & Lee, C. H. (2022). Treatment and outcomes in undifferentiated and dedifferentiated endometrial carcinoma. *Journal of Gynecologic Oncology*, 33(3). <https://doi.org/10.3802/jgo.2022.33.e25>
 7. Ji, J. X., Cochrane, D., Gian Luca Negri, Colborne, S., Spencer, S. E., Hoang, L. N., Farnell, D., Tessier-Cloutier, B., Huvila, J., Thompson, E., Leung, S., Chiu, D., Chow, C., Ta, M., **Köbel, M.**, Feil, L., Anglesio, M., Goode, E., Bolton, K., & Morin, G. B. (2022). The proteome of clear cell ovarian carcinoma. *The Journal of Pathology*, 258(4), 325–338. <https://doi.org/10.1002/path.6006>
 8. Jiang, W. N., Chan, A. M. Y., Ugoiti, G. R., Kornaga, E., Mathen, P., Yeung, R., Konno, M., Lee, S., Box, A., **Köbel, M.**, Joseph, K. J., & Doll, C. M. (2018). The Prognostic Impact of PD-L1 and CD8 Expression in Anal Cancer Patients Treated with Chemoradiotherapy. *International Journal of Radiation Oncology Biology Physics*, 102(3), S62–S63.
<https://doi.org/10.1016/j.ijrobp.2018.06.175>
 9. ***Köbel, M.**, & Kang, E. Y. (2022). The Evolution of Ovarian Carcinoma Subclassification. *Cancers*, 14(2), 416. <https://doi.org/10.3390/cancers14020416>
 10. ***Köbel, M.**, Rui Zhe Yang, Eun Young Kang, Al-Shamma, Z., Cook, L. S., Kinloch, M., Carey, M. S., Hopkins, L., Nelson, G. S., McManus, K. J., Vizeacoumar, F. S., Vizeacoumar, F. J., Freywald, A., Fu, Y., Reuss, D. E., & Lee, C.-H. (2023). Survey of NF1 inactivation by surrogate immunohistochemistry in ovarian carcinomas. *Gynecologic Oncology*, 178, 80–88.
<https://doi.org/10.1016/j.ygyno.2023.09.016>
 11. ***Koebel, M.**, & Clarke, B. A. (2022). Perspectives in Endometrial Pathology. *AJSP: Reviews & Reports*, 27(5), 177–178. <https://doi.org/10.1097/pcr.0000000000000530>
 12. Kommos, F. K. F., Chiang, S., **Köbel, M.**, Koelsche, C., Chang, K. T.-E., Irving, J. A., Dickson, B., Thiryayi, S., Rouzbahman, M., Rasty, G., von Deimling, A., Lee, C.-H., & Turashvili, G. (2022). Endometrial Stromal Sarcomas With BCOR Internal Tandem Duplication and Variant BCOR/BCORL1 Rearrangements Resemble High-grade Endometrial Stromal Sarcomas With Recurrent CDK4 Pathway Alterations and MDM2 Amplifications. *American Journal of Surgical Pathology*, 46(8), 1142–1152. <https://doi.org/10.1097/pas.0000000000001909>
 13. Kommos, F. K., Tessier-Cloutier, B., Witkowski, L., Forgo, E., Koelsche, C., **Köbel, M.**, Foulkes, W. D., Lee, C.-H., Kolin, D. L., von Deimling, A., & Howitt, B. E. (2022). Cellular context determines DNA methylation profiles in SWI/SNF-deficient cancers of the gynecologic tract. *The Journal of Pathology*, 257(2), 140–145. <https://doi.org/10.1002/path.5889>
 14. Martell, K., McIntyre, J. B., Abedin, T., Kornaga, E. N., Chan, A. M. Y., Enwere, E., **Köbel, M.**, Dean, M. L., Phan, T., Ghatage, P., Lees-Miller, S. P., & Doll, C. M. (2023). Prevalence and Prognostic Significance of PIK3CA Mutation and CNV Status and Phosphorylated AKT Expression in

- Patients With Cervical Cancer Treated With Primary Surgery. *International Journal of Gynecological Pathology*, 43(2), 158–170. <https://doi.org/10.1097/pgp.0000000000000978>
15. Praetorius, T. H., Leonova, A., Lac, V., Senz, J., Tessier-Cloutier, B., Nazeran, T. M., **Köbel, M.**, Grube, M., Kraemer, B., Yong, P. J., Kommoss, S., & Anglesio, M. S. (2022). Molecular analysis suggests oligoclonality and metastasis of endometriosis lesions across anatomically defined subtypes. *Fertility and Sterility*, 118(3), 524–534. <https://doi.org/10.1016/j.fertnstert.2022.05.030>
 16. Scott, S. A., Llaurodo Fernandez, M., Kim, H., Elit, L., Nourmoussavi, M., Glaze, S., Roberts, L., Offman, S. L., Rahimi, K., Lytwyn, A., Sur, M., Gilks, C. B., Matheson, K., **Köbel, M.**, Dawson, A., Tinker, A. V., Kwon, J. S., Hoskins, P., Santos, J. L., & Cheung, A. (2022). Corrigendum to “Low-grade serous carcinoma (LGSC): A Canadian multicenter review of practice patterns and patient outcomes” [Gynecologic Oncology, Volume 157, Issue 1, April 2020, Pages 36–45]. *Gynecologic Oncology*, 167(2), 399–399. <https://doi.org/10.1016/j.ygyno.2022.10.009>
 17. Shakfa, N., Li, D., Conseil, G., Lightbody, E. D., Wilson-Sanchez, J., Hamade, A., Chenard, S., Jawa, N. A., Laight, B. J., Afriyie-Asante, A., Tyryshkin, K., **Koebel, M.**, & Koti, M. (2023). Cancer cell genotype associated tumor immune microenvironment exhibits differential response to therapeutic STING pathway activation in high-grade serous ovarian cancer. *Journal for ImmunoTherapy of Cancer*, 11(4), e006170–e006170. <https://doi.org/10.1136/jitc-2022-006170>
 18. Tessier-Cloutier, B., Kang, E., Alex, D., Stewart, C. J. R., McCluggage, W. G., **Köbel, M.**, & Lee, C. (2022). Endometrial neuroendocrine carcinoma and undifferentiated carcinoma are distinct entities with overlap in neuroendocrine marker expression. *Histopathology*, 81(1), 44–54. <https://doi.org/10.1111/his.14639>
 19. Weir, A., Kang, E.-Y., Meagher, N. S., Nelson, G. S., Ghatage, P., Lee, C.-H., Riggan, M. J., Gentry-Maharaj, A., Ryan, A., Singh, N., Widschwendter, M., Alsop, J., Anglesio, M. S., Beckmann, M. W., Berger, J., Bisinotto, C., Boros, J., Brand, A. H., Brenton, J. D., & Brooks-Wilson, A. (2022). Increased FOXJ1 protein expression is associated with improved overall survival in high-grade serous ovarian carcinoma: an Ovarian Tumor Tissue Analysis Consortium Study. *British Journal of Cancer*, 128(1), 137–147. <https://doi.org/10.1038/s41416-022-02014-y>

Koro, Konstantin

1. Eszlinger, M., Stewardson, P., McIntyre, J. B., Box, A., Khalil, M., Hyrcza, M., **Koro, K.**, Ruether, D., Wu, J., & Paschke, R. (2022). Systematic population-based identification of NTRK and RET fusion-positive thyroid cancers. *European Thyroid Journal*, 11(1). <https://doi.org/10.1530/etj-21-0061>
2. Kishlyansky, D., Kline, G., Mahajan, A., **Koro, K.**, Pasiaka, J. L., & Champagne, P. (2022). Ectopic Cushing’s syndrome from an ACTH-producing pheochromocytoma with a non-functioning pituitary adenoma. *Endocrinology, Diabetes & Metabolism Case Reports*, 2022. <https://doi.org/10.1530/edm-21-0189>
3. Nguyen, V. V., **Koro, K.**, Coffin, C. S., Wang, W., Syeda, N., Meatherall, B., & Lee, S. S. (2022). Herpes simplex virus hepatitis in a renal transplant recipient seronegative pre-transplant. *Canadian Liver Journal*. <https://doi.org/10.3138/canlivj-2022-0007>
4. *Slack, J. C., Bründler, M.-A., Box, A., & **Koro, K.** (2022). A Subset of Pancreatoblastomas May Arise From an Adenomatous Precursor: An Ampullary Pancreatoblastoma and Adjacent Adenoma with a Shared Molecular Phenotype in an Adult Patient. *Pancreas*, 51(10), 1455–1460. <https://doi.org/10.1097/mpa.0000000000002189>

Kurek, Kyle

1. Ceccherini, I., **Kurek, K. C.**, & Weese-Mayer, D. E. (2022). Developmental disorders affecting the respiratory system: CCHS and ROHHAD. *Handbook of Clinical Neurology*, 53–91. <https://doi.org/10.1016/b978-0-323-91532-8.00005-7>
2. Khaytin, I., Victor, A. K., Barclay, S. F., Benson, L. A., Slattery, S. M., Rand, C. M., **Kurek, K. C.**, & Weese-Mayer, D. E. (2023). Rapid-onset obesity with hypothalamic dysfunction, hypoventilation, and autonomic dysregulation (ROHHAD): a collaborative review of the current understanding. *Clinical Autonomic Research*, 33(3), 251–268. <https://doi.org/10.1007/s10286-023-00936-y>
3. Kryshchalskyj, M. T., Agi, J., Ramien, M. L., **Kurek, K. C.**, & Kherani, F. (2023). Lichen sclerosus of the upper eyelid in a paediatric patient: a novel presentation. *Canadian Journal of Ophthalmology*, 58(6), e267–e268. <https://doi.org/10.1016/j.jcjo.2023.07.012>
4. Park, P. C., **Kurek, K. C.**, DeCoteau, J., Howlett, C. J., Hawkins, C., Izevbaye, I., Carter, M. D., Redpath, M., Lo, B., Alex, D., Yousef, G., Yip, S., & Maung, R. (2022). CAP-ACP Workload Model for Advanced Diagnostics in Precision Medicine. *American Journal of Clinical Pathology*. <https://doi.org/10.1093/ajcp/aqac012>
5. Perez-Atayde, A. R., Debelenko, L., Al-Ibraheemi, A., Eng, W., Ruiz-Gutierrez, M., O'Hare, M., Croteau, S. E., Trenor, C. C., Boyer, D., Balkin, D. M., Barclay, S. F., Belinda Hsi Dickie, Liang, M. G., Chaudry, G., Alomari, A. I., Mulliken, J. B., Adams, D. M., **Kurek, K. C.**, Fishman, S. J., & Kozakewich, H. P. W. (2022). Kaposiform Lymphangiomatosis. *The American Journal of Surgical Pathology*, 46(7), 963–976. <https://doi.org/10.1097/pas.0000000000001898>
6. Slattery, S. M., Perez, I. A., Ceccherini, I., Chen, M. L., **Kurek, K. C.**, Kai Lee Yap, Keens, T. G., Khaytin, I., Ballard, H. A., Sokol, E. A., Mittal, A., Rand, C. M., & Weese-Mayer, D. E. (2022). Transitional care and clinical management of adolescents, young adults, and suspected new adult patients with congenital central hypoventilation syndrome. *Clinical Autonomic Research*, 33(3), 231–249. <https://doi.org/10.1007/s10286-022-00908-8>
7. Wright, J. R., Chan, S., Morgen, E. K., Maung, R. T. A., Brundler, M.-A., El Demellawy, D., Fraser, R. B., **Kurek, K. C.**, Magee, F., Nizalik, E., Oligny, L. L., Somers, G. R., Stefanovici, C., & Terry, J. (2022). Workload Measurement in Subspecialty Placental Pathology in Canada. *Pediatric and Developmental Pathology*, 25(6), 604–610. <https://doi.org/10.1177/10935266221118150>

Langdon, Kristopher

1. Dimond, D., Tehrani, N., Ambikkumar, A., Madill, J., Naert, K., **Langdon, K. D.**, & Camara-Lemarroy, C. (2023). Dermatomyositis Immunosuppression in Bacillus Calmette–Guerin-Treated Urothelial Cancer. *Canadian Journal of Neurological Sciences*, 1–3. <https://doi.org/10.1017/cjn.2023.238>
2. Roczowski, A., Limonta, D., Fernandes, J. P., Branton, W. G., Clarke, M., Hlavay, B., Noyce, R. S., Joseph, J. T., Ogando, N. S., Das, S. K., Elaish, M., Arbour, N., Evans, D. H., **Langdon, K.**, Hobman, T. C., & Power, C. (2023). COVID-19 Induces Neuroinflammation and Suppresses Peroxisomes in the Brain. *Annals of Neurology*, 94(3), 531–546. <https://doi.org/10.1002/ana.26679>
3. Sjonnesen, K., Hader, W., Xu, Q., Jacobs-Levan, J., Federico, P., **Langdon, K. D.**, & Appendino, J. P. (2023). Chronic Herpes Simplex Virus Encephalitis with Unexpected Neuropathological Findings. *Canadian Journal of Neurological Sciences*, 1–2. <https://doi.org/10.1017/cjn.2023.311>

Larios, Oscar

1. *Acosta, N., Bautista, M. A., Waddell, B., Du, K., McC Calder, J., Pradhan, P., Sedaghat, N., Papparis, C., Alexander Buchner Beaudet, Chen, J., Van Doorn, J., Xiang, K., Chan, L., Vivas, L., Low, K., Lu, X., Lee, J., Westlund, P., Chekouo, T., & Dai, X, **Larios, O.** (2023). Surveillance for SARS-CoV-2 and its variants in wastewater of tertiary care hospitals correlates with increasing case burden and outbreaks. *Journal of Medical Virology*, 95(2). <https://doi.org/10.1002/jmv.28442>
2. Bresee, L. C., Lamont, N., Ocampo, W., Holroyd-Leduc, J., Sabuda, D., Leal, J., Dalton, B., Kaufman, J., Missaghi, B., Kim, J., **Larios, O. E.**, Henderson, E., Raman, M., Fletcher, J. R., Faris, P., Kraft, S., Shen, Y., Louie, T., & Conly, J. M. (2023). Implementation strategies for hospital-based probiotic administration in a stepped-wedge cluster randomized trial design for preventing hospital-acquired *Clostridioides difficile* infection. *BMC Health Services Research*, 23(1). <https://doi.org/10.1186/s12913-023-10350-9>
3. King, T. L., Chew, D. S., Leal, J., Cannon, K., Exner, D. V., Smith, S., **Larios, O.**, Bush, K., Yuen, B., Zhang, Z., & Rennert-May, E. (2023). Complex cardiac implantable electronic device infections in Alberta, Canada: An epidemiologic cohort study of validated administrative data. *Infection Control & Hospital Epidemiology*, 44(10), 1607–1613. <https://doi.org/10.1017/ice.2023.48>
4. *Leal, J., Farkas, B., Mastikhina, L., Flanagan, J., Skidmore, B., Salmon, C., Dixit, D., Smith, S., Tsekrekos, S., Lee, B., Vayalumkal, J., Dunn, J., Harrison, R., Cordoviz, M., Dubois, R., Chandran, U., Clement, F., Bush, K., Conly, J., & **Larios, O.** (2022). Risk of transmission of respiratory viruses during aerosol-generating medical procedures (AGMPs) revisited in the COVID-19 pandemic: a systematic review. *Antimicrobial Resistance and Infection Control*, 11(1). <https://doi.org/10.1186/s13756-022-01133-8>
5. Leal, J., Shen, Y., Faris, P., Dalton, B., Sabuda, D., Ocampo, W., Bresee, L., Chow, B., Fletcher, J. R., Henderson, E., Kaufman, J., Kim, J., Raman, M., Kraft, S., Lamont, N. C., **Larios, O.**, Missaghi, B., Holroyd-Leduc, J., Louie, T., & Conly, J. (2023). Effectiveness of Bio-K+ for the prevention of *Clostridioides difficile* infection: Stepped-wedge cluster-randomized controlled trial. *Infection Control and Hospital Epidemiology*, 1–9. <https://doi.org/10.1017/ice.2023.169>
6. Loeb, M., Bartholomew, A., Hashmi, M., Tarhuni, W., Hassany, M., Youngster, I., Somayaji, R., **Larios, O.**, Kim, J., Missaghi, B., Vayalumkal, J. V., Mertz, D., Chagla, Z., Cividino, M., Ali, K., Mansour, S., Castellucci, L. A., Frenette, C., Parkes, L., & Downing, M. (2022). Medical Masks Versus N95 Respirators for Preventing COVID-19 Among Health Care Workers. *Annals of Internal Medicine*. <https://doi.org/10.7326/m22-1966>
7. Rennert-May, E., Chew, D. S., Cannon, K., Zhang, Z., Smith, S., King, T., Exner, D. V., **Larios, O.**, & Leal, J. (2023). The economic burden of cardiac implantable electronic device infections in Alberta, Canada: a population-based study using validated administrative data. *Antimicrobial Resistance and Infection Control*, 12(1). <https://doi.org/10.1186/s13756-023-01347-4>
8. Rennert-May, E., Leal, J., MacDonald, M. K., Cannon, K., Smith, S., Exner, D., **Larios, O. E.**, Bush, K., & Chew, D. (2022). Validating administrative data to identify complex surgical site infections following cardiac implantable electronic device implantation: a comparison of traditional methods and machine learning. *ProQuest*, 1–9. <https://doi.org/10.1186/s13756-022-01174-z>

Lee, Sandra

1. Li, X., Quick, C., Zhou, H., Gaynor, S. M., Liu, Y., Chen, H., Margaret Sunitha Selvaraj, Sun, R., **Lee, S.**, Dey, R., Arnett, D. K., Bielak, L. F., Bis, J. C., Blangero, J., Boerwinkle, E., Bowden, D. W., Brody, J. A., Cade, B. E., Correa, A., Cupples, L. A., & Curran, J. E. (2022). Powerful, scalable and resource-efficient meta-analysis of rare variant associations in large whole genome sequencing studies. *Nature Genetics*, 55(1), 154–164. <https://doi.org/10.1038/s41588-022-01225-6>
2. Li, Z., Li, X., Zhou, H., Gaynor, S. M., Margaret Sunitha Selvaraj, Arapoglou, T., Quick, C., Liu, Y., Chen, H., Sun, R., Dey, R., Arnett, D. K., Auer, P. L., Bielak, L. F., **Lee, S.**, Bis, J. C., Blackwell, T. W.,

- Blangero, J., Boerwinkle, E., Bowden, D. W., & Brody, J. A. (2022). A framework for detecting noncoding rare-variant associations of large-scale whole-genome sequencing studies. *Nature Methods*, *19*(12), 1599–1611. <https://doi.org/10.1038/s41592-022-01640-x>
3. Margaret Sunitha Selvaraj, Li, X., Li, Z., Pampana, A., Zhang, D. Y., Park, J., Aslibekyan, S., Bis, J. C., Brody, J. A., Cade, B. E., Chuang, L., Chung, R., **Lee, S.**, Curran, J. E., Lisa, Paul, Duggirala, R., Freedman, B. I., Graff, M., Guo, X., & Heard-Costa, N. L. (2022). Whole genome sequence analysis of blood lipid levels in >66,000 individuals. *Nature Communications*, *13*(1). <https://doi.org/10.1038/s41467-022-33510-7>
 4. Thompson, E. F., Huvila, J., Jamieson, A., Leung, S., Lum, A., Offman, S., Lytwyn, A., Sur, M. L., Hoang, L., Irving, J., van der Westhuizen, N., **Lee, S.**, Morin, C., Bicamumpaka, C., Azordegan, N., Gougeon, F., Ennour-Idrissi, K., Senz, J., McConechy, M. K., Aguirre-Hernandez, R., & Lui, V. (2022). Variability in endometrial carcinoma pathology practice: opportunities for improvement with molecular classification. *Modern Pathology*, *35*(12), 1974–1982. <https://doi.org/10.1038/s41379-022-01165-w>
 5. Wainschtein, P., Jain, D., Zheng, Z., Aslibekyan, S., Becker, **Lee, S.**, D., Bi, W., Brody, J., Carlson, J. C., Correa, A., Du, M. M., Fernandez-Rhodes, L., Ferrier, K. R., Graff, M., Guo, X., He, J., Heard-Costa, N. L., Highland, H. M., Hirschhorn, J. N., Howard-Claudio, C. M., & Isasi, C. R. (2022). Assessing the contribution of rare variants to complex trait heritability from whole-genome sequence data. *Nature Genetics*, *54*(3), 263–273. <https://doi.org/10.1038/s41588-021-00997-7>
 6. Weinstock, J. S., **Lee, S.**, Gopakumar, J., Burugula, B. B., Uddin, M. M., Jahn, N., Belk, J. A., Bouzid, H., Daniel, B., Miao, Z., Ly, N., Mack, T. M., Luna, S. E., Prothro, K. P., Mitchell, S. R., Laurie, C. A., Broome, J. G., Taylor, K. D., Guo, X., Sinner, M. F., & von Falkenhausen, A. S. (2023). Aberrant activation of TCL1A promotes stem cell expansion in clonal haematopoiesis. *Nature*, 1–11. <https://doi.org/10.1038/s41586-023-05806-1>
 7. Wong, W. J., Emdin, C., Bick, A. G., Zekavat, S. M., Niroula, A., Pirruccello, J. P., Dichtel, L., Griffin, G., Uddin, M. M., Gibson, C. J., Kovalcik, V., Lin, A. E., **Lee, S.**, McConkey, M. E., Vromman, A., Sellar, R. S., Kim, P. G., Agrawal, M., Weinstock, J., Long, M. T., & Yu, B. (2023). Clonal haematopoiesis and risk of chronic liver disease. *Nature*, *616*(7958), 747–754. <https://doi.org/10.1038/s41586-023-05857-4>
 8. Bassi, A., Nelson, G., Lee, C. H., Ogilvie, T., Cota, A., & **Lee, S.** (2022). Somatic yolk sac differentiation in tumors of the gynecologic tract: A report of two cases and review of the literature. *Gynecologic Oncology Reports*, *44*, 101119–101119. <https://doi.org/10.1016/j.gore.2022.101119>
 9. Jiang, W. N., Chan, A. M. Y., Urgoiti, G. R., Kornaga, E., Mathen, P., Yeung, R., Konno, M., **Lee, S.**, Box, A., Köbel, M., Joseph, K. J., & Doll, C. M. (2018). The Prognostic Impact of PD-L1 and CD8 Expression in Anal Cancer Patients Treated with Chemoradiotherapy. *International Journal of Radiation Oncology Biology Physics*, *102*(3), S62–S63. <https://doi.org/10.1016/j.ijrobp.2018.06.175>
 10. Köbel, M., Kang, E., **Lee, S.**, Terzic, T., Karnezis, A. N., Prafull Ghatage, Woo, L., Lee, C., Meagher, N. S., Ramus, S. J., & Gorringer, K. L. (2023). Infiltrative pattern of invasion is independently associated with shorter survival and desmoplastic stroma markers FAP and THBS2 in mucinous ovarian carcinoma. *Histopathology*. <https://doi.org/10.1111/his.15128>
 11. ***Lee, S.** (2022). Chronic Endometritis: Diagnostic Considerations in Patients With Infertility. *AJSP: Reviews and Reports*, *27*(5), 222–226. <https://doi.org/10.1097/pcr.0000000000000523>
 12. Wong, W. J., Connor Emdin, Bick, A. G., Zekavat, S. M., Niroula, A., Pirruccello, J. P., Dichtel, L., Griffin, G., Uddin, M. M., Gibson, C. J., Kovalcik, V., Lin, A. E., McConkey, M. E., Vromman, A., Sellar, R. S., Kim, P. G., Agrawal, M., Weinstock, J., Long, M. T., & Yu, B. (2023). Author Correction: Clonal haematopoiesis and risk of chronic liver disease. *Nature*, *619*(7970), E47–E47. <https://doi.org/10.1038/s41586-023-06375-z>

Lisboa, Luiz

1. *Mah, J., Bakker, A., Tseng, C., Lafay-Cousin, L., Kuhn, S., Brundler, M.-A., & **Lisboa, L. F.** (2022). Isolated Pulmonary Emergomycosis in an Immunocompetent Patient in Alberta, Canada. *Open Forum Infectious Diseases*, 9(3). <https://doi.org/10.1093/ofid/ofac021>
2. Mossman, A. K., Svishchuk, J., Waddell, B., Izydorczyk, C., Buckley, P. T., Hilliard, J. J., Al-Ghalith, G. A., Zheng, L., Lynch, A. S., Mody, C. H., **Lisboa, L. F.**, Gregson, D. B., & Parkins, M. D. (2022). *Staphylococcus aureus* in Non-Cystic Fibrosis Bronchiectasis: Prevalence and Genomic Basis of High Inoculum β -Lactam Resistance. *Annals of the American Thoracic Society*, 19(8), 1285–1293. <https://doi.org/10.1513/annalsats.202108-965oc>
3. Svishchuk, J., Ebbert, K., Waddell, B., Izydorczyk, C., Acosta, N., Somayaji, R., Rabin, H. R., Bjornson, C. L., **Lisboa, L.**, Gregson, D. B., Conly, J. M., Surette, M. G., & Parkins, M. D. (2023). Epidemiology and impact of methicillin-sensitive *Staphylococcus aureus* with β -lactam antibiotic inoculum effects in adults with cystic fibrosis. *Antimicrobial Agents and Chemotherapy*, 67(12). <https://doi.org/10.1128/aac.00136-23>

Mahe, Etienne

1. Bégin, P., Callum, J., Jamula, E., Cook, R., Heddle, N. M., Tinmouth, A., Zeller, M. P., Beaudoin-Bussièrès, G., Amorim, L., Bazin, R., Loftsgard, K. C., Carl, R., Chassé, M., Cushing, M. M., **Mahe, E.**, Daneman, N., Devine, D. V., Dumaresq, J., Fergusson, D. A., Gabe, C., & Glesby, M. J. (2022). Author Correction: Convalescent plasma for hospitalized patients with COVID-19: an open-label, randomized controlled trial. *Nature Medicine*, 28(1), 212–212. <https://doi.org/10.1038/s41591-021-01667-1>
2. Agbani, E. O., Chow, L., Nicholas, J., Skeith, L., Schneider, P., Gregory, A., **Mahe, E.**, Yamaura, L., Young, D., Dufour, A., Paul, P. P., Walker, A. M., Mukherjee, P. G., Poole, A. W., Poon, M.-C., & Lee, A. (2023). Overexpression of facilitative glucose transporter-3 and membrane procoagulation in maternal platelets of preeclamptic pregnancy. *Journal of Thrombosis and Haemostasis*, 21(7), 1903–1919. <https://doi.org/10.1016/j.jtha.2023.03.014>
3. Agbani, E. O., **Mahe, E.**, Chaturvedi, S., Yamaura, L., Schneider, P., Barber, M. R. W., Choi, M., Lee, A., & Skeith, L. (2022). Platelets and neutrophils co-drive procoagulant potential in secondary antiphospholipid syndrome during pregnancy. *Thrombosis Research*, 220, 141–144. <https://doi.org/10.1016/j.thromres.2022.10.018>
4. Banoei, M. M., **Mahé, E.**, Mansoor, A., Stewart, D., Winston, B. W., Habibi, H. R., & Shabani-Rad, M.-T. (2022). NMR-based metabolomic profiling can differentiate follicular lymphoma from benign lymph node tissues and may be predictive of outcome. *Scientific Reports*, 12(1). <https://doi.org/10.1038/s41598-022-12445-5>
5. *Bégin, P., Callum, J., Cook, R., Jamula, E., Liu, Y., Finzi, A., Bégin, P., Callum, J., Jamula, E., Cook, R., Heddle, N. M., Tinmouth, A., Zeller, M. P., Beaudoin-Bussièrès, G., Amorim, L., Bazin, R., Loftsgard, K. C., Carl, R., Chassé, M., & Cushing, M. M., **Mahe, E.** (2022). Reply to: Concerns about estimating relative risk of death associated with convalescent plasma for COVID-19. *Nature Medicine*, 28(1), 53–58. <https://doi.org/10.1038/s41591-021-01639-5>
6. Bergeron, J., Capo-Chichi, J.-M., Tsui, H., **Mahe, E.**, Berardi, P., Minden, M. D., Brandwein, J. M., & Schuh, A. C. (2023). The Clinical Utility of FLT3 Mutation Testing in Acute Leukemia: A Canadian Consensus. *Current Oncology*, 30(12), 10410–10436. <https://doi.org/10.3390/curroncol30120759>
7. Elzinga, K., Khayambashi, S., Hahn, C., **Mahe, E.**, & Fine, N. M. (2023). Amyloidosis and Carpal Tunnel Syndrome: Surgical Technique for Extended Carpal Tunnel Release with Tenosynovium and Transverse Carpal Ligament Biopsies. *Plastic and Reconstructive Surgery - Global Open*, 11(1), e4757. <https://doi.org/10.1097/gox.0000000000004757>

8. Gillrie, M. R., Rosin, N., Sinha, S., Kang, H., Farias, R., Nguyen, A., Volek, K., Mah, J., **Mahe, E.**, Fritzier, M. J., Yipp, B. G., & Biernaskie, J. (2023). Case report: Immune profiling links neutrophil and plasmablast dysregulation to microvascular damage in post-COVID-19 Multisystem Inflammatory Syndrome in Adults (MIS-A). *Frontiers in Immunology*, *14*. <https://doi.org/10.3389/fimmu.2023.1125960>
9. Khayambashi, S., Elzinga, K., Hahn, C., Chhibber, S., **Mahe, E.**, Miller, R. J. H., White, J. A., Howlett, J. G., Jimenez-Zepeda, V., & Fine, N. M. (2022). Amyloidosis Tissue Confirmation for Tafamidis Eligibility Using Transverse Carpal Ligament and Tenosynovium Biopsy. *Canadian Journal of Cardiology*, *38*(10), 1643–1646. <https://doi.org/10.1016/j.cjca.2022.06.012>
10. Lewis, E., Fine, N., Miller, R. J. H., Hahn, C., Chhibber, S., **Mahe, E.**, Tay, J., Duggan, P., McCulloch, S., Bahlis, N., Neri, P., & Jimenez-Zepeda, V. H. (2022). Amyloidosis and COVID-19: experience from an amyloid program in Canada. *Annals of Hematology*, *101*(10), 2307–2315. <https://doi.org/10.1007/s00277-022-04964-y>
11. Lewis, E., Lee, H., Fine, N., Miller, R., Hahn, C., Tay, J., Chhibber, S., **Mahe, E.**, & Jimenez-Zepeda, V. H. (2023). Monoclonal Gammopathy of Undetermined Significance in Patients With Transthyretin Amyloidosis (ATTR): Analysis Using the iStopMM Criteria. *Clinical Lymphoma, Myeloma & Leukemia*, *23*(3), 211–217. <https://doi.org/10.1016/j.clml.2022.12.012>
12. Lewis, E., McCulloch, S., **Mahe, E.**, Bahlis, N., Neri, P., Tay, J., Duggan, P., & Jimenez-Zepeda, V. H. (2023). Effect of the Presence of t(11;14) for Patients With AL Amyloidosis Treated With Bortezomib-Containing Regimens: Experience From the Amyloidosis Program of Calgary. *Clinical Lymphoma, Myeloma & Leukemia*, *23*(10), e331–e334. <https://doi.org/10.1016/j.clml.2023.07.004>

Mansoor, Adnan

1. Banoei, M. M., Mahé, E., **Mansoor, A.**, Stewart, D., Winston, B. W., Habibi, H. R., & Shabani-Rad, M.-T. (2022). NMR-based metabolomic profiling can differentiate follicular lymphoma from benign lymph node tissues and may be predictive of outcome. *Scientific Reports*, *12*(1). <https://doi.org/10.1038/s41598-022-12445-5>
2. Bavi, P., **Mansoor, A.**, Auer, I., & Hardin, J. (2023). In Situ Follicular B-Cell Neoplasm Presenting as Miliary Facial Papules: A Unique Clinical Presentation and Spontaneous Regression. *The American Journal of Dermatopathology*. <https://doi.org/10.1097/dad.0000000000002579>
3. *Elyamany, G., Rizwan, H., Akhter, A., Aljabry, M. S., Alotaibi, S., Albalawi, M. A. H., Shabani-Rad, M.-T., Roshan, T. M., & **Mansoor, A.** (2023). “Losing the Brakes”—Suppressed Inhibitors Triggering Uncontrolled Wnt/ β -Catenin Signaling May Provide a Potential Therapeutic Target in Elderly Acute Myeloid Leukemia. *Current Issues in Molecular Biology*, *45*(1), 604–613. <https://doi.org/10.3390/cimb45010040>
4. ***Mansoor, A.**, Akhter, A., Hamidi, M., Roshan, T. M., Shabani-Rad, M.-T., & Stewart, D. (2023). Exploring *TBL1XR1* and *NCOR1* Expression in B-cell Lymphoma Subtypes: Interaction With DNA Damage Repair Genes. *Anticancer Research*, *43*(11), 4801–4807. <https://doi.org/10.21873/anticancer.16677>
5. ***Mansoor, A.**, Akhter, A., Kamran, H., Minoo, P., & Stewart, D. (2023). Unraveling the molecular landscape: a comparative analysis of PI3K and MAPK signaling pathways in plasmablastic lymphoma and diffuse large B-cell lymphoma with therapeutic implications. *Human Pathology*, *141*, 102–109. <https://doi.org/10.1016/j.humpath.2023.07.009>
6. ***Mansoor, A.**, Akhter, A., Shabani-Rad, M., Deschenes, J., Yilmaz, A., Trpkov, K., & Stewart, D. (2023). Primary testicular lymphoma demonstrates overexpression of the Wilms tumor 1 gene and different mRNA and miRNA expression profiles compared to nodal diffuse large B-cell lymphoma. *Hematological Oncology*, *41*(5), 828–837. <https://doi.org/10.1002/hon.3190>

7. ***Mansoor, A.**, Kamran, H., Akhter, A., Seno, R., Torlakovic, E. E., Roshan, T. M., Shabani-Rad, M.-T., Elyamany, G., Minoo, P., & Stewart, D. (2023). Identification of Potential Therapeutic Targets for Plasmablastic Lymphoma Through Gene Expression Analysis: Insights into RAS and Wnt Signaling Pathways. *Modern Pathology*, *36*(8), 100198. <https://doi.org/10.1016/j.modpat.2023.100198>
8. ***Mansoor, A.**, Kamran, H., Rizwan, H., Akhter, A., Roshan, T. M., Shabani-Rad, M.-T., Bavi, P., & Stewart, D. (2023). Expression of “DNA damage response” pathway genes in diffuse large B-cell lymphoma: The potential for exploiting synthetic lethality. *Hematological Oncology*. <https://doi.org/10.1002/hon.3225>
9. Vose, J. M., Fu, K., Wang, L., **Mansoor, A.**, Stewart, D. A., Cheng, H., Smith, L. M., Ji, Y., Hina Naushad Qureishi, Link, B. K., Cessna, M. H., Barr, P. M., Kahl, B. S., McKinney, M., Khan, N., Advani, R. H., Martin, P., Goy, A., Phillips, T., & Mehta, A. (2023). Integrative analysis of clinicopathological features defines novel prognostic models for mantle cell lymphoma in the immunochemotherapy era: a report from The North American Mantle Cell Lymphoma Consortium. *Journal of Hematology & Oncology*, *16*(1). <https://doi.org/10.1186/s13045-023-01520-7>

Minoo, Parham

1. *Chezar, K., & **Minoo, P.** (2022). Appendiceal sessile serrated lesions are distinct from their right-sided colonic counterparts and may be precursors for appendiceal mucinous neoplasms. *Human Pathology*, *122*, 40–49. <https://doi.org/10.1016/j.humpath.2022.01.008>
2. *Kubik, T., & **Minoo, P.** (2022). TEMPI syndrome associated with IgM monoclonal gammopathy. *Blood*, *139*(8), 1254–1254. <https://doi.org/10.1182/blood.2021014393>
3. Lih, T. M., Cao, L., **Minoo, P.**, Omenn, G. S., Hruban, R. H., Chan, D. W., Bathe, O. F., & Zhang, H. (2024). Detection of pancreatic ductal adenocarcinoma-associated proteins in serum. *Molecular & Cellular Proteomics*, *23*(1), 100687–100687. <https://doi.org/10.1016/j.mcpro.2023.100687>
4. Mansoor, A., Akhter, A., Kamran, H., **Minoo, P.**, & Stewart, D. (2023). Unraveling the molecular landscape: a comparative analysis of PI3K and MAPK signaling pathways in plasmablastic lymphoma and diffuse large B-cell lymphoma with therapeutic implications. *Human Pathology*, *141*, 102–109. <https://doi.org/10.1016/j.humpath.2023.07.009>
5. Mansoor, A., Kamran, H., Akhter, A., Seno, R., Torlakovic, E. E., Roshan, T. M., Shabani-Rad, M.-T., Elyamany, G., **Minoo, P.**, & Stewart, D. (2023). Identification of Potential Therapeutic Targets for Plasmablastic Lymphoma Through Gene Expression Analysis: Insights into RAS and Wnt Signaling Pathways. *Modern Pathology*, *36*(8), 100198. <https://doi.org/10.1016/j.modpat.2023.100198>
6. *Meagher, N. S., Gorringer, K. L., Wakefield, M. J., Bolithon, A., Nam, C., Chiu, D. S., Anglesio, M. S., Mallitt, K.-A., Doherty, J. A., Harris, H. R., Schildkraut, J. M., Berchuck, A., Cushing-Haugen, K. L., Chezar, K., Chou, A., Tan, A., Alsop, J., Barlow, E., Beckmann, M. W., & Boros, J., **Minoo, P.** (2022). Gene-Expression Profiling of Mucinous Ovarian Tumors and Comparison with Upper and Lower Gastrointestinal Tumors Identifies Markers Associated with Adverse Outcomes. *Clinical Cancer Research*, *28*(24), 5383–5395. <https://doi.org/10.1158/1078-0432.ccr-22-1206>

Naert, Karen

1. Dimond, D., Tehrani, N., Ambikkumar, A., Madill, J., **Naert, K.**, Langdon, K. D., & Camara-Lemarroy, C. (2023). Dermatomyositis Immunosuppression in Bacillus Calmette–Guerin-Treated Urothelial Cancer. *Canadian Journal of Neurological Sciences*, 1–3. <https://doi.org/10.1017/cjn.2023.238>

2. *Wiedemeyer, K., Brenn, T., & **Naert, K.** (2022). Necrotizing Infundibular Crystalline Folliculitis—A Case Report of a Rare Entity and Review of the Literature. *The American Journal of Dermatopathology*, 44(10), 774–777. <https://doi.org/10.1097/dad.0000000000002239>
3. Woo, T., **Naert, K.**, & Chia, J. C. (2022). Pigmented squamous cell carcinoma in situ with amyloid deposition mimicking melanoma. *JAAD Case Reports*, 27, 137–139. <https://doi.org/10.1016/j.jdcr.2022.07.028>

Naugler, Christopher

1. Campbell, D. J. T., Ghuttura, H., Mladenovic, A., Smith, J., Leigh, R., Desveaux, L., Ivers, N., Manns, B., Tonelli, M., **Naugler, C.**, Hemmelgarn, B., & McBrien, K. A. (2022). Variation in Patient-Described Barriers to and Facilitators of Diabetes Management by Individual-Level Characteristics: A Cross-Sectional, Open-Ended Survey. *Clinical Diabetes*, 40(3). <https://doi.org/10.2337/cd21-0060>
2. *Church, D. L., & **Naugler, C.** (2022). Using a systematic approach to strategic innovation in laboratory medicine to bring about change. *Critical Reviews in Clinical Laboratory Sciences*, 59(3), 178–202. <https://doi.org/10.1080/10408363.2021.1997899>
3. de Koning, L., Orton, D., Seiden Long, I., Boyd, J., Kellogg, M., Abdullah, A., **Naugler, C.**, Tsui, A., Strange, B., & Glaser, D. (2023). Distribution of videos demonstrating best practices in preventing hemolysis is associated with reduced hemolysis among nurse-collected specimens in hospitals. *Clinical Biochemistry*, 119, 110632. <https://doi.org/10.1016/j.clinbiochem.2023.110632>
4. *Guo, E., Gupta, M., Deng, J., Park, Y.-J., Paget, M., & **Naugler, C.** (2024). Automated Paper Screening for Clinical Reviews Using Large Language Models: Data Analysis Study. *Journal of Medical Internet Research*, 26, e48996–e48996. <https://doi.org/10.2196/48996>
5. Harvey, A., Paget, M., McLaughlin, K., Busche, K., Touchie, C., **Naugler, C.**, & Desy, J. (2023). *How much is enough? Proposing achievement thresholds for core EPAs of graduating medical students in Canada.* 1–7. <https://doi.org/10.1080/0142159x.2023.2215910>
6. Lang, R., Gill, M. J., Viczko, J., **Naugler, C.**, & Church, D. (2022). Risk Factors and Outcomes of Bloodstream Infections Among People With Human Immunodeficiency Virus: A Longitudinal Cohort Study From 2000 to 2017. *PubMed*, 9(8), ofac318–ofac318. <https://doi.org/10.1093/ofid/ofac318>
7. Yang, J., Dong, Y., **Naugler, C. T.**, & de Koning, L. (2022). Serum 25-hydroxyvitamin D and risk of cancer in a large community population under investigation for cardiovascular disease: a retrospective cohort study. *BMJ Open*, 12(1), e056543–e056543. <https://doi.org/10.1136/bmjopen-2021-056543>

Ng, Denise

1. Sarnat, H. B., Chan, E. S., **Ng, D.**, & Yu, W. (2023). Maturation of metastases in peripheral neuroblastic tumors (neuroblastoma) of children. *Journal of Neuropathology and Experimental Neurology*, 82(10), 853–864. <https://doi.org/10.1093/jnen/nlad068>
2. Watson, P. N., Midha, R., & Ng, D. W. (2023). Causalgia: A Review of Nerve Resection, Amputation, Immunotherapy, and Amputated Limb CRPS II Pathology. *Canadian Journal of Neurological Sciences*, 1–6. <https://doi.org/10.1017/cjn.2023.260>

Nikolic, Ana

1. Chen, H.-M., **Nikolic, A.**, Singhal, D., & Gallo, M. (2022). Roles of Chromatin Remodelling and Molecular Heterogeneity in Therapy Resistance in Glioblastoma. *Cancers*, *14*(19), 4942–4942. <https://doi.org/10.3390/cancers14194942>
2. Mirzaei, R., D’Mello, C., Liu, M., **Nikolic, A.**, Kumar, M., Visser, F., Bose, P., Gallo, M., & Yong, V. W. (2023). Single-Cell Spatial Analysis Identifies Regulators of Brain Tumor–Initiating Cells. *Cancer Research*, OF1–OF17. <https://doi.org/10.1158/0008-5472.can-22-3004>
3. ***Nikolic, A.**, Maule, F., Bobyn, A., Ellestad, K., So Ya Paik, Marhon, S. A., Mehdipour, P., Lun, X., Chen, H.-M., Mallard, C., Hay, A., Johnston, M. J., Gafuik, C., Zemp, F. J., Shen, Y., Ninkovic, N., Osz, K., Labit, E., Berger, N. D., & Brownsey, D. K. (2023). macroH2A2 antagonizes epigenetic programs of stemness in glioblastoma. *Nature Communications*, *14*(1). <https://doi.org/10.1038/s41467-023-38919-2>

Nohr, Erik

1. Chan, E. S., Haider, S., Subramanian, S., Yu, W., **Nohr, E. W.**, & de Koning, L. (2022). Congenital cytomegalovirus infection is associated with congenital rickets: a retrospective autopsy cohort study. *Archives of Disease in Childhood: Fetal & Neonatal*, *108*(4), 435–436. <https://doi.org/10.1136/archdischild-2022-324760>
2. de Koning, L., Crawford, S., **Nohr, E.**, Chadha, R., Horn, C., Wright, J. R., & Chan, E. S. (2022). Recurrence risk of villitis of unknown etiology: Analysis of a large retrospective cohort study, systematic review and meta-analysis. *Placenta*, *120*, 32–39. <https://doi.org/10.1016/j.placenta.2022.02.006>
3. Husereau, D., Bombard, Y., Stockley, T., Carter, M., Davey, S., Lemaire, D., **Nohr, E.**, Park, P., Spatz, A., Williams, C., Pollett, A., Lo, B., Yip, S., El Hallani, S., & Feilotter, H. (2023). Future Role of Health Technology Assessment for Genomic Medicine in Oncology: A Canadian Laboratory Perspective. *Current Oncology*, *30*(11), 9660–9669. <https://doi.org/10.3390/currenocol30110700>

Ogilvie, Travis

1. Bassi, A., Nelson, G., Lee, C. H., **Ogilvie, T.**, Cota, A., & Lee, S. (2022). Somatic yolk sac differentiation in tumors of the gynecologic tract: A report of two cases and review of the literature. *Gynecologic Oncology Reports*, *44*, 101119–101119. <https://doi.org/10.1016/j.gore.2022.101119>

Orton, Dennis

1. de Koning, L., **Orton, D.**, Seiden Long, I., Boyd, J., Kellogg, M., Abdullah, A., Naugler, C., Tsui, A., Strange, B., & Glaser, D. (2023). Distribution of videos demonstrating best practices in preventing hemolysis is associated with reduced hemolysis among nurse-collected specimens in hospitals. *Clinical Biochemistry*, *119*, 110632. <https://doi.org/10.1016/j.clinbiochem.2023.110632>
2. *Ezra, S., Winstone, T. M. L., Singh, R., & **Orton, D. J.** (2023). Agreement of LC-MS assays for IGF-1 traceable to NIST and WHO standards permits harmonization of reference intervals between laboratories. *Clinical Biochemistry*, *116*, 75–78. <https://doi.org/10.1016/j.clinbiochem.2023.04.002>
3. *Wickramarachchi, P., Boyd, J. M., & **Orton, D. J.** (2022). Biological variation in clozapine and metabolite reporting during therapeutic drug monitoring. *Clinica Chimica Acta*, *531*, 183–187. <https://doi.org/10.1016/j.cca.2022.04.005>

Paul, Heather

1. Johnson, J. M., Walsh, J. D., Okun, N. B., Metcalfe, A., Pastuck, M. L., Maxey, C. M., Soliman, N., Mahallati, H., Kuret, V. H., Dwinnell, S. J., Chada, R., O'Quinn, C. P., Schacher, J., Somerset, D. A., Paterson, K., Suchet, I. B., Silang, K. A., **Paul, H.**, Nerenberg, K. A., & Johnson, D. W. (2023). The Implementation of Preeclampsia Screening and Prevention (IMPRESS) Study. *American Journal of Obstetrics & Gynecology MFM*, 5(2), 100815. <https://doi.org/10.1016/j.ajogmf.2022.100815>
2. ***Paul, H. A.**, Chi, Q., Gifford, J. L., & Seiden-Long, I. (2022). Not T too! False elevations in high-sensitivity cardiac troponin T (hs-TnT) following specimen transport. *Clinical Biochemistry*. <https://doi.org/10.1016/j.clinbiochem.2022.08.008>
3. Reid, M. S., **Paul, H. A.**, Mostoufi, A., Robinson, J. L., & Sadrzadeh, S. M. H. (2023). Evaluation of the stability of fecal immunochemical test specimens. *Clinical Biochemistry*, 115, 92–96. <https://doi.org/10.1016/j.clinbiochem.2022.11.015>
4. Wayne Lewis, C., Butorin, Y., de Koning, L., **Paul, H. A.**, Gifford, J. L., Venner, A. A., & Seiden-Long, I. (2023). Low partial pressure of oxygen causes significant and unrecognized under-recovery of glucose on blood gas analyzers. *Clinical Biochemistry*, 111, 60–65. <https://doi.org/10.1016/j.clinbiochem.2022.10.008>

Pillai, Dylan

1. *Castañeda-Mogollón, D., Doolan, C., Toppings, N., Amarasekara, R., Tran, T.-A., & **Pillai, D. R.** (2022). Correlation of Clinical Severity With Stool Microbiome Changes in *Clostridioides difficile* Infection. *Archives of Pathology & Laboratory Medicine*, 147(7), 774–785. <https://doi.org/10.5858/arpa.2021-0636-oa>
2. *Castañeda-Mogollón, D., Toppings, N. B., Kamaliddin, C., Lang, R., Kuhn, S., & **Pillai, D. R.** (2023). Amplicon Deep Sequencing Reveals Multiple Genetic Events Lead to Treatment Failure with Atovaquone-Proguanil in *Plasmodium falciparum*. 67(6). <https://doi.org/10.1128/aac.01709-22>
3. *Doolan, C. P., Sahragard, B., Leal, J., Sharma, A., Kim, J., Spackman, E., Hollis, A., & **Pillai, D. R.** (2023). *Clostridioides difficile* Near-Patient Testing Versus Centralized Testing: A Pragmatic Cluster Randomized Crossover Trial. *Clinical Infectious Diseases*. <https://doi.org/10.1093/cid/ciad046>
4. *Freedman, S. B., Oberding, L. K., Kim, K., Xie, J., Berenger, B. M., Goulden, R., Weisbeck, S., & **Pillai, D. R.** (2022). SARS-CoV-2 Viral Load Quantification, Clinical Findings and Outcomes in Children Seeking Emergency Department Care: Prospective Cohort Study. *Pediatric Infectious Disease Journal*, 41(7), 566–569. <https://doi.org/10.1097/inf.0000000000003527>
5. *Lieu, A., Mah, J., Desgagnés, N., & **Pillai, D. R.** (2022). The impact of the COVID-19 pandemic on malaria in returning travellers in Canada: a retrospective population-based cohort study. *Journal of Travel Medicine*, 29(4). <https://doi.org/10.1093/jtm/taac061>
6. *Linton, K., Oberding, L. K., Ward, L., Kim, J., & **Pillai, D. R.** (2022). Digital Droplet PCR to Track SARS-CoV-2 Outbreak in a Hospital Transitional Care Unit. *American Journal of Infection Control*. <https://doi.org/10.1016/j.ajic.2022.06.010>
7. *Mohon, A. N., Toppings, N., Castañeda-Mogollón, D., Kamaliddin, C., & **Pillai, D. R.** (2023). Ultrasensitive Reverse Transcriptase Loop-Mediated Isothermal Amplification (US-LAMP)-Based Detection of Malaria Infection from Dried Blood Spots. *Methods in Molecular Biology*, 325–337. https://doi.org/10.1007/978-1-0716-2950-5_19
8. ***Pillai, D. R.**, Feleke, S. M., & Bayih, A. G. (2023). Artemisinin-Resistant HRP2-Negative Malaria in Eritrea. *The New England Journal of Medicine*, 389(26), 2497–2498. <https://doi.org/10.1056/nejmc2312559>
9. *Rediet Fikru Gebresenbet, Kamaliddin, C., Bekele, Z. M., Teferi, M., Tegegne, B., Yewhalaw, D., Bayih, A. G., & **Pillai, D. R.** (2022). Active case detection and treatment of malaria in pregnancy

using LAMP technology (LAMPREG): a pragmatic randomised diagnostic outcomes trial—study protocol. *BMJ Open*, 12(7), e058397–e058397. <https://doi.org/10.1136/bmjopen-2021-058397>

10. *Toppings, N. B., Oberding, L. K., Lin, Y.-C., Evans, D., & **Pillai, D. R.** (2022). The Role of Subgenomic RNA in Discordant Results From Reverse Transcription-Polymerase Chain Reaction Tests for COVID-19. *Archives of Pathology & Laboratory Medicine*, 146(7), 805–813. <https://doi.org/10.5858/arpa.2021-0630-sa>

Pitout, Johann

1. *Ibadin, E. & Ogefere, H. & Peirano, G. & **Pitout, J.** (2022). High prevalence of NDM genes among Carbapenamase-producing clinical Gram-negative bacilli in Benin City, Nigeria: *Pseudomonas aeruginosa*-a leading culprit. *New Zealand Journal of Medical Laboratory Science*. 76. 90-92.
2. *Chen, L., Peirano, G., Kreiswirth, B. N., Devinney, R., & **Pitout, J. D. D.** (2022). Acquisition of genomic elements were pivotal for the success of *Escherichia coli* ST410. *Journal of Antimicrobial Chemotherapy*, 77(12), 3399–3407. <https://doi.org/10.1093/jac/dkac329>
3. *Chen, L., Peirano, G., Yen, K., Wang, B., Terlecky, A., DeVinney, R., Kreiswirth, B. N., & **Pitout, J.** (2023). CRISPR-Cas9-mediated IncF plasmid curing in extraintestinal pathogenic *Escherichia coli*. *Microbiology Spectrum*. <https://doi.org/10.1128/spectrum.03692-23>
4. *Dingle, T. C., & **Pitout, J. D. D.** (2022). The Ins and Outs of Susceptibility Testing for New β -Lactam/ β -Lactamase Inhibitor Combinations for Gram-Negative Organisms. *Journal of Clinical Microbiology*, 60(7). <https://doi.org/10.1128/jcm.00807-21>
5. Faustinos Tatenda Takawira, **Pitout, J.**, Thilliez, G., Mashe, T., Gutierrez, A. V., Kingsley, R. A., Peirano, G., Matheu, J., Stanley Munyaradzi Midzi, Lusubilo Witson Mwamakamba, Gally, D. L., Tarupiwa, A., Mukavhi, L., Ehlers, M. M., Mtapuri-Zinyowera, S., & Kock, M. M. (2022). Faecal carriage of ESBL producing and colistin resistant *Escherichia coli* in avian species over a 2-year period (2017-2019) in Zimbabwe. *Frontiers in Cellular and Infection Microbiology*, 12. <https://doi.org/10.3389/fcimb.2022.1035145>
6. *Mataseje, L. F., Chen, L., Peirano, G., Fakharuddin, K., Kreiswirth, B., Mulvey, M., & **Pitout, J. D. D.** (2022). *Klebsiella pneumoniae* ST147: and then there were three carbapenemases. *European Journal of Clinical Microbiology & Infectious Diseases*, 41(12), 1467–1472. <https://doi.org/10.1007/s10096-022-04514-4>
7. Mataseje, L. F., **Pitout, J. D. D.**, Croxen, M., Mulvey, M. R., & Dingle, T. C. (2023). Three separate acquisitions of blaNDM-1 in three different bacterial species from a single patient. *European Journal of Clinical Microbiology & Infectious Diseases*, 42(10), 1275–1280. <https://doi.org/10.1007/s10096-023-04651-4>
8. *Nobrega, D., Peirano, G., Matsumura, Y., & **Pitout, J. D. D.** (2023). Molecular Epidemiology of Global Carbapenamase-Producing *Citrobacter* spp. (2015–2017). *Microbiology Spectrum*, 11(2). <https://doi.org/10.1128/spectrum.04144-22>
9. *Nobrega, D., Peirano, G., & **Pitout, J.** (2022). *Escherichia coli* sequence type 73 bloodstream infections in a centralized Canadian region and their association with companion animals: an ecological study. *Infection*, 50(6), 1579–1585. <https://doi.org/10.1007/s15010-022-01856-1>
10. *Peirano, G., Chen, L., Nobrega, D., Finn, T. J., Kreiswirth, B. N., DeVinney, R., & **Pitout, J. D. D.** (2022). Genomic Epidemiology of Global Carbapenamase-Producing *Escherichia coli*, 2015–2017. *Emerging Infectious Diseases*, 28(5). <https://doi.org/10.3201/eid2805.212535>
11. ***Pitout, J. D. D.**, & Chen, L. (2023). The Significance of Epidemic Plasmids in the Success of Multidrug-Resistant Drug Pandemic Extraintestinal Pathogenic *Escherichia coli*. *Infectious Diseases and Therapy*. <https://doi.org/10.1007/s40121-023-00791-4>

12. ***Pitout, J. D. D.**, Peirano, G., Chen, L., DeVinney, R., & Matsumura, Y. (2022). Escherichia coli ST1193: Following in the Footsteps of E. coli ST131. *Antimicrobial Agents and Chemotherapy*, 66(7). <https://doi.org/10.1128/aac.00511-22>
13. ***Pitout, J. D.**, Peirano, G., & DeVinney, R. (2023). The contributions of multidrug resistant clones to the success of pandemic Extra-intestinal Pathogenic Escherichia coli. *Expert Review of Anti-Infective Therapy*. <https://doi.org/10.1080/14787210.2023.2184348>

Roshan, Tariq

1. Elyamany, G., Rizwan, H., Akhter, A., Aljabry, M. S., Alotaibi, S., Albalawi, M. A. H., Shabani-Rad, M.-T., **Roshan, T. M.**, & Mansoor, A. (2023). “Losing the Brakes”—Suppressed Inhibitors Triggering Uncontrolled Wnt/ β -Catenin Signaling May Provide a Potential Therapeutic Target in Elderly Acute Myeloid Leukemia. *Current Issues in Molecular Biology*, 45(1), 604–613. <https://doi.org/10.3390/cimb45010040>
2. Mansoor, A., Akhter, A., Hamidi, M., **Roshan, T. M.**, Shabani-Rad, M.-T., & Stewart, D. (2023). Exploring *TBL1XR1* and *NCOR1* Expression in B-cell Lymphoma Subtypes: Interaction With DNA Damage Repair Genes. *Anticancer Research*, 43(11), 4801–4807. <https://doi.org/10.21873/anticancer.16677>
3. Mansoor, A., Kamran, H., Akhter, A., Seno, R., Torlakovic, E. E., **Roshan, T. M.**, Shabani-Rad, M.-T., Elyamany, G., Minoo, P., & Stewart, D. (2023). Identification of Potential Therapeutic Targets for Plasmablastic Lymphoma Through Gene Expression Analysis: Insights into RAS and Wnt Signaling Pathways. *Modern Pathology*, 36(8), 100198. <https://doi.org/10.1016/j.modpat.2023.100198>
4. Mansoor, A., Kamran, H., Rizwan, H., Akhter, A., **Roshan, T. M.**, Shabani-Rad, M.-T., Bavi, P., & Stewart, D. (2023). Expression of “DNA damage response” pathway genes in diffuse large B-cell lymphoma: The potential for exploiting synthetic lethality. *Hematological Oncology*. <https://doi.org/10.1002/hon.3225>
5. *Shameli, A., & **Roshan, T. M.** (2022). CD200 expression on Sezary cells: A valuable tool for flow cytometric assessment of peripheral blood T-cell neoplasms. *Cytometry Part B: Clinical Cytometry*, 102(4), 303–311. <https://doi.org/10.1002/cyto.b.22080>

Seiden Long, Isolde

1. Bibby, H. L., de Koning, L., **Seiden-Long, I.**, Zelyas, N., Church, D. L., & Berenger, B. M. (2022). A pragmatic randomized controlled trial of rapid on-site influenza and respiratory syncytial virus PCR testing in paediatric and adult populations. *BMC Infectious Diseases*, 22(1). <https://doi.org/10.1186/s12879-022-07796-3>
2. Butorin, Y., **Seiden-Long, I.**, & de Koning, L. (2024). A verification and implementation plan for 50 blood gas analyzers across a large geographic area. *Clinical Biochemistry*, 123, 110702–110702. <https://doi.org/10.1016/j.clinbiochem.2023.110702>
3. de Koning, L., Orton, D., **Seiden Long, I.**, Boyd, J., Kellogg, M., Abdullah, A., Naugler, C., Tsui, A., Strange, B., & Glaser, D. (2023). Distribution of videos demonstrating best practices in preventing hemolysis is associated with reduced hemolysis among nurse-collected specimens in hospitals. *Clinical Biochemistry*, 119, 110632. <https://doi.org/10.1016/j.clinbiochem.2023.110632>
4. *Gifford, J. L., & **Seiden-Long, I.** (2022). Filling in the GAPS: validation of anion gap (AGAP) measurement uncertainty estimates for use in clinical decision making. *Clinical Chemistry and Laboratory Medicine*, 60(6), 851–858. <https://doi.org/10.1515/cclm-2021-1279>
5. Langman, L. J., Rushton, A. M., Thomas, D., Colbourne, P., **Seiden-Long, I.**, Brun, M. M., Colantonio, D., & Jannetto, P. J. (2022). Drug Testing In Support Of The Diagnosis Of Neonatal

Abstinence Syndrome: The Current Situation. *Clinical Biochemistry*.
<https://doi.org/10.1016/j.clinbiochem.2022.11.002>

6. *Paul, H. A., Chi, Q., Gifford, J. L., & **Seiden-Long, I.** (2022). Not T too! False elevations in high-sensitivity cardiac troponin T (hs-TnT) following specimen transport. *Clinical Biochemistry*.
<https://doi.org/10.1016/j.clinbiochem.2022.08.008>
7. *Wayne Lewis, C., Butorin, Y., de Koning, L., Paul, H. A., Gifford, J. L., Venner, A. A., & **Seiden-Long, I.** (2023). Low partial pressure of oxygen causes significant and unrecognized under-recovery of glucose on blood gas analyzers. *Clinical Biochemistry*, *111*, 60–65.
<https://doi.org/10.1016/j.clinbiochem.2022.10.008>

Shabani-Rad, Meer-Taher

1. Asma Amir Ali, **Shabani-Rad, M.-T.**, & Chia, J. C. (2022). Solitary pink papule in an elderly man. *JAAD Case Reports*, *24*, 48–51. <https://doi.org/10.1016/j.jdc.2022.04.004>
2. *Banoei, M. M., Mahé, E., Mansoor, A., Stewart, D., Winston, B. W., Habibi, H. R., & **Shabani-Rad, M.-T.** (2022). NMR-based metabolomic profiling can differentiate follicular lymphoma from benign lymph node tissues and may be predictive of outcome. *Scientific Reports*, *12*(1).
<https://doi.org/10.1038/s41598-022-12445-5>
3. Elyamany, G., Rizwan, H., Akhter, A., Aljabry, M. S., Alotaibi, S., Albalawi, M. A. H., **Shabani-Rad, M.-T.**, Roshan, T. M., & Mansoor, A. (2023). “Losing the Brakes”—Suppressed Inhibitors Triggering Uncontrolled Wnt/ β -Catenin Signaling May Provide a Potential Therapeutic Target in Elderly Acute Myeloid Leukemia. *Current Issues in Molecular Biology*, *45*(1), 604–613.
<https://doi.org/10.3390/cimb45010040>
4. Mansoor, A., Akhter, A., Hamidi, M., Roshan, T. M., **Shabani-Rad, M.-T.**, & Stewart. (2023). Exploring *TBL1XR1* and *NCOR1* Expression in B-cell Lymphoma Subtypes: Interaction With DNA Damage Repair Genes. *Anticancer Research*, *43*(11), 4801–4807.
<https://doi.org/10.21873/anticancer.16677>
5. Mansoor, A., Akhter, A., **Shabani-Rad, M.**, Deschenes, J., Yilmaz, A., Trpkov, K., & Stewart, D. (2023). Primary testicular lymphoma demonstrates overexpression of the Wilms tumor 1 gene and different mRNA and miRNA expression profiles compared to nodal diffuse large B-cell lymphoma. *Hematological Oncology*, *41*(5), 828–837. <https://doi.org/10.1002/hon.3190>
6. Mansoor, A., Kamran, H., Akhter, A., Seno, R., Torlakovic, E. E., Roshan, T. M., **Shabani-Rad, M.-T.**, Elyamany, G., Minoo, P., & Stewart, D. (2023). Identification of Potential Therapeutic Targets for Plasmablastic Lymphoma Through Gene Expression Analysis: Insights into RAS and Wnt Signaling Pathways. *Modern Pathology*, *36*(8), 100198.
<https://doi.org/10.1016/j.modpat.2023.100198>
7. Mansoor, A., Kamran, H., Rizwan, H., Akhter, A., Roshan, T. M., **Shabani-Rad, M.-T.**, Bavi, P., & Stewart, D. (2023). Expression of “DNA damage response” pathway genes in diffuse large B-cell lymphoma: The potential for exploiting synthetic lethality. *Hematological Oncology*.
<https://doi.org/10.1002/hon.3225>
8. *Shameli, A., Dharmani-Khan, P., Auer, I., & **Shabani-Rad, M.-T.** (2023). Deep immunophenotypic analysis of the bone marrow progenitor cells in myelodysplastic syndromes. *Leukemia Research*, *134*, 107401–107401.
<https://doi.org/10.1016/j.leukres.2023.107401>
9. Tripathi, G., Khanolkar, R. A., Faridi, R. M., Kalra, A., Dharmani-Khan, P., **Shabani-Rad, M.-T.**, Berka, N., Daly, A., Storek, J., & Khan, F. M. (2022). Donor Genetic Predisposition to High Interleukin-10 Production Appears Protective against Acute Graft-Versus-Host Disease. *International Journal of Molecular Sciences*, *23*(24), 15888–15888.
<https://doi.org/10.3390/ijms232415888>

Shameli, Afshin

1. ***Shameli, A.**, Dharmani-Khan, P., Auer, I., & Shabani-Rad, M.-T. (2023). Deep immunophenotypic analysis of the bone marrow progenitor cells in myelodysplastic syndromes. *Leukemia Research*, *134*, 107401–107401. <https://doi.org/10.1016/j.leukres.2023.107401>
2. ***Shameli, A.**, & Roshan, T. M. (2022). CD200 expression on Sezary cells: A valuable tool for flow cytometric assessment of peripheral blood T-cell neoplasms. *Cytometry Part B: Clinical Cytometry*, *102*(4), 303–311. <https://doi.org/10.1002/cyto.b.22080>
3. Srinivasan, C., **Shameli, A.**, Ritchie, B., & Adatia, A. (2023). Investigation of a synonymous mutation in *Btk* in a patient with agammaglobulinemia: A case report. *PubMed*, *11*(10). <https://doi.org/10.1002/iid3.1049>

Siadat, Farshid

1. Alaghehbandan, R., Agaimy, A., Ali, L., Alvarado-Cabrero, I., Amin, M. B., Boudova, L., Caliò, A., Comperat, E., Damjanov, I., Daum, O., Farcas, M., Gatalica, Z., Gill, A. J., Hartmann, A., Hayes, M. M., Hora, M., Kojima, F., Kristiansen, G., Kuroda, N., **Siadat, F.** & Atienza, I. (2022). A tribute to Prof. Ondrej Hes, MD, PhD (1968–2022). *Modern Pathology*, *35*(12), 2023–2024. <https://doi.org/10.1038/s41379-022-01159-8>
2. Alaghehbandan, R., **Siadat, F.**, & Trpkov, K. (2023). What's new in the WHO 2022 classification of kidney tumours? *Pathologica*, 1–15. <https://doi.org/10.32074/1591-951x-818>
3. Mansoor, M., **Siadat, F.**, & Trpkov, K. (2022). Low-grade oncocytic tumor (LOT) - a new renal entity ready for a prime time: An updated review. *PubMed*, *37*(5), 405–413. <https://doi.org/10.14670/hh-18-435>
4. Mansoor, M., Young-Speirs, M., Ren, B., Gotto, G., Merten, L., Sawhney, S., **Siadat, F.**, Acosta, A. M., Agaimy, A., & Trpkov, K. (2022). Extrarenal renal cell carcinoma arising in the kidney proximity but without an identifiable renal primary – an intriguing dilemma: report of three cases and review of the literature. *Histopathology*, *81*(5), 635–643. <https://doi.org/10.1111/his.14736>
5. Sangoi, A. R., Chan, E., Abdulfatah, E., Stohr, B. A., Nguyen, J., Trpkov, K., **Siadat, F.**, Hirsch, M., Falzarano, S., Udager, A. M., & Kunju, L. P. (2022). p53 null phenotype is a “positive result” in urothelial carcinoma in situ. *Modern Pathology*, *35*(9), 1287–1292. <https://doi.org/10.1038/s41379-022-01062-2>
6. Sangoi, A. R., Maclean, F., Mohanty, S., Hes, O., Daniel, R., Lal, P., Canete-Portillo, S., Magi-Galluzzi, C., Cornejo, K. M., Collins, K., Hwang, M., Falzarano, S. M., Feely, M. M., Dababneh, M., Harik, L., Tretiakova, M., Akgul, M., Manucha, V., Chan, E., **Siadat, F.** & Kao, C. (2022). Granulomas associated with renal neoplasms: A multi-institutional clinicopathological study of 111 cases. *Histopathology*, *80*(6), 922–927. <https://doi.org/10.1111/his.14633>
7. ***Siadat, F.**, Mansoor, M., Hes, O., & Trpkov, K. (2022). Kidney Tumors. *Surgical Pathology Clinics*, *15*(4), 713–728. <https://doi.org/10.1016/j.path.2022.07.006>
8. Zhou, M., Amin, A., Fine, S. W., Rao, P., **Siadat, F.**, & Shah, R. B. (2023). Should grade group 1 prostate cancer be reclassified as “non-cancer”? A pathology community perspective. *Urologic Oncology: Seminars and Original Investigations*, *41*(2), 62–64. <https://doi.org/10.1016/j.urolonc.2022.09.028>

Sidhu, Davinder

1. Bégin, P., Callum, J., Cook, R., Jamula, E., Liu, Y., Finzi, A., Bégin, P., Callum, J., Jamula, E., Cook, R., Heddle, N. M., Tinmouth, A., Zeller, M. P., Beaudoin-Bussièrès, G., Amorim, **Sidhu, D.**, L., Bazin, R., Loftsgard, K. C., Carl, R., Chassé, M., & Cushing, M. M. (2022). Reply to: Concerns about estimating relative risk of death associated with convalescent plasma for COVID-19. *Nature Medicine*, 28(1), 53–58. <https://doi.org/10.1038/s41591-021-01639-5>
2. Bégin, P., Callum, J., Jamula, E., Cook, R., Heddle, N. M., Tinmouth, A., Zeller, M. P., Beaudoin-Bussièrès, G., Amorim, L., Bazin, R., Loftsgard, K. C., Carl, R., **Sidhu, D.**, Chassé, M., Cushing, M. M., Daneman, N., Devine, D. V., Dumaresq, J., Fergusson, D. A., Gabe, C., & Glesby, M. J. (2022). Author Correction: Convalescent plasma for hospitalized patients with COVID-19: an open-label, randomized controlled trial. *Nature Medicine*, 28(1), 212–212. <https://doi.org/10.1038/s41591-021-01667-1>
3. Harmon, M., Riazi, K., Callum, J., Arnold, D. M., Barty, R., **Sidhu, D.**, Heddle, N. M., MacLeod, L., & Li, N. (2023). Immunoglobulin utilization in Canada: a comparative analysis of provincial guidelines and a scoping review of the literature. *Allergy, Asthma & Clinical Immunology*, 19(1). <https://doi.org/10.1186/s13223-023-00841-z>
4. Li, N., Riazi, K., Arnold, D., **Sidhu, D.**, Barty, R., Heddle, N., Callum, J., & Down, D. (2023). P-TS-10 | An Unsupervised Learning Approach to Identify Immunoglobulin Utilization Patterns Using Electronic Health Records. *Transfusion*, 63(S5). https://doi.org/10.1111/trf.347_17554
5. Riazi, K., Ly, M., Barty, R., Callum, J., Arnold, D. M., Heddle, N. M., Down, D. G., **Sidhu, D.**, & Li, N. (2023). An unsupervised learning approach to identify immunoglobulin utilization patterns using electronic health records. *Transfusion*, 63(12), 2234–2247. <https://doi.org/10.1111/trf.17585>

Simpson, Roderick

1. Baněčková, M., Thompson, L. D. R., Hyrcza, M. D., Vaněček, T., Agaimy, A., Laco, J., **Simpson, R. H. W.**, Di Palma, S., Stevens, T. M., Brcic, L., Etebarian, A., Dimnik, K., Majewska, H., Stárek, I., O'Regan, E., Salviato, T., Helliwell, T., Horáková, M., Biernat, W., & Onyuma, T. (2023). Salivary Gland Secretory Carcinoma. *The American Journal of Surgical Pathology*, 47(6), 661–677. <https://doi.org/10.1097/pas.0000000000002043>
2. Chiesa-Estomba, C. M., Thompson, L., Agaimy, A., Zidar, N., **Simpson, R.**, Franchi, A., Rodrigo, J. P., Mäkitie, A. A., Almangush, A., Leivo, I., & Ferlito, A. (2023). Predictive value of tumor budding in head and neck squamous cell carcinoma: an update. *Virchows Archiv*, 483(4), 441–449. <https://doi.org/10.1007/s00428-023-03630-6>
3. Hellquist, H., Agaimy, A., Stenman, G., Franchi, A., Nadal, A., Skalova, A., Leivo, I., Zidar, N., **Simpson, R. H. W.**, Slootweg, P. J., Hernandez-Prera, J. C., & Ferlito, A. (2022). Development of head and neck pathology in Europe. *Virchows Archiv*, 480(5), 951–965. <https://doi.org/10.1007/s00428-022-03275-x>
4. Rooper, L. M., Agaimy, A., Gagan, J., **Simpson, R. H. W.**, Thompson, L. D. R., Trzcinska, A. M., Ud Din, N., & Bishop, J. A. (2022). Comprehensive Molecular Profiling of Sinonasal Teratocarcinoma Highlights Recurrent SMARCA4 Inactivation and CTNNB1 Mutations. *The American Journal of Surgical Pathology*, 47(2), 224–233. <https://doi.org/10.1097/pas.0000000000001976>
5. Skalova, A., Leivo, I., Hellquist, H., **Simpson, R. H. W.**, Vander Poorten, V., Willems, S. M., Mosaieby, E., Slouka, D., & Ferlito, A. (2022). Clear Cell Neoplasms of Salivary Glands: A Diagnostic Challenge. *Advances in Anatomic Pathology*, 29(4), 217–226. <https://doi.org/10.1097/pap.0000000000000339>

Teman, Carolin

1. Wu, G., Cheligeer, C., Brisson, A.-M., Quan, M. L., Cheung, W. Y., Brenner, D., Lupichuk, S., **Teman, C.**, Robert Barkev Basmadjian, Popwich, B., & Xu, Y. (2022). A New Method of Identifying Pathologic Complete Response After Neoadjuvant Chemotherapy for Breast Cancer Patients Using a Population-Based Electronic Medical Record System. *Annals of Surgical Oncology*, 30(4), 2095–2103. <https://doi.org/10.1245/s10434-022-12955-6>
2. Wu, G., Cheligeer, C., Brisson, A.-M., Quan, M. L., Cheung, W. Y., Brenner, D., Lupichuk, S., **Teman, C.**, Robert Barkev Basmadjian, Popwich, B., & Xu, Y. (2023). ASO Visual Abstract: A New Method of Identifying Pathologic Complete Response Following Neoadjuvant Chemotherapy for Breast Cancer Patients Using a Population-Based Electronic Medical Record System. *Annals of Surgical Oncology*, 30(4), 2106–2107. <https://doi.org/10.1245/s10434-022-13023-9>

Tordon, Bryan

1. Khandelwal, A., Vandermeulen, H., **Tordon, B.**, & Pavenski, K. (2022). Applying principles of patient blood management during COVID-19 pandemic: a literature review. *Annals of Blood*, 0. <https://doi.org/10.21037/aob-22-1>
2. ***Tordon, B.**, Drews, S. J., Flahr, F., Bennett, K., Gaziano, T., Anderson, D., Nahirniak, S., Gerges, H., Tyrrell, G. J., Mah, J., Ndao, M., Bigham, M., & Seftel, M. (2023). Canadian Blood Services traceback investigation of a suspected case of transfusion-transmitted malaria. *Transfusion*, 63(10), 2001–2006. <https://doi.org/10.1111/trf.17549>
3. ***Tordon, B.**, Montemayor, C., Clarke, G., O'Brien, S. F., & Goldman, M. (2023). Use of selective phenotyping and genotyping to identify rare blood donors in Canada. *Vox Sanguinis*, 118(5), 398–401. <https://doi.org/10.1111/vox.13419>
4. Yang, J., Yang, L., **Tordon, B.**, Bucher, O., Nugent, Z., Landego, I., Bourrier, N., Uminski, K., Brown, K., Squires, M., Marshall, A. J., Katyal, S., Mahmud, S., Decker, K., Geirnaert, M., Dawe, D. E., Gibson, S. B., Johnston, J. B., & Banerji, V. (2023). Clinical Outcomes in a Large Canadian Centralized CLL Clinic Based on Treatment and Molecular Factors over a Decade. *Current Oncology*, 30(7), 6411–6431. <https://doi.org/10.3390/curroncol30070472>

Trpkov, Kiril

1. ***Trpkov, K.**, & Zhou, M. (2022). Dedication. *Surgical Pathology Clinics*, 15(4), xiii–xiii. <https://doi.org/10.1016/j.path.2022.09.002>
2. Acosta, A. M., Gordetsky, J. B., Collins, K., Osunkoya, A. O., Sangoi, A. R., Miyamoto, H., Kao, C.-S., **Trpkov, K.**, Leenders, V., Wobker, S. E., Maclean, F., Lal, P., Daniel, R. E., Brimo, F., Wasco, M., Hirsch, M. S., Baniak, N., Diaz-Perez, J. A., Cornejo, K. M., & Choy, B. (2022). Clinicopathologic Spectrum of Secondary Solid Tumors of the Prostate of Nonurothelial Origin: Multi-institutional Evaluation of 85 Cases. *The American Journal of Surgical Pathology*, 46(9), 1269–1276. <https://doi.org/10.1097/pas.0000000000001907>
3. Acosta, A. M., Sangoi, A. R., Maclean, F., **Trpkov, K.**, Osunkoya, A. O., Collins, K., Miyamoto, H., Hirsch, M. S., Chan, E., Tretiakova, M., Mohanty, S. K., Kaushal, S., Cornejo, K. M., Aron, M., Quiroga-Garza, G., Arora, K., Nguyen, J. K., Williamson, S. R., Epstein, J. I., & Matoso, A. (2022). Prostatic malakoplakia: clinicopathological assessment of a multi-institutional series of 49 patients. *Histopathology*, 81(4), 520–528. <https://doi.org/10.1111/his.14729>
4. Al-Obaidy, K. I., Saleeb, R., **Trpkov, K.**, Williamson, S. R., Sangoi, A. R., Nassiri, M., Hes, O., Montironi, R., Cimadamore, A., Acosta, A., Alruwaili, Z., Alkashash, A., Hassan, O., Gupta, N., Osunkoya, A. O., Sen, J. D., Baldrige, L. A., Sakr, W., Idrees, M. T., & Eble, J. N. (2022). Recurrent KRAS mutations are early events in the development of papillary renal neoplasm with reverse polarity. *Modern Pathology*, 35(9), 1279–1286. <https://doi.org/10.1038/s41379-022-01018-6>

5. Alaghehbandan, R., Agaimy, A., Ali, L., Alvarado-Cabrero, I., Amin, M. B., Boudova, L., Caliò, A., Comperat, E., Damjanov, I., Daum, O., Farcas, M., Gatalica, Z., Gill, A. J., Hartmann, A., Hayes, M. M., Hora, M., Kojima, F., Kristiansen, G., Kuroda, N., **Trpkov, K.** & Atienza, I. (2022). A tribute to Prof. Ondrej Hes, MD, PhD (1968–2022). *Modern Pathology*, 35(12), 2023–2024. <https://doi.org/10.1038/s41379-022-01159-8>
6. *Alaghehbandan, R., Siadat, F., & **Trpkov, K.** (2023). What's new in the WHO 2022 classification of kidney tumours? *Pathologica*, 1–15. <https://doi.org/10.32074/1591-951x-818>
7. Anderson, W. J., Mariño-Enríquez, A., **Trpkov, K.**, Hornick, J. L., Nucci, M. R., Dickson, B. C., & Fletcher, C. D. M. (2023). Expanding the Clinicopathologic and Molecular Spectrum of Lipoblastoma-Like Tumor in a Series of 28 Cases. *Modern Pathology*, 36(10), 100252–100252. <https://doi.org/10.1016/j.modpat.2023.100252>
8. Castillo, V. F., Masoomian, M., **Trpkov, K.**, Downes, M., Brimo, F., van, Yousef, G. M., Zakhary, A., Rotondo, F., Saad, G., Nguyen, V., Kidanewold, W., Streutker, C., Rowsell, C., Hamdani, M., & Saleeb, R. M. (2023). ABCC2 brush-border expression predicts outcome in papillary renal cell carcinoma: a multi-institutional study of 254 cases. *Histopathology*, 83(6), 949–958. <https://doi.org/10.1111/his.15042>
9. Collins, K., Sholl, L. M., Siegmund, S., Dickson, B. C., Colecchia, M., Michalová, K., Hwang, M., Ulbright, T. M., Kao, C., Leenders, van, Mehta, V., **Trpkov, K.**, Yilmaz, A., Cimadamore, A., Matoso, A., Epstein, J. I., Maclean, F., Comperat, E., Anderson, W. J., & Fletcher, C. D. M. (2022). Myoid gonadal stromal tumours are characterised by recurrent chromosome-level copy number gains: molecular assessment of a multi-institutional series. *Histopathology*, 82(3), 431–438. <https://doi.org/10.1111/his.14825>
10. *Hes, O., & **Trpkov, K.** (2022). Do we need an updated classification of oncocytic renal tumors? Emergence of low-grade oncocytic tumor (LOT) and eosinophilic vacuolated tumor (EVT) as novel renal entities. *Modern Pathology*, 35(9), 1140–1150. <https://doi.org/10.1038/s41379-022-01057-z>
11. *Hora, M., Albiges, L., Bedke, J., Campi, R., Capitanio, U., Giles, R. H., Ljungberg, B., Marconi, L., Klatte, T., Volpe, A., Abu-Ghanem, Y., Dabestani, S., Fernández-Pello, S., Hofmann, F., Kuusk, T., Tahbaz, R., Powles, T., Bex, A., & **Trpkov, K.** (2023). European Association of Urology Guidelines Panel on Renal Cell Carcinoma Update on the New World Health Organization Classification of Kidney Tumours 2022: The Urologist's Point of View. *European Urology*, 83(2), 97–100. <https://doi.org/10.1016/j.eururo.2022.11.001>
12. Jacox, N., Yao, H. H.-I., Baverstock, R., **Trpkov, K.**, & Carlson, K. (2023). Periurethral and Anterior Vaginal Wall Masses: Etiology, Presentation, and Treatment Outcomes. *Obstetrical & Gynecological Survey*, 78(3), 150–152. <https://doi.org/10.1097/01.ogx.0000923064.68826.bd>
13. *Kapur, P., Brugarolas, J., & **Trpkov, K.** (2023). Recent Advances in Renal Tumors with TSC/mTOR Pathway Abnormalities in Patients with Tuberous Sclerosis Complex and in the Sporadic Setting. *Cancers*, 15(16), 4043. <https://doi.org/10.3390/cancers15164043>
14. ***Kiril Trpkov.** (2023). TSC/MTOR-associated Eosinophilic Renal Tumors Exhibit a Heterogeneous Clinicopathologic Spectrum: Several Distinct Entities or a Tumor Family? *The American Journal of Surgical Pathology*, 47(4), 518–519. <https://doi.org/10.1097/pas.0000000000002020>
15. Klontzas, M. E., Koltsakis, E., Kalarakis, G., **Trpkov, K.**, Papatomas, T., Karantanas, A. H., & Tzortzakakis, A. (2023). Machine Learning Integrating 99mTc Sestamibi SPECT/CT and Radiomics Data Achieves Optimal Characterization of Renal Oncocytic Tumors. *Cancers*, 15(14), 3553–3553. <https://doi.org/10.3390/cancers15143553>
16. Klontzas, M. E., Koltsakis, E., Kalarakis, G., **Trpkov, K.**, Papatomas, T., Sun, N., Walch, A., Karantanas, A. H., & Tzortzakakis, A. (2023). A pilot radiometabolomics integration study for the characterization of renal oncocytic neoplasia. *Scientific Reports*, 13(1), 12594. <https://doi.org/10.1038/s41598-023-39809-9>

17. Mannan, R., Wang, X., Bawa, P. S., Chugh, S., Chinnaiyan, A. K., Rangaswamy, R., Zhang, Y., Cao, X., Smith, S. C., **Trpkov, K.**, Williamson, S., Sangoi, A. R., Mohanty, S., McKenney, J. K., Gupta, S., Magi-Galluzzi, C., Argani, P., Osunkoya, A. O., Chinnaiyan, A. M., & Dhanasekaran, S. M. (2023). Characterization of protein S-(2-succino)-cysteine (2SC) succination as a biomarker for fumarate hydratase-deficient renal cell carcinoma. *Human Pathology*, *134*, 102–113. <https://doi.org/10.1016/j.humpath.2022.12.013>
18. Mansoor, A., Akhter, A., Shabani-Rad, M., Deschenes, J., Yilmaz, A., **Trpkov, K.**, & Stewart, D. (2023). Primary testicular lymphoma demonstrates overexpression of the Wilms tumor 1 gene and different mRNA and miRNA expression profiles compared to nodal diffuse large B-cell lymphoma. *Hematological Oncology*, *41*(5), 828–837. <https://doi.org/10.1002/hon.3190>
19. *Mansoor, M., Siadat, F., & **Trpkov, K.** (2022). Low-grade oncocytic tumor (LOT) - a new renal entity ready for a prime time: An updated review. *PubMed*, *37*(5), 405–413. <https://doi.org/10.14670/hh-18-435>
20. *Mansoor, M., Young-Speirs, M., Ren, B., Gotto, G., Merten, L., Sawhney, S., Siadat, F., Acosta, A. M., Agaimy, A., & **Trpkov, K.** (2022). Extrarenal renal cell carcinoma arising in the kidney proximity but without an identifiable renal primary – an intriguing dilemma: report of three cases and review of the literature. *Histopathology*, *81*(5), 635–643. <https://doi.org/10.1111/his.14736>
21. Mohanty, S. K., Lobo, A., Williamson, S. R., Shah, R. B., **Trpkov, K.**, Varma, M., Sirohi, D., Aron, M., Kandukari, S. R., Balzer, B. L., Luthringer, D. L., Ro, J., Osunkoya, A. O., Desai, S., Menon, S., Nigam, L. K., Sardana, R., Roy, P., Kaushal, S., & Midha, D. (2022). Reporting Trends, Practices, and Resource Utilization in Neuroendocrine Tumors of the Prostate Gland: A Survey among Thirty-Nine Genitourinary Pathologists. *International Journal of Surgical Pathology*, *31*(6), 993–1005. <https://doi.org/10.1177/10668969221116629>
22. Nova-Camacho, L. M., Acosta, A. M., **Trpkov, K.**, Sangoi, A. R., Pierre, A., Chou, A., Yilmaz, A., Morini, A., Rodrigues, Â., Fletcher, C. DM., Perez-Montiel, D., Maclean, F., Contreras, F., Francisco Javier Queipo, Gorka Muñiz Unamunzaga, Mesa, H., de Torres, I., Ruiz, I., Alvarado-Cabrero, I., & Lobo, J. (2023). Metastatic solid tumors to the testis: a clinicopathologic evaluation of 157 cases from an international collaboration. *Human Pathology*, *139*, 37–46. <https://doi.org/10.1016/j.humpath.2023.06.002>
23. Nova-Camacho, L. M., Collins, K., **Trpkov, K.**, Acosta, A. M., Sangoi, A. R., Akgul, M., Chou, A., Polonia, A., Rodrigues, Â., Yilmaz, A., Perez-Montiel, D., Maclean, F., Javier, F., Contreras, F., Wu, H. H., Alvarado-Cabrero, I., de Torres, I., Ruiz, I., Lobo, J., & Prendeville, S. (2023). Metastatic solid tumours to the penis: a clinicopathologic evaluation of 109 cases from an international collaboration. *Histopathology*, *83*(1), 31–39. <https://doi.org/10.1111/his.14927>
24. *Ricci, C., Ambrosi, F., Franceschini, T., Giunchi, F., Grillini, A., Franchini, E., Grillini, M., Schiavina, R., Massari, F., Mollica, V., Tateo, V., Federico Mineo Bianchi, Bianchi, L., Droghetti, M., Maloberti, T., Tallini, G., Colecchia, M., Andres Martin Acosta, Lobo, J., & **Trpkov, K.** (2023). Evaluation of an institutional series of low-grade oncocytic tumor (LOT) of the kidney and review of the mutational landscape of LOT. *Virchows Archiv*, *483*(5), 687–698. <https://doi.org/10.1007/s00428-023-03673-9>
25. Sangoi, A. R., Chan, E., Abdulfatah, E., Stohr, B. A., Nguyen, J., **Trpkov, K.**, Siadat, F., Hirsch, M., Falzarano, S., Udager, A. M., & Kunju, L. P. (2022). p53 null phenotype is a “positive result” in urothelial carcinoma in situ. *Modern Pathology*, *35*(9), 1287–1292. <https://doi.org/10.1038/s41379-022-01062-2>
26. *Siadat, F., Mansoor, M., Hes, O., & **Trpkov, K.** (2022). Kidney Tumors: New and Emerging Kidney Tumor Entities. *Surgical Pathology Clinics*, *15*(4), 713–728. <https://doi.org/10.1016/j.path.2022.07.006>
27. Siegmund, S. E., Sholl, L. M., Tsai, H. K., Yang, Y., Vasudevaraja, V., Tran, I., Snuderl, M., Fletcher, C. D. M., Cornejo, K. M., Idrees, M. T., Al-Obaidy, K. I., Collins, K., Gordetsky, J. B., Wobker, S. E.,

- Hirsch, M. S., **Trpkov, K.**, Yilmaz, A., Anderson, W. J., Quiroga-Garza, G., & Magi-Galluzzi, C. (2022). Clinicopathologic and molecular spectrum of testicular sex cord-stromal tumors not amenable to specific histopathologic subclassification. *Modern Pathology*, *35*(12), 1944–1954. <https://doi.org/10.1038/s41379-022-01155-y>
28. Tzortzakakis, A., Papatomas, T., Gustafsson, O., Gabrielson, S., **Trpkov, K.**, Ekström-Ehn, L., Arvanitis, A., Holstensson, M., Karlsson, M., Kokaraki, G., & Axelsson, R. (2022). ^{99m}Tc-Sestamibi SPECT/CT and histopathological features of oncocytic renal neoplasia. *Scandinavian Journal of Urology*, *56*(5-6), 375–382. <https://doi.org/10.1080/21681805.2022.2119273>
29. Williamson, S. R., Hes, O., **Trpkov, K.**, Aggarwal, A., Satapathy, A., Mishra, S., Sharma, S., Sangoi, A. R., Cheng, L., Akgul, M., Idrees, M. T., Levin, A., Sadasivan, S. M., San, P., Rogala, J., Compérat, E., Berney, D. M., Bulimbasic, S., McKenney, J. K., & Jha, S. (2022). Low-grade oncocytic tumour of the kidney is characterised by genetic alterations of *TSC1*, *TSC2*, *MTOR* or *PIK3CA* and consistent GATA3 positivity. *Histopathology*, *82*(2), 296–304. <https://doi.org/10.1111/his.14816>

Urbanski, Stefan

1. Almishri, W., Swain, L. A., D'Mello, C., Le, T. S., **Urbanski, S. J.**, & Nguyen, H. H. (2022). ADAM Metalloproteinase Domain 17 Regulates Cholestasis-Associated Liver Injury and Sickness Behavior Development in Mice. *Frontiers in Immunology*, *12*. <https://doi.org/10.3389/fimmu.2021.779119>
2. Scheidl, T. B., Wager, J. L., Baker, L. G., Brightwell, A. L., Melan, K. M., Larion, S., Sarr, O., Regnault, T., **Urbanski, S. J.**, & Thompson, J. (2023). High maternal adiposity during pregnancy programs an imbalance in the lipidome and predisposes to diet-induced hepatosteatosis in the offspring. *Bioscience Reports*, *43*(10). <https://doi.org/10.1042/bsr20231060>
3. Swain, L. A., Ambasta, A., Erika Prando Munhoz, Omodon, O., **Urbanski, S. J.**, & Nguyen, H. H. (2023). Acute severe hepatitis as a presenting symptom in clinically stable patients admitted with SARS-CoV-2 Omicron infection. *Hepatology Communications*, *7*(4). <https://doi.org/10.1097/hc9.000000000000115>

Venner, Allison

1. Bohn, M. K., Bailey, D., Balion, C., Cembrowski, G., Collier, C., De Guire, V., Higgins, V., Jung, B., Zahraa Mohammed Ali, Seccombe, D., Taher, J., Tsui, A. K. Y., **Venner, A.**, & Adeli, K. (2023). Reference Interval Harmonization: Harnessing the Power of Big Data Analytics to Derive Common Reference Intervals across Populations and Testing Platforms. *Clinical Chemistry*, *69*(9), 991–1008. <https://doi.org/10.1093/clinchem/hvad099>
2. M.A, N., Higgins, V., **Venner, A. A.**, Bailey, D., Beriault, D., Collier, C. M., & Adeli, K. (2022). Canadian Society of Clinical Chemists Harmonized Clinical Laboratory Lipid Reporting Recommendations on the Basis of the 2021 Canadian Cardiovascular Society Lipid Guidelines. *Canadian Journal of Cardiology*, *38*(8), 1180–1188. <https://doi.org/10.1016/j.cjca.2022.03.019>
3. Shaw, J. L. V., Arnoldo, S., Beach, L., Bouhtiauy, I., Brinc, D., Brun, M., Collier, C., Kostantin, E., Fung, A. W. S., Füzéry, A. K., Huang, Y., Kaur, S., Knauer, M., Labrecque, L., Leung, F., Shea, J. L., Thakur, V., Thorlacius, L., **Venner, A. A.**, & Yip, P. M. (2023). Establishing quality indicators for point of care glucose testing: recommendations from the Canadian Society for Clinical Chemists Point of Care Testing and Quality Indicators Special Interest Groups. *Clinical Chemistry and Laboratory Medicine*, *61*(7), 1280–1287. <https://doi.org/10.1515/cclm-2023-0147>
4. Stokes, W., Berenger, B. M., **Venner, A. A.**, Deslandes, V., & Julie. (2022). Point of care molecular and antigen detection tests for COVID-19: current status and future prospects. *Expert Review of Molecular Diagnostics*, *22*(8), 797–809. <https://doi.org/10.1080/14737159.2022.2122712>

5. Stokes, W., **Venner, A. A.**, Buss, E., Tipples, G., & Berenger, B. M. (2023a). Evaluation of the ID NOW among symptomatic individuals during the Omicron wave. *Journal of Medical Microbiology*, 72(2). <https://doi.org/10.1099/jmm.0.001669>
6. Stokes, W., **Venner, A. A.**, Buss, E., Tipples, G., & Berenger, B. M. (2023b). Prospective population-level validation of the Abbott ID NOW severe acute respiratory syndrome coronavirus 2 device implemented in multiple settings for testing asymptomatic and symptomatic individuals. *Clinical Microbiology and Infection*, 29(2), 247–252. <https://doi.org/10.1016/j.cmi.2022.08.025>
7. Wayne Lewis, C., Butorin, Y., de Koning, L., Paul, H. A., Gifford, J. L., **Venner, A. A.**, & Seiden-Long, I. (2023). Low partial pressure of oxygen causes significant and unrecognized under-recovery of glucose on blood gas analyzers. *Clinical Biochemistry*, 111, 60–65. <https://doi.org/10.1016/j.clinbiochem.2022.10.008>

Walker, Simon

1. Gamallat, Y., Bakker, A., Khosh Kish, E., Choudhry, M., **Walker, S.**, Aldakheel, S., Seyedi, S., Huang, K.-C., Ghosh, S., Gotto, G., & Bismar, T. A. (2022). The Association between Cyclin Dependent Kinase 2 Associated Protein 1 (CDK2AP1) and Molecular Subtypes of Lethal Prostate Cancer. *International Journal of Molecular Sciences*, 23(21), 13326. <https://doi.org/10.3390/ijms232113326>

Wang, Yinong

1. Hijazi, W., Jefferson, L., **Wang, Y.**, Howarth, A., Kanani, R., Shaw, J., Kent, W., & Miller, R. J. H. (2022). Eosinophilic Myocarditis Complicated by Massive Right Ventricular Thrombus. *Circulation: Cardiovascular Imaging*, 15(8). <https://doi.org/10.1161/circimaging.121.013873>

Wright, James (Jim)

1. ***Wright, J. R.** (2023). J.J.R. Macleod: Misunderstood, Misinterpreted, and Maligned. *Canadian Bulletin of Medical History*, 40(2), 430–465. <https://doi.org/10.3138/cjmh.598-072022>
2. Beers, C. A., Pond, G. R., **Wright, J. R.**, Tsakiridis, T., Okawara, G. S., & Swaminath, A. (2023). The impact of staging FDG-PET/CT on treatment for stage III NSCLC - an analysis of population-based data from Ontario, Canada. *Frontiers in Oncology*, 13. <https://doi.org/10.3389/fonc.2023.1210945>
3. de Koning, L., Crawford, S., Nohr, E., Chadha, R., Horn, C., **Wright, J. R.**, & Chan, E. S. (2022). Recurrence risk of villitis of unknown etiology: Analysis of a large retrospective cohort study, systematic review and meta-analysis. *Placenta*, 120, 32–39. <https://doi.org/10.1016/j.placenta.2022.02.006>
4. Flávia Kessler Borges, Devereaux, P. J., Cuerden, M., Sontrop, J. M., Bhandari, M., Guerra-Farfan, E., Patel, A., Sigamani, A., Umer, M., Neary, J., Tiboni, M., Tandon, V., **Wright, J. R.** Mmampapatla Thomas Ramokgopa, Sancheti, P., Lawendy, A.-R., Balaguer-Castro, M., Jenkinson, R., Ślęczka, P., Aamer Nabi Nur, & Wood, G. (2022). Accelerated Surgery Versus Standard Care in Hip Fracture (HIP ATTACK-1): A Kidney Substudy of a Randomized Clinical Trial. *American Journal of Kidney Diseases*, 80(5), 686–689. <https://doi.org/10.1053/j.ajkd.2022.01.431>
5. Goldberg, M., Parpia, S., Rakovitch, E., Chang, L., Bowen, J., Lukka, H., Perera, F., Fyles, A., **Wright, J.**, Sussman, J., & Whelan, T. (2023). Long-term outcomes and effects of hypofractionated radiotherapy in microinvasive breast cancer: Analysis from a randomized trial. *The Breast*, 68, 189–193. <https://doi.org/10.1016/j.breast.2023.02.005>
6. Gouran-Savadkoohi, M., Mesci, A., Pond, G. R., Swaminath, A., Quan, K., **Wright, J.**, & Tsakiridis, T. (2023). Contemporary real-world radiotherapy outcomes of unresected locally advanced

- non-small cell lung cancer. *Journal of Thoracic Disease*, 15(2), 423–433.
<https://doi.org/10.21037/jtd-22-925>
7. Lee, J., Nguyen, N., **Wright, J. R.**, Yeung, K.-K. D., Sagar, S. M., Kim, D.-H., Ostapiak, O., Doerwald-Munoz, L., & Whelan, T. J. (2023). A phase 2 study of stereotactic body radiation therapy for squamous cell carcinoma of the head and neck (SHINE): a single arm clinical trial protocol. *BMC Cancer*, 23(1). <https://doi.org/10.1186/s12885-023-10807-4>
 8. Sur, R., Pond, G., Falkson, C., Pan, M., **Wright, J.**, Bezjak, A., Dagnault, A., Yu, E., Almahmudi, M., Puksa, S., Gopaul, D., Tsakiridis, T., Swaminath, A., Ellis, P., & Whelan, T. (2023). BRACHY: A Randomized Trial to Evaluate Symptom Improvement in Advanced Non-Small Cell Lung Cancer Treated With External Beam Radiation With or Without High-Dose-Rate Intraluminal Brachytherapy. *International Journal of Radiation Oncology Biology Physics*, 116(3), 601–610. <https://doi.org/10.1016/j.ijrobp.2022.12.049>
 9. Swaminath, A., Wierzbicki, M., Parpia, S., Kundapur, V., Faria, S., Ahmed, N., Bujold, A., Hirmiz, K., Owen, T., Leong, N., Ramchandrar, K., Filion, E., Lau, H., Thompson, R., Yaremko, B., Gabos, Z., Mehiri, S., **Wright, J. R.**, Tsakiridis, T. K., & Cline, K. (2022). Lung SBRT credentialing in the Canadian OCOG-LUSTRE randomized trial. *Clinical and Translational Radiation Oncology*, 37, 145–152. <https://doi.org/10.1016/j.ctro.2022.10.002>
 10. Whelan, T. J., Smith, S., Parpia, S., Fyles, A., Bane, A., Liu, F., Rakovitch, E., Chang, L., Stevens, C., Bowen, J., Provencher, S., Théberge, V., Mulligan, A. M., Kos, Z., Akra, M., Voduc, K. D., Hijal, T., Dayes, I. S., Pond, G. R., & **Wright, J. R.** (2023). Omitting Radiotherapy after Breast-Conserving Surgery in Luminal A Breast Cancer. *The New England Journal of Medicine*, 389(7), 612–619. <https://doi.org/10.1056/nejmoa2302344>
 11. ***Wright, J. R.** (2022a). Broca’s legacy: fame not shame. *Brain*. <https://doi.org/10.1093/brain/awac131>
 12. ***Wright, J. R.** (2022b). Co-occurrence of Eosinophilic/T-Cell Chorionic Vasculitis and Acute Chorionitis, Acute Chorionic Vasculitis, and Funisitis in a Term Placenta. *Pediatric and Developmental Pathology*, 25(4), 485–488. <https://doi.org/10.1177/10935266221086249>
 13. ***Wright, J. R.** (2022c). Société Mutuelle d’Autopsie, American Anthropometric Society, and the Wilder Brain Collection. *Archives of Pathology & Laboratory Medicine*. <https://doi.org/10.5858/arpa.2021-0623-hp>
 14. ***Wright, J. R.** (2023). Maud Menten: Pioneering Pediatric-Perinatal Pathologist, Clinician-Scientist, and “the Most Wonderful Human Being in the World.” *Pediatric and Developmental Pathology*. <https://doi.org/10.1177/10935266231202934>
 15. ***Wright, J. R.**, Chan, S., Morgen, E. K., Maung, R. T. A., Brundler, M.-A., El Demellawy, D., Fraser, R. B., Kurek, K. C., Magee, F., Nizalick, E., Oligny, L. L., Somers, G. R., Stefanovici, C., & Terry, J. (2022). Workload Measurement in Subspecialty Placental Pathology in Canada. *Pediatric and Developmental Pathology*, 25(6), 604–610. <https://doi.org/10.1177/10935266221118150>
 16. ***Wright, J. R.**, & de Koning, L. (2023). Eosinophilic/T-cell Chorionic Vasculitis: Its Incidence Is Increasing but It Does Not Appear to Recur in Subsequent Pregnancies. *Pediatric and Developmental Pathology*, 26(3), 281–286. <https://doi.org/10.1177/10935266231156332>
 17. ***Wright, J. R.**, & Hibbitts, W. A. (2023). Dick van Velzen and the Burnside warehouse organ scandal in Nova Scotia, Canada. *Clinical Anatomy*, 37(1), 102–113. <https://doi.org/10.1002/ca.24115>
 18. ***Wright, J. R.**, Martin, B., Eggen, A., & Knight, C. G. (2023). Fetal bovine islets and the discovery of insulin. *General and Comparative Endocrinology*, 330, 114136–114136. <https://doi.org/10.1016/j.ygcen.2022.114136>
 19. ***Wright, J. R.**, Yu, W., & de Koning, L. (2022). Author(s) Reply - Vitelline Vessel Remnant – Derived Funisitis: Semantics, Severity, and Significance. *Pediatric and Developmental Pathology*, 25(5), 572–573. <https://doi.org/10.1177/10935266221113450>

20. *Yu, W., Thomas, M. A., Mills, L., & **Wright, J. R.** (2023). Prenatal Diagnosis of Isolated Right Ventricular Non-Compaction Cardiomyopathy with an *MYH7* Likely Pathogenic Variant. *Fetal and Pediatric Pathology*, 42(3), 464–471. <https://doi.org/10.1080/15513815.2022.2120785>

Yilmaz, Asli

1. Collins, K., Sholl, L. M., Siegmund, S., Dickson, B. C., Colecchia, M., Michalová, K., Hwang, M., Ulbright, T. M., Kao, C., Leenders, van, Mehta, V., Trpkov, K., **Yilmaz, A.**, Cimadamore, A., Matoso, A., Epstein, J. I., Maclean, F., Comperat, E., Anderson, W. J., & Fletcher, C. D. M. (2022). Myoid gonadal stromal tumours are characterised by recurrent chromosome-level copy number gains: molecular assessment of a multi-institutional series. *Histopathology*, 82(3), 431–438. <https://doi.org/10.1111/his.14825>
2. Mansoor, A., Akhter, A., Shabani-Rad, M., Deschenes, J., **Yilmaz, A.**, Trpkov, K., & Stewart, D. (2023). Primary testicular lymphoma demonstrates overexpression of the Wilms tumor 1 gene and different mRNA and miRNA expression profiles compared to nodal diffuse large B-cell lymphoma. *Hematological Oncology*, 41(5), 828–837. <https://doi.org/10.1002/hon.3190>
3. Nova-Camacho, L. M., Acosta, A. M., Trpkov, K., Sangoi, A. R., Pierre, A., Chou, A., **Yilmaz, A.**, Morini, A., Rodrigues, Â., Fletcher, C. DM., Perez-Montiel, D., Maclean, F., Contreras, F., Francisco Javier Queipo, Gorka Muñiz Unamunzaga, Mesa, H., de Torres, I., Ruiz, I., Alvarado-Cabrero, I., & Lobo, J. (2023). Metastatic solid tumors to the testis: a clinicopathologic evaluation of 157 cases from an international collaboration. *Human Pathology*, 139, 37–46. <https://doi.org/10.1016/j.humpath.2023.06.002>
4. Nova-Camacho, L. M., Collins, K., Trpkov, K., Acosta, A. M., Sangoi, A. R., Akgul, M., Chou, A., Polonia, A., Rodrigues, Â., **Yilmaz, A.**, Perez-Montiel, D., Maclean, F., Javier, F., Contreras, F., Wu, H. H., Alvarado-Cabrero, I., de Torres, I., Ruiz, I., Lobo, J., & Prendeville, S. (2023). Metastatic solid tumours to the penis: a clinicopathologic evaluation of 109 cases from an international collaboration. *Histopathology*, 83(1), 31–39. <https://doi.org/10.1111/his.14927>
5. Siegmund, S. E., Sholl, L. M., Tsai, H. K., Yang, Y., Vasudevaraja, V., Tran, I., Snuderl, M., Fletcher, C. D. M., Cornejo, K. M., Idrees, M. T., Al-Obaidy, K. I., Collins, K., Gordetsky, J. B., Wobker, S. E., Hirsch, M. S., Trpkov, K., **Yilmaz, A.**, Anderson, W. J., Quiroga-Garza, G., & Magi-Galluzzi, C. (2022). Clinicopathologic and molecular spectrum of testicular sex cord-stromal tumors not amenable to specific histopathologic subclassification. *Modern Pathology*, 35(12), 1944–1954. <https://doi.org/10.1038/s41379-022-01155-y>

Yu, Weiming

1. Chan, S. E., Suchet, I., **Yu, W.**, Somerset, D., Soliman, N., Kuret, V., Chadha, R. (2023) Absence of Ductus Venosus: A Comparison of 2 Distinctive Fetal Autopsy Cases and Embryologic Perspectives. *Pediatric Developmental Pathology*. <https://doi.org/10.1177/10935266231211760>
2. Chan, E. S., de Koning, L., **Yu, W.**, & Chadha, R. (2023). C4d Staining Is Present in Normal Placentas From Pregnancies Prior to Pregnancy Loss Associated With Chronic Histiocytic Intervillositis and Is Reduced by Immunomodulatory Therapy in Subsequent Pregnancies. *Pediatric and Developmental Pathology*, 26(4), 374–387. <https://doi.org/10.1177/10935266231176682>
3. Chan, E. S., Haider, S., Subramanian, S., **Yu, W.**, Nohr, E. W., & de Koning, L. (2022). Congenital cytomegalovirus infection is associated with congenital rickets: a retrospective autopsy cohort

study. *Archives of Disease in Childhood: Fetal & Neonatal*, 108(4), 435–436.
<https://doi.org/10.1136/archdischild-2022-324760>

4. Fraleigh, R., Wei, X.-C., **Yu, W.**, & Miettunen, P. M. (2023). Chronic recurrent multifocal osteomyelitis with a comprehensive approach to differential diagnosis of paediatric skull pain. *Case Reports*, 16(1), e252471–e252471. <https://doi.org/10.1136/bcr-2022-252471>
5. *Sarnat, H. B., Chan, E. S., Ng, D., & **Yu, W.** (2023). Maturation of metastases in peripheral neuroblastic tumors (neuroblastoma) of children. *Journal of Neuropathology and Experimental Neurology*, 82(10), 853–864. <https://doi.org/10.1093/jnen/nlad068>
6. *Sarnat, H. B., & **Yu, W.** (2022). Ganglion cell maturation in peripheral neuroblastic tumours of children. *Clinical Neuropathology*, 41(05), 101–113. <https://doi.org/10.5414/np301450>
7. Wright, J. R., **Yu, W.**, & de Koning, L. (2022). Author(s) Reply - Vitelline Vessel Remnant – Derived Funisitis: Semantics, Severity, and Significance. *Pediatric and Developmental Pathology*, 25(5), 572–573. <https://doi.org/10.1177/10935266221113450>
8. ***Yu, W.**, Thomas, M. A., Mills, L., & Wright, J. R. (2023). Prenatal Diagnosis of Isolated Right Ventricular Non-Compaction Cardiomyopathy with an *MYH7* Likely Pathogenic Variant. *Fetal and Pediatric Pathology*, 42(3), 464–471. <https://doi.org/10.1080/15513815.2022.2120785>

Zargham, Ramin

1. Hussain, M., Abbott, M., **Zargham, R.**, Pabani, A., & Khan, O. F. (2022). Evolution of an invasive ductal carcinoma to a small cell carcinoma of the breast. *Medicine*, 101(2), e28433. <https://doi.org/10.1097/md.00000000000028433>

Zhang, Kunyan

1. *De Filippis, I., & **Zhang, K.** (2022). “Staphylococci” Molecular typing in bacterial infections. Volume II (pp. 169–192). Springer.